

Archives of Agronomy and Soil Science



ISSN: 0365-0340 (Print) 1476-3567 (Online) Journal homepage: https://www.tandfonline.com/loi/gags20

Identifying optimum rates of fertilizer nitrogen application to maximize economic return and minimize nitrous oxide emission from rice-wheat systems in the Indo-Gangetic Plains of India

Tek B. Sapkota, Love K. Singh, Arvind K. Yadav, Arun Khatri-Chhetri, Hanuman S. Jat, Parbodh C. Sharma, Mangi L. Jat & Clare M. Stirling

To cite this article: Tek B. Sapkota, Love K. Singh, Arvind K. Yadav, Arun Khatri-Chhetri, Hanuman S. Jat, Parbodh C. Sharma, Mangi L. Jat & Clare M. Stirling (2020): Identifying optimum rates of fertilizer nitrogen application to maximize economic return and minimize nitrous oxide emission from rice–wheat systems in the Indo-Gangetic Plains of India, Archives of Agronomy and Soil Science, DOI: <u>10.1080/03650340.2019.1708332</u>

To link to this article: https://doi.org/10.1080/03650340.2019.1708332

9	© 2020 International Maize and Wheat Improvement Center (CIMMYT). Published by Informa UK Limited, trading as Taylor & Francis Group	+	View supplementary material 🕼
	Published online: 06 Jan 2020.		Submit your article to this journal 🕝
111	Article views: 1656	Q	View related articles 🖸
CrossMark	View Crossmark data 🗗		



OPEN ACCESS Check for updates

Identifying optimum rates of fertilizer nitrogen application to maximize economic return and minimize nitrous oxide emission from rice-wheat systems in the Indo-Gangetic Plains of India

Tek B. Sapkota ^[b]^a, Love K. Singh^b, Arvind K. Yadav^c, Arun Khatri-Chhetri^d, Hanuman S. Jat^{e,f}, Parbodh C. Sharma^f, Mangi L. Jat⁹ and Clare M. Stirling^h

^aSustainable Intensification Program, International Maize and Wheat Improvement Centre (CIMMYT), El Batan, Mexico; ^bBorlaug Institute for South Asia (BISA)/CIMMYT, Ludhiana, India; ^cDepartment of Agronomy, Sri Karan Narendra Agricultural University, Jobner, India; ^dCGIAR Research Program on Climate Change, Agriculture and Food Security (CCAFS), Borlaug Institute for South Asia (BISA)/CIMMYT, New Delhi, India; ^eSustainable Intensification Program, International Maize and Wheat Improvement Centre (CIMMYT), CSSRI, Karnal, India; ^fICAR-Central Soil Salinity Research Institute (CSSRI), Karnal, India; ^gSustainable Intensification Program, International Maize and Wheat Improvement Centre (CIMMYT), New Delhi, India; ^hSustainable Intensification Program, International Maize and Wheat Improvement Centre (CIMMYT), World Agroforestry Centre (ICRAF), Nairobi, Kenya

ABSTRACT

Rice-wheat (RW) cropping system in India is a major source of N₂O emissions. In such system, defining N rates that deliver minimal N₂O emissions and economically optimum yield would benefit both food production and the environment. We measured yield and N2O fluxes from RW systems in Northwest IGP under two tillage systems and five N rates (0, 75, 150, 225 and 300 kg N ha⁻¹) for both rice and wheat using static chamber method. Seasonal pattern of N₂O emission was mainly influenced by fertilizer and water application events with no significant effect of tillage systems. Mean annual N₂O emission from RW system was 1.49 kg N ha⁻¹ in N75 plot and 2.97–3.04 in the plots receiving \geq 150 kg N ha⁻¹. On average, the yield-scaled N₂O emissions of rice and wheat were 0.25 and 0.52 kg N₂O–N mg⁻¹, respectively. Our finding suggests that N rates between 120–200 kg N ha⁻¹ in rice and 50–185 kg ha⁻¹ in wheat provide the most economical returns and application rates beyond these ranges would be both economically and environmentally unsustainable. Within the range of N rate studied, fertilizer-induced N₂O-EF for rice and wheat were 0.41% and 0.79%, respectively.

ARTICLE HISTORY

Received 29 October 2018 Accepted 19 December 2019

KEYWORDS

Nitrous oxide; emission factor; greenhouse gas; rice-wheat system

Introduction

Nitrous oxide (N₂O) plays important roles in influencing stratospheric chemistry and regional and global climate change (IPCC 2013). Although present in small quantities, N₂O has a global warming potential approximately 265 time greater than that of CO₂ (Myhre et al. 2013) over a 100 years' time horizon. N₂O is also responsible for depleting stratospheric ozone (Ravishankara et al. 2009). Therefore, developing and implementing methods to reduce N₂O emissions from agricultural croplands is important. Agricultural soils account for approximately 60% global anthropogenic N₂O emission (Foley et al. 2011), N fertilization in croplands being one of the major sources (Cole et al. 1997). Addition of N fertilizer (both synthetic and organic) to agricultural soils leads to N₂O emissions,

CONTACT Tek B. Sapkota Stapkota@cgiar.org Dinternational Maize and Wheat Improvement Centre (CIMMYT), NASC complex, New Delhi 110012, India

Supplemental data for this article can be accessed here.

© 2020 International Maize and Wheat Improvement Center (CIMMYT). Published by Informa UK Limited, trading as Taylor & Francis Group. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives License (http:// creativecommons.org/licenses/by-nc-nd/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited, and is not altered, transformed, or built upon in any way.

2 👄 T. B. SAPKOTA ET AL.

predominantly by the microbial processes of nitrification (oxidation of ammonium to nitrate) and denitrification (reduction of nitrate to N_2 with N_2O as intermediary product). Numerous studies have shown that N_2O emissions from agricultural soils are directly related to N application rates (Bouwman et al. 2002; Halvorson et al. 2011; Hoben et al. 2011). Therefore, farmers in many regions of the world are being incentivized through polices and supply chains to improve nutrient use efficiency (NUE) through better in-field management to protect water resources and minimize GHG emissions whilst sustaining food production.

The rice-wheat (RW) system covering 16 million ha of productive land in Asia (Pathak et al. 2002) is important for the food security of millions of people (Ladha et al. 2009). This production system is also a major consumer of fertilizer N in the region. The majority of rice and wheat farmers in the IGP and elsewhere in South Asia apply fertilizer N following blanket recommendations based upon crop response data averaged over large geographic areas. Given the wide spatial variability in indigenous nutrient supplying capacity of soils in different agro-ecologies, crop fertilization following such blanket recommendation results in under-fertilization in some cases and over-fertilization in others. Diagnostic surveys in the IGP revealed that farmers often apply greater than recommended rates of fertilizer N and P, but overlook the need to balance this with sufficient application of potassium and other secondary and micro-nutrients (Singh et al. 2005). Because the government of India (Gol) provides more subsidy on nitrogenous fertilizer than other nutrients, farmers often apply fertilizer N in doses even higher than the blanket recommendations. Such imbalances and inadequate use of nutrients reduces the NUE and profitability and may increase environmental risks associated with loss of unutilized nutrient through emissions or leaching (Sapkota et al. 2014b). At present, total fertilizer-N consumption in India is about 17 million tonnes (Tewatia and Chanda 2017), about 70% of which is consumed in cereal production (Bijay-Singh 2017). The necessity to increase food production to feed an increasing population means that the consumption of fertilizer-N will continue to grow in the future and is projected to be around 24 million tonnes by 2030 (Tewatia and Chanda 2017). Therefore, fertilizer-induced N₂O emission from agriculture is expected to increase further in India.

