


RESEARCH ARTICLE

Open Access



Design, synthesis, antiviral bioactivities and interaction mechanisms of penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold

Lijuan Chen^{1†}, Xiaobin Wang^{1,2†}, Xu Tang¹, Rongjiao Xia¹, Tao Guo¹, Cheng Zhang¹, Xiangyang Li¹ and Wei Xue^{1*} 

Abstract

Background: penta-1,4-diene-3-one oxime ether and quinazolin-4(3*H*)-one derivatives possess favorable agricultural activities. Aiming to discover novel molecules with highly-efficient agricultural activities, a series of penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold were synthesized and evaluated for their antiviral activities.

Result: Antiviral bioassays indicated that some title compounds exhibited significant antiviral activity against tobacco mosaic virus (TMV). In particular, compounds **8c**, **8j** and **8k** possessed appreciable curative activities against TMV *in vivo*, with half-maximal effective concentration (EC₅₀) values of 138.5, 132.9 and 125.6 μg/mL, respectively, which are better than that of ningnanmycin (207.3 μg/mL). Furthermore, the microscale thermophoresis experiments (MST) on the interaction of compound **8k** with TMV coat protein (TMV CP) showed **8k** bound to TMV CP with a dissociation constant of 0.97 mmol/L. Docking studies provided further insights into the interaction of **8k** with the Arg90 of TMV CP.

Conclusions: Sixteen penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold were designed, synthesized, and their antiviral activities against TMV were evaluated. Antiviral bioassays indicated that some target compounds exhibited remarkable antiviral activities against TMV. Furthermore, through the MST and docking studies, we can speculate that **8k** inhibited the virulence of TMV by binding Arg90 in TMV CP. These results indicated that this kind of penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold could be further studied as potential alternative templates in the search for novel antiviral agents.

Keywords: Penta-1,4-diene-3-one, Oxime ether, Quinazolin-4(3*H*)-one, Antiviral activity, Tobacco mosaic virus, Interaction mechanisms, Molecular docking study

*Correspondence: wxue@gzu.edu.cn

[†]Lijuan Chen and Xiaobin Wang contributed equally to this work

¹ State Key Laboratory Breeding Base of Green Pesticide and Agricultural Bioengineering, Key Laboratory of Green Pesticide and Agricultural Bioengineering, Ministry of Education, Guizhou University, Guiyang 550025, China

Full list of author information is available at the end of the article



Background

Tobacco mosaic virus (TMV) is an important plant virus that caused serious crop loss [1], and unfortunately, there are few effective antiviral agents to control the infection of TMV [2]. Ningnanmycin, exhibited a dual targeting ability against the TMV coat protein [3], is the best anti-TMV agent that was isolated from *Streptomyces noursei* var. [4]. However, its application