Fertilizer N amount, source, timing and placement can all influence N₂O flux from agricultural soils (Snyder et al. 2007). Stehfest and Bouwman (2006), in an extensive review of published studies, concluded that soil N₂O flux is best predicted by N application rate, N source, crop type, soil pH, soil texture, climate and soil organic matter (SOM). Among these, total input of N to the soil is considered one of the strongest predictors of N_2O emissions and this together with the emission factors (EF) are widely used to estimate N₂O emissions (Shcherbak et al. 2014). Through review of peer-reviewed articles and modeling, Albanito et al. (2017) reported mean N_2O EF as 1.2% for tropics and subtropics, which is within the uncertainty range of IPCC Tier 1 EF, i.e. 0.3–3% (De Klein et al. 2006). This means, for every 100 kg of N input, 1.2 kg of N in the form of N₂O is estimated to be emitted directly from the soil. However, results from a growing number of field experiments indicate that the fraction of applied N emitted as direct N₂O increases with increasing rate of N application (McSwiney and Robertson 2005; Ma et al. 2010; Hoben et al. 2011; Shcherbak et al. 2014; Millar et al. 2018). This is likely due to competition for available N between plant uptake and N_2O producing microbes; such that when crop demand is met more N_2O is produced per unit of additional N (Ma et al. 2010). Given this biological threshold, use of single EF across the fertilizer rates may underestimate fertilizerinduced N_2O emission where fertilizer N application is higher than crop demand.

According to India's second national communication to the United Nationals Framework Convention on Climate Change (UNFCCC), fertilizer-induced field emission of N₂O in India was estimated to be 60 million tonnes CO₂e (GOI/MoEF 2012). A dearth of measurements under different crops, cropping systems and agro-ecology makes national and sub-national estimates highly uncertain. Analysis of yield and N input of the cropping systems in India during 1961–2009 demonstrates that crop N input (N rate) is increasing over time and corresponding NUE is decreasing (Bijay-Singh 2017). This indicates that substantial amounts of unused N under high N-input cropping systems may have been lost through direct and indirect emissions suggesting that fertilizer-induced N₂O EF may be much higher in these production systems. In the production systems such as RW system of IGP where fertilizer N is often applied at a rate exceeding crop demand. In such systems defining N rates where yields are maximized and environmental harm are minimized could benefit both food production, human health and the environment. We therefore conducted a field study to quantify yield and N₂O emissions under five N rates in RW systems of North-West IGP. The objectives of the study were to (i) identify a target range of N rates for RW system for optimal yields, profits and N₂O emissions that are compatible with sustainable intensification and (ii) develop fertilizer-induced N₂O EFs for intensive RW systems. This will potentially improve the national N₂O inventory as RW system is one of the most extensive and input-intensive cropping systems in India.

Materials and methods

Study site and experimental details

The study was conducted in the research farm of Central Soil Salinity Research Institute (CSSRI) of Indian Council of Agricultural Research (ICAR) located in Karnal district of Haryana, India (29°70' N, 76°96'E.; 250 m above sea level). The experimental site is a reclaimed alkali loam soil and the field was under a continuous RW system for over half a century before establishment of the experimental platform. The soil characteristics of the site are given in Supplementary Table 1. The climate of the location is semi-arid with average annual rainfall of 700 mm, 75–80% received during June–September. The lowest temperature is observed during January (daily minimum ranged from 0 to 4°C) and the highest temperature is observed during June (daily maximum ranges from 40 to 44°C). Monthly rainfall distribution along with minimum and maximum temperature during the experimental seasons is presented in Supplementary Figure S1.

We started the experiment from the wheat season in 2014–2015 (Nov–March), continued through the rice season in 2015 (July–October) to the wheat season in 2015–2016. We evaluated five N rates (i.e. 0, 75, 150, 225 and 300 kg N ha⁻¹) in both rice and wheat grown under two tillage systems (i.e. conventional tillage, CT and zero-tillage, ZT). Five N rates (sub-plot factors) were factorially combined with two tillage systems (main-plot factor) in split-plot design with three replications giving a total of 30 sub-plots of 10 by 10 m each.

Crop management

Crop management including land preparation, variety, seed rate, sowing/planting, as well as management of water, nutrient and pest for rice and wheat are summarized in Table 1. Tillage operations in CT-based rice involved three passes of dry tillage with harrow to the depth of 15 cm, two passes of cultivator in ponded water (puddling) followed by planking (levelling). CT-based wheat received two passes of harrow, one pass of cultivator followed by one planking. In CT systems, rice seedlings (variety CSR-30) were raised using a seed rate of 12 kg ha^{-1} and 30 days old seedlings were transplanted manually in random geometry with about 30 seedlings m^{-2} . In this system, wheat seeds were broadcasted after all tillage operations followed by planking. In the zero-tillage system, both rice and wheat were seeded without any preparatory tillage at a row spacing of 22.5 cm using ZT seed-cum-fertilizer drill having inclined-rotary-plate seed metering systems. In ZT system, the seed rate was 25 and 100 kg ha⁻¹ for rice and wheat, respectively. Seeding depth was maintained at \sim 2 cm and 5 cm in rice and wheat crop, respectively using a depth-control wheel of the planter. The field remained fallow after harvesting of the first year wheat until establishment of rice in rainy season. Crops were fertilized by manually broadcasting fertilizer as per the treatment. Both rice and wheat received 60 kg P_2O_5 and K_2O ha⁻¹ through single super phosphate (SSP) and muriate of potash (MoP), respectively. N fertilization was done as per the treatments using Urea. Full amount of P and K and 33% of N was applied as basal at the time of seeding/transplanting and the remaining two-third of N was applied in two equal splits at 20–25 and 40–45 days after seeding/transplanting in rice and wheat.

	C		Z	
Activity/Operations	Rice	Wheat	Rice	Wheat
Cultivar	CSR-30	HD-2967	CSR-30	HD-2967
Field preparation	Harrowing (×3), cultivators (×2) for	Harrowing (×2), cultivators (×1)	ZT	ZT
	puddling followed by planking.	followed by planking		
Date of sowing/transplanting	30 July 2015	13 November 2014	31 June 2015	13 November 2014
		18 November 2015		18 November 2015
Seed rate (kg ha $^{-1}$)	12	150	25	100
Method	Transplanting in random geometry	Broadcasted in random	Drill (line sowing)	Drill (line sowing)
		geometry		
Row spacing (cm)	Random	Random	20	20
Nutrient management	P_2O_5 and K_2O : 60 kg ha ⁻¹ through SSP	P_2O_5 and K_2O : 60 kg ha ⁻¹	P_2O_5 and K_2O : 60 kg ha ⁻¹ through SSP and MoP,	P_2O_5 and K_2O : 60 kg ha ⁻¹
	and MoP, respectively	through SSP and MoP,	respectively	through SSP and MoP,
	N: as per treatment	respectively	N: as per treatment	respectively
		N: as per treatment		N: as per treatment
Water management	Continuous flooding of 5-cm for one	Four irrigation at CRI, tillering,	First irrigation at sowing. Second irrigation one	Four irrigation at CRI, tillering,
	month, then irrigation applied at	flowering and grain filling	week after sowing. Subsequent irrigations at	flowering and grain filling
	appearance of hair-line crack		appearance of hair-line crack	
Weed management	Post-emergence: bispyribac sodium	Post-emergence: metsulfuron	Pre-sowing: glyphosate (1.25 g a.i. ha ⁻¹)	Pre-sowing: glyphosate (1.25 g a.
	$(25 \text{ mL a.i. ha}^{-1})$	methyl (20 g a.i. ha ^{_1})+	Pre-emergence: Pendimethalin (1.0 L a.i. ha ⁻¹)	i. ha ^{_1})
		clodinafop (400 g a.i. ha ^{–1})	Post-emergence: bispyribac sodium (25 g a.i.	Post-emergence: metsulfuron
			ha ⁻¹)	methyl (20 g a.i. ha ⁻¹)+
				clodinafop (400 g a.i. ha ^{_1})
Harvesting	26 October 2015	14 April 2015 09 April 2016	26 October 2015	14 April 2015 09 April 2016

Table 1. Details of tillage, crop establishment and residue management of rice and wheat under two tillage systems (main-plot factors).

During the rice cycle, CT plots were kept continuously flooded (5 cm standing water) for the initial one month and the subsequent irrigations were scheduled when soil matric potential reaches around -40 to -50 kPa determined through a visual inspection of hairline cracks on the soil surface (Gathala et al. 2011). In ZT plots, the first irrigation was applied immediately after sowing. The second irrigation was given one week after seeding and subsequent irrigations were applied as described under CT system. Irrespective of the tillage systems, wheat received four irrigations (6-7 cm each) at 20–25, 45–50, 75–80 and 95–100 days after sowing. In ZT plots, weeds prior to seeding of rice and wheat were killed by pre-plant application of glyphosate but no herbicides were applied in CT plots before seeding. Weed management after seeding/transplanting in rice and wheat were done by using appropriate pre- and post-emergence herbicide as required (Table 1).

Grain and biomass yield estimation

At maturity, crops were harvested manually from three 1-m² quadrats randomly selected within each plot for both rice and wheat. The plants within at least 0.5 m from the border were not considered for yield determination. The harvested crops were sun-dried and threshed to determine grain and straw yield. Grain and straw yield of rice and wheat were reported at 14% moisture content.

Collection of gas sample

Gas samples were collected using two-part static chambers as described by Sapkota et al. (2014a). The base of the chamber (43 cm i.d.) made up of the galvanized steel was semi-permanently installed (12 cm of the base inserted into the soil keeping 17.5 cm above soil surface) in the plots keeping 5–10 plants inside the chamber. It consisted of a circular channel to hold the upper part of the chamber. At the time of sampling, the upper part of the chamber was placed over the base of the chamber giving a total headspace volume of 105.41 L. The circular channel was filled with water and vents were sealed with adhesive to make the assembly airtight. The chamber top was equipped with a batterypowered fan to facilitate mixing of the gas in chamber headspace. Gas sampling was commenced one day before seeding in each crop. Thereafter, gas samples were collected once a week and for five consecutive days after every N fertilization events. Gas samples were collected through a septum fitted on the side of the chamber using a 50 mL polypropylene disposable syringe with three-way luer lock. The gas (50 mL) in the syringe was injected into the pre-evacuated and labelled 30 mL vials, which ensured higher pressure inside the vial to avoid contamination from ambient air. At each sampling, gas samples were collected four times within a total chamber deployment period of 30 min at 10-min interval. Sampling was performed between 9.00 and 11.30 am when soil surface temperature is believed to be equal to the daily average (Sapkota et al. 2014b). Chamber temperature was also recorded during gas sampling event using a thermometer fitted on the chamber top to be used for flux calculation. Depth of flood water when present and plant volumes inside the chamber were determined and subtracted from chamber volume to calculate effective chamber volume.

Gas analysis and calculations

Collected air samples were analyzed for N₂O using a Gas Chromatograph (GC) (model: Bruker 450) equipped with Electron Capture Detector (ECD) with the temperature settings of 300°C. Argon and 5% methane were used as carrier gases with a flow rate of 60 mL min⁻¹.

Concentrations of gases were calculated by comparing relative peak areas against the curves prepared from known concentrations (0.5, 1 and 10 ppm) of standard gases from Linde Engineering India Pvt. Ltd. To address the issue of GC drift, GC was calibrated periodically using N₂O standards of known concentration. Gas concentration at each sampling period was converted into mole of gas by using ideal gas law as below.

$$PV = nRT$$
(1)

where P = pressure, V = volume, n = number of moles of gas, R = the gas law constant and T = temperature.

The mole unit of gas was then converted into weight of gas considering the molecular weight of a particular gas. The daily N₂O emission rate was calculated from the linear increase (slope) in N₂O concentration over time. The increase in gas concentration over time was carefully monitored and only the data-points where *t*-test of the slope of the regression were significant (p = 0.05) and were included in cumulative seasonal emission calculations. The fluxes in between two sampling dates were estimated by linear interpolation. Seasonal cumulative N₂O emission was calculated from the sum of daily emission rates between planting to harvesting of the crop. The direct N₂O emission factor (EF%) induced by the N fertilizer was calculated as EF = $100 \times (E_F - E_0)/N$, where E_F (kg N ha⁻¹) is the seasonal cumulative N₂O flux for non-fertilized treatment, and N is the seasonal N fertilizer application rate (kg N ha⁻¹). To evaluate the environmental relevance of N₂O emissions under different N fertilizer management practices, the seasonal cumulative N₂O emission. Total annual grain yield, cumulative emission, emission intensity and EF were calculated using the data from 2015 to 2016 rice and wheat seasons.

Economic and statistical analyses

The amount of N applied may vary depending on whether the priority is to increase crop productivity, reduce emissions or maximize the rate of return from the use of N fertilizer. The best-case scenario would be to maximize yield and return whilst minimizing emissions from crop cultivation (multi-objective target). This study estimated productivity, emissions and return in relation to N rate. Grain production was converted to a money value based on the Minimum Support Price (MSP) for the study area, Karnal District of Haryana, India. The MSP is an agriculture product price set by Food Corporation of India (FCI) to purchase directly from the farmer (http://fci.gov.in). This rate is to safeguard the farmer to a minimum profit for the harvest, if the open market price is less than the cost of cultivation. The commission for Agricultural Cost and Prices (CACP), under the Ministry of Agriculture and Farmers welfare determines the MSP of particular agricultural products based on cost of cultivation and market prices of inputs and outputs in a particular location (https://cacp.dacnet.nic.in/).

Prices of paddy and wheat straws were collected from IndiaMART (https://dir.indiamart.com) for the study sites. Both grain and straw prices of paddy and wheat were also verified with the farmers from the study district. This study estimated an economically feasible range of N use in rice and wheat crops. The economically feasible range of N use indicates that return from additional N use in the crop cultivation is greater than the additional cost of N. When additional cost of N use is greater than additional return, application of N is no more economically suitable. Price of N fertilizer was collected from the Indian Farmers Fertilizer Cooperative (IFFCO: www.iffco.in) for the study sites which was further verified with the farmers from the study district. The incremental benefits of N use were estimated for different levels N rate for both paddy and wheat cultivation using Equation (2).

Incremental Benefit (IB) = Incremental Return
$$-$$
 Incremental Cost of N use (2)

A functional form of return to N use and emissions intensity was estimated using a nonlinear regression model. We used quadratic function to estimate the N rate response to economic return. Economic return is directly related to N rate response to crop output (yield), which is normally nonlinear (quadratic). Quadratic function indicates that N-response to yield initially increases and after reaching biological optimum the response starts decreasing. All data were analyzed following an Analysis of Variance (ANOVA) for split-plot design using the Costat Software (CoHort 2012). The difference between treatment means was compared using a LSD test at P < 0.05 (Gomez and Gomez 1984).

Results

Weather and environmental conditions

The growing season mean air temperature for 2014–2015 wheat, 2015 rice and 2015–2016 wheat were 16, 27 and 18°C, respectively (Supplementary Figure S1), which was similar to the long-term average of the location. Although minimum temperature was similar in both the years, maximum temperature particularly in April was higher in 2015–2016 wheat season than in 2014–2015 wheat season. The 2015 rice season received 370 mm rainfall, about 70% of the total annual precipitation (524 mm). The 2014–2015 wheat season received 247 mm rainfall which was about five times more than that received in 2015–2016 wheat season (56 mm) and 2.5 times more than the long-term average (110 mm).

Crop production

The effect of tillage system was not significant for grain as well as biomass yield in both rice and wheat in both the years (Table 2). Except for grain yield in 2014–2015, N rate resulted in significant difference in grain and above-ground biomass yield of rice and wheat in both the years (Table 2, Figure 1). There was no significant interaction between tillage and N rate. Therefore, we report grain and straw yields of each crop averaged over two tillage treatments. Figure 1 shows the grain and straw yield of rice and wheat receiving different N rates averaged over the two tillage systems. Grain yield and above ground biomass were always significantly lower in control (zero N) plots compared to plots where N fertilizer was applied. However, no significant effect of N was observed on grain, straw and aboveground biomass yield for N rate of 150 kg ha⁻¹ and above in both the crops in all years (Figure 1).

Seasonal trend of N₂O emission

Nitrogen fertilization rates clearly influenced the daily soil N₂O emissions during the wheat season in both years whereas no effect of tillage system was evident (Supplementary Figures S2 and S4). In

Source of variation	df	Grain yield (mg ha ⁻¹)	Total biomass yield (mg ha ⁻¹)	N ₂ O emission (kg ha ⁻¹)	N ₂ O emission intensity (kg CO2e/mg grain)	N ₂ O EF (%)
			Wheat 2014–2015			
Tillage (T)	1	ns	ns	ns	ns	ns
N rate (N)	4	ns	**	***	ns	ns
$T \times N$	4	ns	ns	ns	ns	ns
		Rice 2015				
Tillage (T)	1	ns	ns	ns	ns	*
N rate (N)	4	**	**	***	**	ns
$T \times N$	4	ns	ns	ns	ns	ns
		Wheat 2015-2016				
Tillage (T)	1	ns	ns	ns	ns	ns
N rate (N)	4	**	***	***	***	*
$T \times N$	4	ns	ns	ns	ns	ns
		Rice-Wheat system				
Tillage (T)	1	ns	Ns	ns	ns	ns
N rate (N)	4	**	***	***	***	**
$T \times N$	4	ns	ns	ns	ns	ns

Table 2. Significance of effects of tillage systems, N rate (N) and their interactions on grain yield, total biomass yield, cumulative N_2O emission, N_2O emission intensity and N_2O emission factor as resulting from analysis of variance (ANOVA).

*, ** and *** are significant at 0.05, 0.01 and 0.001 probability level, respectively. ns is non-significant.



Figure 1. Rice and wheat grain and straw yield at maturity under different N rates (averaged over two tillage systems and three replications) during the experimental period of 2014–2016. The straw yield is stacked over grain yield to show total biomass yield (grain and straw). In each panel, the bars bearing different lowercase letters are significantly different from each other at P < 0.05. Error bars (standard error of the means) for grain and straw yield are separately provided.

general, daily soil N₂O emissions from the unfertilized control plots were lower than the N fertilized plots. Among the N fertilized plots, daily N₂O emission were lower in plots fertilized with 75 kg N ha⁻¹ than those fertilizers at higher N rates, i.e. 150, 225 and 300 kg N ha⁻¹. The trend of N₂O emission was similar among the plots receiving 150–300 kg N ha⁻¹. During both wheat seasons daily N₂O emissions from N fertilized plots increased immediately after sowing and after each fertilization event reaching to a maximum of 14.67 mg N₂O-N m⁻² day⁻¹ in 2014–2015 wheat season and 24 mg N₂O-N m⁻² day⁻¹ in 2015–2016 wheat season. The peak N₂O emission was observed 2–3 days after each fertilization event (Supplementary Figures S2 and S4). Although the N₂O emission trend was similar under CT and ZT plots, slightly higher N₂O emissions were recorded under ZT than CT plots across all fertilization treatments after the last fertilizer event in both the years.

In the rice-growing season, both tillage systems as well as N fertilization rates affected the trend of N₂O emission. Irrespective of the tillage systems, daily emissions of N₂O were smaller in the control plot than the N fertilized plots. The plots receiving 75 kg N ha⁻¹ also recorded lower daily and seasonal N₂O emission than the plots receiving higher N rates, irrespective of the tillage system. The daily N₂O emission from N fertilized plots during rice growing season ranged from 0 to 12.78 mg N₂O–N m⁻² day⁻¹ (mean: 1.23 mg N₂O–N m⁻² day⁻¹) in CT plots and 0 to 22.39 mg N₂O–N m⁻² day⁻¹ (mean: 2.31 mg N₂O–N m⁻² day⁻¹) in ZT plot. In ZT plots, N₂O emissions were observed immediately after the first N fertilization whereas in CT plots no N₂O emission was observed after the first dose of N application (Supplementary Figure S3). The magnitude of daily N₂O emission in CT plots were smaller than those in ZT plots even after second dose of N fertilization. However, pronounced N₂O peaks appeared following third dose of N application, particularly from the plots with higher N rate, i. e. 225 and 300 kg N ha⁻¹. In ZT system, all the plots receiving 150 kg N ha⁻¹ or more recorded much higher daily N₂O emission than other plots after second and third fertilization event.

Cumulative N₂O emission

Nitrogen fertilization rates had significant effect on cumulative N₂O emissions in both rice and wheat growing seasons, whereas the effect of tillage systems was not significant. Similarly, tillage by N rate interaction effect was also not significant for soil N₂O emissions from both rice and wheat in both the years (Table 2). Therefore, only the mean effect of N rates averaged over tillage systems are presented in Table 3.

N rates (kg ha ⁻¹)	Cumulative N ₂ O emission (kg N ha ⁻¹)	Grain yield (mg ha ⁻¹)	Emission intensity (kg N ₂ O–N mg ⁻¹)	N ₂ O emission factor (%)	
0	0.88 ± 0.13 c	2.10 ± 0.36	0.54 ± 0.20 b		
75	1.48 ± 0.23 b	2.54 ± 0.45	0.72 ± 0.18 ab	0.81 ± 0.30	
150 225	2.22 ± 0.15 a 2.38 + 0.07 a	2.84 ± 0.32 3.76 ± 0.23	0.88 ± 0.15 a 0.65 + 0.05 ab	0.84 ± 0.12 0.67 ± 0.08	
300	2.40 ± 0.12 a	3.27 ± 0.51	0.88 ± 0.18 a	0.51 ± 0.05	
	2015 Rice				
0	$0.28\pm0.06~c$	2.05 ± 0.05	0.14 ± 0.03 b		
75	$0.62 \pm 0.07 \text{ b}$	3.51 ± 0.08	$0.18 \pm 0.02 \text{ b}$	0.45 ± 0.12	
150	1.15 ± 0.10 a	3.73 ± 0.05	0.31 ± 0.03 a	0.54 ± 0.09	
225	1.14 ± 0.13 a	3.72 ± 0.07	0.31 ± 0.04 a	0.38 ± 0.06	
300	1.16 ± 0.16 a	3.57 ± 0.06	0.33 ± 0.05 a	0.29 ± 0.06	
		no wheat			
0	$0.09 \pm 0.01 \text{ c}$	2.89 ± 0.22	$0.03 \pm 0.01 \text{ c}$		
75	0.87 ± 0.09 b	3.62 ± 0.41	0.25 ± 0.03 b	1.04 ± 0.11 a	
150	1.83 ± 0.18 a	4.33 ± 0.22	0.42 ± 0.04 a	1.09 ± 0.09 a	
225	1.89 ± 0.18 a	4.35 ± 0.14	0.44 ± 0.05 a	0.80 ± 0.08 ab	
300	1.87 ± 0.28 a	4.44 ± 0.10	0.43 ± 0.07 a	0.59 ± 0.09 b	
	2015–2016 Rice–Wheat System				
0	$0.37 \pm 0.07 \ c$	4.94 ± 0.17	0.07 ± 0.01 c		
75	1.49 ± 0.13 b	7.13 ± 0.41	0.21 ± 0.02 b	$0.75 \pm 0.08 \text{ ab}$	
150	2.97 ± 0.24 a	8.06 ± 0.24	0.37 ± 0.03 a	0.81 ± 0.08 a	
225	3.04 ± 0.22 a	8.07 ± 0.17	0.38 ± 0.03 a	0.59 ± 0.05 bc	
300	3.02 ± 0.44 a	8.00 ± 0.14	0.38 ± 0.06 a	0.44 ± 0.08 c	

Table 3. Seasonal and annual cumulative N₂O fluxes, grain yield, emission intensity and N₂O emission factor under different nitrogen rates from rice, wheat and rice–wheat. Within each crop and cropping system, means in columns bearing same lowercase letter are significantly different from each other based LSD test (p < 0.05). Values are presented as means \pm SEM (standard error of the mean).

10 🔄 T. B. SAPKOTA ET AL.

The seasonal cumulative N₂O emission was significantly lower in control plot followed by N75, in which cumulative N₂O emission was significantly higher than control but significantly lower than the plots receiving higher N rates (i.e. N150, N225 and N300) in both the years (Table 3). The seasonal cumulative N₂O emission during the rice growing season ranged from 0.097 to 0.52 kg N ha⁻¹ (mean value: 0.28 kg N ha⁻¹) for control plot. In the plots receiving N fertilization, average seasonal cumulative N₂O emission ranged from 0.62 kg N ha⁻¹ (in N75 plot) to 1.16 kg N ha⁻¹ (in N300 plot). Total cumulative N₂O emission from N75 plot in this season was significantly higher than the control but significantly lower than the plots receiving higher N rates. The plots receiving 150–300 kg N ha⁻¹ recorded the highest cumulative N₂O emission in this season (Table 3).

Seasonal cumulative emission from control plot in the wheat growing season ranged from 0.33 to 1.23 kg N ha⁻¹ (mean value: 0.88 kg h ha⁻¹) during first year and 0.05 to 0.13 kg N ha⁻¹ (mean value: 0.09 kg N ha⁻¹) during second year. In N75 plot, the cumulative N₂O emission for wheat season ranged from 0.99 to 2.33 kg N ha⁻¹ (mean value: 1.48 kg N ha⁻¹) during first year and 0.58 to 1.14 kg N ha⁻¹ (mean value: 0.87 kg N ha⁻¹) during second year (Table 3). N150, N225 and N300 plots had similar cumulative N₂O emission during both wheat growing season which ranged from 1.40 to 3.2 kg N ha⁻¹ (mean value: 2.10 kg N ha⁻¹).

Annual N₂O emission (emission from entire crop year consisting of 2015 rice and 2015–2016 wheat season) ranged from 0.17 to 0.66 kg N ha⁻¹ (mean value: 0.37 kg N ha⁻¹) for control which was significantly lower than the annual N₂O emission from other treatment (Table 3). Cumulative N₂O emission from annual RW rotation from N75 plot ranged from 1.08 to 1.88 kg N ha⁻¹ (mean value: 1.49 kg N ha⁻¹), which was significantly higher than that of control plot but significantly lower than the plots receiving higher N rates (Table 3). All plots receiving N rates higher than 150 kg ha⁻¹ (i.e. N150, N225 and N300) resulted in similar cumulative emissions.

Yield-scaled N₂O emission

Yield-scaled N₂O emission (emission intensity) of rice and wheat in the studied year is presented in Table 3. Irrespective of the crops and year, N₂O emission intensity was higher in all plots receiving N rate 150 kg ha⁻¹ or more. The plots receiving 75 kg N ha⁻¹ resulted into significantly lower N₂O emission intensity than other N fertilized plots but significantly higher than the control plot (Table 3). Emission intensity of rice ranged from 0.04–0.55 kg N₂O–N mg⁻¹ whereas that of wheat ranged from 0.25–1.61 kg N₂O–N mg⁻¹ during 2014–2015 growing season and 0.02–0.77 kg N₂O_N mg⁻¹ during 2015–2016 growing season. As in rice and wheat season, N₂O emission intensity of RW rotation was intermediate in N75, control plots having significantly lower and the plots receiving ≥150 kg N ha⁻¹ having significantly higher emission intensity (Table 3).

Soil N₂O emission factor (EF)

Irrespective of treatments, EF for rice ranged from 0.08% to 0.91% whereas that for wheat range from 0.14% to 2.12% in 2014–2015 growing season and 0.435 to 1.41% in 2015–2016 growing season. Annual N₂O EF from RW system ranged from 0.26% to 1.08% across different N rates. The effect of tillage system and tillage by N rates interaction were not significant for seasonal and annual N₂O EF (Table 2). Main effect of N rates on N₂O EF was significant during 2015–2016 wheat (seasonal) as well as RW (annual) rotation (Tables 2 and 3). Overall, N₂O EF was higher with smaller N rates and *vice-versa*.

Optimum N rate

A regression analysis was used to identify the relationship between marginal rate of return, N rate, and emissions intensity in both rice and wheat crops. This analysis estimated how N rate and emissions intensity behave with total economic return in both crops. Results of quadratic function showed that the N rate and emissions intensity in both rice and wheat crops were non-linearly



Figure 2. Observed and predicted grain yield, N₂O emission intensity and predicted marginal rate of return under different N rates in rice (upper panel) and wheat (lower panel), 2015–2016.

related to total economic returns (Supplementary Table 2). The total economic return increased up to certain level of N-rate and emissions intensity, then started to decrease with increase in N rate (Square N Rate) and emissions intensity (Square Emissions Intensity) in both crops. Figure 2 indicates an area of 'optimum' N rate for rice (upper panel) and wheat (lower panel) crops. The optimum N ranges from the point of diminishing marginal rate of return to the point where yield stop increasing. The optimum rate of N fertilizer application in rice ranges from 120 to 200 kg ha⁻¹. Similarly, the optimum rate of N fertilizer application in wheat ranges from 50 to 185 kg ha⁻¹.

Discussion

Seasonal N₂O emission trend

In both the crops, the fluxes of N_2O were highly variable and demonstrated strong association with fertilization events. Although the mechanism of N_2O generation and consumption in the soil is highly complex, it can be speculated that higher soil N content after urea application may have induced nitrification and denitrification-led N_2O emission. Further, each fertilizer application event was followed by irrigation that may have triggered the activity of nitrifying and denitrifying microbes, which resulted in greater fluxes of N_2O following fertilization event (Pathak et al. 2002; Sapkota et al. 2015).

The effect of tillage system on the seasonal trend of N₂O emission was evident only in rice (Supplementary Figure S3) not in wheat seasons (Supplementary Figures S2 and S4). In ZT rice, N₂O emission was observed right after basal fertilizer application though in smaller rate (Supplementary Figure S3 upper panel). In CT rice, on the other hand, the magnitude of N₂O emission was smaller even after second fertilizer application probably because anaerobic conditions in the field following conventional tillage and flooding in this system might have suppressed the nitrification and hence associated N₂O generation (Pandey et al. 2012; Butterbach-Bahl et al. 2013).

Cumulative N₂O emission from rice and wheat crops

N₂O fluxes from rice and wheat fields have been measured in different countries and also in different cropping systems and different agro-ecologies in India (Pathak et al. 2002; Malla et al. 2005; Bhattacharyya et al. 2012; Pandey et al. 2012; Sapkota et al. 2015). Seasonal cumulative N_2O emissions reported in the present study (0.58–3.23 and 0.38–1.93 kg N_2O-N ha⁻¹ for wheat and rice season, respectively) are within the reported range in the previous literature. Relatively higher seasonal emission in 2014–2015 wheat season than 2015–2016 wheat season (Table 3) was probably due to evenly distributed rainfall during 2014–2015 wheat season (Supplementary Figure S1) resulting in soil conditions conducive (i.e. moist but not water-logged condition) to N_2O emissions. Friedl et al. (2016) demonstrated through soil incubation study that denitrification-led N₂O emission is higher in soil with 80% water-filled pore space than in 100% saturated soil. This is also evident from the higher background emissions (i.e. N_2O emission from zero-N treatment) in the 2014-2015 wheat season than in any other crop seasons studied (Table 3). The background emission in our study ranged from 0.33–1.23, 0.09–0.52 and 0.07–0.13 kg N_2O-N ha⁻¹ during first-year wheat, rice and second-year wheat, respectively (means shown in Table 3). In this study, background N₂O emission accounted for 5–41% of total in fertilized treatments, which is comparable to the values (26–30%) reported by Gu et al. (2009). In terms of magnitude, background emission in this study was slightly smaller than that from rice-rapeseed systems in China, i.e. 0.66 kg N ha⁻¹ (Zhou et al. 2015). The differences in background emission can be attributed to different climatic conditions such as semi-arid climatic condition and soil properties such as lower soil total nitrogen content and low soil organic carbon content in our site. Such climate and soil conditions strongly affect the background emission of N₂O (Gu et al. 2009).

Annual N₂O emission from RW system in the fertilized treatments ranged from 1.07 to 5.17 (mean = 2.18) kg N₂O–N ha⁻¹. The annual N₂O emission from RW system in N75 treatment in our study (1.49 kg N₂O–N) was similar to the values (1.42 kg N₂O–N) reported by Malla et al. (2005) from IGP with application of 120 kg N ha⁻¹ in both rice and wheat, whereas emission from the plots receiving higher rates of N were much higher (Table 3). On an average, rice and wheat contributed 40% and 60% of annual N₂O emission, respectively. The contribution of rice and wheat to total annual N₂O emission was respectively 36% and 64% in CT system and 42% and 58% in ZT system.

N₂O emission factor

The N_2O EF, the percentage of fertilizer N applied that is transformed into fertilizer-induced emission, was determined individually for the three crop seasons studied, i.e. wheat 2014–2015, rice 2015 and wheat 2015–2016. Annual N₂O EF represents the data from the entire crop year consisting of 2015 rice and 2015–2016 wheat season. The fertilizer-induced N₂O EF in our study (Table 3; 0.23–0.54% for rice and 0.59–1.09% for wheat) was on slightly lower side as compared to 1% proposed by IPCC (IPCC 2007) and also reported by Albanito et al. (2017) through a review and modelling of published EF from tropical and sub-tropical agricultural systems. As soil water can directly and indirectly influence N₂O emission, prolonged dry period in our wheat field induced by semi-arid climatic condition (Supplementary Figure 1) coupled with limited number of irrigations might have suppressed nitrification- denitrification-induced N₂O emission in our study. N₂O EF in our study was smaller for rice which was comparable to 0.34% reported by Kumar et al. (2002) from the RW systems of IGP. EFs in our study did not increase with increasing N rate, contrary to a recent study (e.g. Shcherbak et al. 2014) who reported increased N₂O-EF with increasing N rate. Given that N₂O emission from soil is controlled by a multitude of soil and climatic variables and controlled by management factors, it is difficult to derive a general pattern of N₂O dependency on N input. For example, Kim et al. (2013) examined the dependency of N₂O emission on N input using published dataset and reported that N₂O-EF remains constant or increase or decrease non-linearly with changing N rates.

We speculate that higher soil pH and low soil organic carbon in our study site (i.e. pH = 8 SOC = 0.56%; Table 1) are the main factors for absence of response of N₂O emission at higher N rates. For example, Wang et al. (2017), through a global meta-analysis of 1,104 field measurements, reported that N₂O emission in acidic soils is more sensitive to fertilization rate than that in alkaline soils. Further, in our C limited soil, increasing N rate above certain amount might have increase soil N beyond the capacity of soil microbes to take up and utilize thereby slowing down the N₂O production rate and finally reaching a steady-state as hypothesized by Kim et al. (2013). Even in the meta-analysis of Shcherbak et al. (2014), higher N₂O EF with higher N rates were evident only in the soils with carbon content >1.5% and pH<7. Although the apparent causality between soil pH and N₂O emission has been studied previously, the influencing mechanisms of soil pH on N₂O emission have not been completely understood. For example, soil liming in laboratory study suppressed N₂O emission (Shaaban et al. 2015) and soil acidification through intensive fertilization significantly enhanced N₂O reductase enzyme, leading to higher N₂O/N₂ ratio during denitrification (Bakken et al. 2012).

Optimization of N fertilizer

An ideal fertilizer rate for any crop would be the one that promotes the dual goals of high crop yield and low N₂O flux (Sapkota et al. 2018). Although, the control treatment resulted in the lowest yieldscaled N₂O emission (Table 3), it should be noted that for a N rate to be financially viable, it must produce sufficient crop yield. Both rice and wheat yields increased until N rate reached to 225 kg N ha⁻¹ (Figure 2), the rate of yield increment slowed down above these N rates in both the crops. The optimum rate of N fertilizer application in rice and wheat ranged from 120–200 to 50–185 kg N ha⁻¹, respectively (Figure 2). This optimum range for N in both crop includes a range between maximum rate of marginal return and maximum yield with positive marginal rate of return. Therefore, application of N fertilizer above these ranges would not be financially viable or environmentally suitable.

Conclusion

This study provides the 'optimum' range for fertilizer N rate for rice and wheat to maximize crop yield and economic benefit and to minimize N_2O emission. Overall, fertilizer-induced N_2O -EF in our study

14 👄 T. B. SAPKOTA ET AL.

was about 0.42% in rice and 0.8% for wheat. Alkaline soil with low organic carbon in the experimental site might be responsible for reducing fertilizer-induced N₂O emission in our study, particularly under high N application rate. Based on this analysis, N rate of 120–200 kg N ha⁻¹ for rice and 50–185 kg N ha⁻¹ for wheat is agronomically productive, economically viable and environmentally sustainable for the RW system of North-West Indo-Gangetic Plains.

Acknowledgements

This work was carried out by International Maize and Wheat Improvement Center (CIMMYT) and implemented as part of the CGIAR Research Program on Climate Change, Agriculture and Food Security (CCAFS), with support from the CGIAR Trust Fund and through bilateral funding agreements. For details please visit https://ccafs.cgiar.org/donors. The views expressed in this document cannot be taken to reflect the official opinions of these organizations. We sincerely acknowledge the input and support provided by field staffs during trial management, GHG sampling and analysis. We are very thankful to ICAR-Central Soil Salinity Research Institute for providing experimental field and hosting GHG analytical laboratory.

Disclosure statement

No potential conflict of interest was reported by the authors.

ORCID

Tek B. Sapkota (D) http://orcid.org/0000-0001-5311-0586

References

- Albanito F, Lebender U, Cornulier T, Sapkota TB, Brentrup F, Stirling C, Hillier J. 2017. Direct nitrous oxide emissions from tropical and sub-tropical agricultural systems A review and modelling of emission factors. Sci Rep. 7:1–12. doi:10.1038/srep44235.
- Bakken LR, Bergaust L, Liu B, Frostegard A. 2012. Regulation of denitrification at the cellular level: a clue to the understanding of N₂O emissions from soils. Philos Trans R Soc B Biol Sci. 367:1226–1234. doi:10.1098/rstb.2011.0321.
- Bhattacharyya P, Roy KS, Neogi S, Adhya TK, Rao KS, Manna MC. 2012. Effects of rice straw and nitrogen fertilization on greenhouse gas emissions and carbon storage in tropical flooded soil planted with rice. Soil Tillage Res. 124:119–130. doi:10.1016/j.still.2012.05.015.
- Bijay-Singh. 2017. Management and use efficiency of fertilizer nitrogen in prodution of cereals in India-issues and strategies. In: Abrol YP, Adhya TK, Aneja VP, Raghuram N, Pathak H, Kulshrestha U, Sharma C, Bijay-Singh, editors. Indian nitrogen assess sources react nitrogen, environ clim eff manag options policies. Duxford (UK): Woodhead Publishing; p. 149–159.
- Bouwman AF, Boumans LJM, Batjes NH. 2002. Emissions of N₂O and NO from fertilized fields: summary of available measurement data. Global Biogeochem Cycles. 16:1058. doi:10.1029/2001GB001811.
- Butterbach-Bahl K, Baggs EM, Dannenmann M, Kiese R, Zechmeister-Boltenstern S. 2013. Nitrous oxide emissions from soils: how well do we understand the processes and their controls? Philos Trans R Soc B Biol Sci. 368. doi:10.1098/ rstb.2013.0122.

CoHort S 2012. CoHort Software. Monterey (CA USA) [Internet]. Available from: www.cohort.com

- Cole CV, Duxbury J, Freney J, Heinemeyer O, Minami K, Mosier A, Paustian K, Rosenberg N, Sampson N, Sauerbeck D. 1997. Global estimates of potential mitigation of greenhouse gas emissions by agriculture. Nutr Cycl Agroecosyst. 49:221–228.
- De Klein C, Novoa RSA, Ogle S, Smith KA, Rochette P, Wirth TC, McConkey BG, Mosier A, Rypdal K, Walsh M. 2006. N₂O emissions from managed soils, and CO₂ emissions from lime and urea application. IPCC Guidel Natl Greenh Gas Invent Prep Natl Greenh Gas Invent Progr. 4:1–54.
- Foley JA, Ramankutty N, Brauman KA, Cassidy ES, Gerber JS, Johnston M, Mueller ND, O'Connell C, Ray DK, West PC, et al. 2011. Solutions for a cultivated planet. Nature. 478:337. doi:10.1038/nature10452.
- Friedl J, Scheer C, Rowlings DW, McIntosh HV, Strazzabosco A, Warner DI, Grace PR. 2016. Denitrification losses from an intensively managed sub-tropical pasture - Impact of soil moisture on the partitioning of N₂ and N₂O emissions. Soil Biol Biochem. 92:58–66. doi:10.1016/j.soilbio.2015.09.016.
- Gathala MK, Ladha JK, Kumar V, Saharawat YS, Kumar V, Sharma PK, Sharma S, Pathak H. 2011. Tillage and crop establishment affects sustainability of south asian rice-wheat system. Agron J. 103:961–971.

- GOI/MoEF. 2012. India: second national communication to the united nations framework convention on climate change. New Delhi (India): Ministry of Environment and Forests, Government of India.
- Gomez K, Gomez A. 1984. Statistical procedures for agricultural research. New York (USA): John Wiley and Sons.
- Gu J, Zheng X, Zhang W. 2009. Background nitrous oxide emissions from croplands in China in the year 2000. Plant Soil. 320:307–320.
- Halvorson AD, Del Grosso SJ, Jantalia CP. 2011. Nitrogen source effects on soil nitrous oxide emissions from strip-till corn. J Environ Qual. 40:1775.
- Hoben JP, Gehl RJ, Millar N, Grace PR, Robertson GP. 2011. Nonlinear nitrous oxide (N₂O) response to nitrogen fertilizer in on-farm corn crops of the US Midwest. Glob Chang Biol. 17:1140–1152.
- IPCC. 2007. Climate change: the physical science basis. summary for policymakers. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge (UK): Cambridge University Press.
- IPCC. 2013. Climate change 2013. The physical science basis. Working group I contribuiton to the fifth assessment report of the intergovernmental panel on climate change. Chapter 8: Anthropogenic and Natural RAdiative Forcing. Intergovernmental Panel on Climate Change.
- Kim DG, Hernandez-Ramirez G, Giltrap D. 2013. Linear and nonlinear dependency of direct nitrous oxide emissions on fertilizer nitrogen input: A meta-analysis. Agric Ecosyst Environ. 168:53–65. doi:10.1016/j.agee.2012.02.021.
- Kumar S, Jain MC, Kumar U. 2002. Emission of nitrous oxide from rice-wheat systems of indo-gangetic plains of india. Envion Monit Assess. 77:163–178.
- Ladha JK, Kumar V, Alam MM, Sharma S, Gathala MK, Chandna P, Saharawat YS, Balasubramanian V. 2009. Integrating crop and resource management technologies for enhanced productivity, profitability, and sustainability of the ricewheat system in South Asia. In: Ladha JK, Yadvinder-Singh, Erenstein O, Hardy B, editors. Integr crop resour manag rice-wheat syst South Asia. Los Baños (Philippines): IRRI; p. 69–108.
- Ma BL, Wu TY, Tremblay N, Deen W, Morrison MJ, Mclaughlin NB, Gregorich EG, Stewart G. 2010. Nitrous oxide fluxes from corn fields: on-farm assessment of the amount and timing of nitrogen fertilizer. Glob Chang Biol. 16:156–170.
- Malla G, Bhatia A, Pathak H, Prasad S, Jain N, Singh J. 2005. Mitigating nitrous oxide and methane emissions from soil in rice–wheat system of the Indo-Gangetic plain with nitrification and urease inhibitors. Chemosphere. 58:141–147.
- McSwiney CP, Robertson GP. 2005. Nonlinear response of N₂O flux to incremental fertilizer addition in a continuous maize (*Zea mays* L.). Cropping Syst Glob Chang Biol. 11:1712–1719.
- Millar N, Urrea A, Kahmark K, Shcherbak I, Robertson GP, Ortiz-Monasterio I. 2018. Nitrous oxide (N₂O) flux responds exponentially to nitrogen fertilizer in irrigated wheat in the Yaqui Valley, Mexico. Agric Ecosyst Environ. 261:125–132. doi:10.1016/j.agee.2018.04.003.
- Myhre G, Shindell D, Bréon F-M, Collins W, Fuglestvedt J, Huang J, Koch D, Lamarque J-F, Lee D, Mendoza B. 2013. Anthropogenic and natural radiative forcing. In: Stocker TF, Qin D, Plattner G-K, Tingor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex V, PM M, et al., editors. Clim Chang 2013. Phys Sci Basis Contrib Work Gr I to Fifth Assess Rep Intergov Panel Clim Change, Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA. p. 659–740.
- Pandey D, Agrawal M, Singh J, Bohra JS. 2012. Greenhouse gas emissions from rice crop with different tillage permutations in rice wheat system. Agric Ecosyst Environ. 159:133–144. doi:10.1016/j.agee.2012.07.008.
- Pathak H, Bhatia A, Prasad S, Singh S, Kumar S, Jain MC, Kumar U. 2002. Emission of nitrous oxide from rice-wheat systems of indo-gangetic plains of India. Environ Monit Assess. 77:163–178.
- Raut N, Dörsch P, Sitaula BK, Bakken LR. 2012. Soil acidification by intensified crop production in South Asia results in higher N₂O/(N₂+ N₂O) product ratios of denitrification. Soil Biol Biochem. 55:104–112.
- Ravishankara AR, Daniel JS, Portmann RW. 2009. Nitrous Oxide (N₂O): the dominant ozone-depleting substance emitted in the 21st century. Science. 123:2007–2010.
- Sapkota TB, Aryal JP, Khatri-chhetri A, Shirsath PB. 2018. Identifying high-yield low-emission pathways for the cereal production in South Asia. Mitig Adapt Strateg Glob Chang. 23:621–641.
- Sapkota TB, Jat ML, Shankar V, Singh LK, Rai M, Grewal MS, Stirling CM. 2015. Tillage, residue and nitrogen management effects on methane and nitrous oxide emission from rice–wheat system of Indian Northwest Indo-Gangetic Plains. J Integr Environ Sci. 12.
- Sapkota TB, Majumdar K, Jat ML, Kumar A, Bishnoi DK, Mcdonald AJ, Pampolino M. 2014b. Precision nutrient management in conservation agriculture based wheat production of Northwest India: profitability, nutrient use efficiency and environmental footprint. F Crop Res. 155:233–244.
- Sapkota TB, Rai M, Singh LK, Gathala MK, Jat ML, Sutaliya JM, Bijarniya D, Jat MK, Jat RK, Parihar CM, et al. 2014a. Greenhouse gas measurement from smallholder production systems: guidelines for static chamber method. New Delhi (India): International Maize and Wheat Improvement Center (CIMMYT).
- Shaaban M, Pengan Q, Hu R, Wu Y, Lin S, Zhao J. 2015. Dolomite application to acidic soils: a promising option for mitigating N₂O emissions. Environ Sci Pollut Res. 22:19961–19970.
- Shcherbak I, Millar N, Robertson GP. 2014. Global metaanalysis of the nonlinear response of soil nitrous oxide (N₂O) emissions to fertilizer nitrogen. Proc Natl Acad Sci. 111:9199–9204.

16 🕒 T. B. SAPKOTA ET AL.

- Singh VK, Dwivedi BS, Shukla AK, Chauhan YS, Yadav RL. 2005. Diversification of rice with pigeonpea in a rice-wheat cropping system on a Typic Ustochrept: effect on soil fertility, yield and nutrient use efficiency. F Crop Res. 92:85–105.
- Snyder CS, Bruulsema TW, Jensen TL. 2007. Greenhouse gas emissions from cropping systems and the influence of fertilizer management-A Literature Review. Norcross (GA): International Plant Nutrition, Institute.
- Stehfest E, Bouwman L. 2006. N₂O and NO emission from agricultural fields and soils under natural vegetation: summarizing available measurement data and modeling of global annual emissions. Nutr Cycl Agroecosyst. 74:207–228.
- Tewatia RK, Chanda TK. 2017. Trends in fertilizer nitrogen proudction and consumption in India. In: Abrol YP, Adhya TK, Aneja VP, Raghuram N, Pathak H, Kulshrestha U, Sharma C, Bijay-Singh, editors. Indian nitrogen assess sources react nitrogen, environ clim eff manag options policies. Duxford (United Kingdom): Woodhead Publishing; p. 45–56.
- Wang Y, Guo J, Vogt RD, Mulder J, Wang J, Zhang X. 2017. Soil pH as the chief modifier for regional nitrous oxide emissions: new evidence and implications for global estimates and mitigation. Glob Chang Biol. 1–10. doi:10.1111/ gcb.13966.
- Zhou M, Zhu B, Brüggemann N, Wang X, Zheng X, Butterbach-bahl K. 2015. Nitrous oxide and methane emissions from a subtropical rice rapeseed rotation system in China : A 3-year fi eld case study. Agric Ecosyst Environ. 212:297–309. doi:10.1016/j.agee.2015.07.010.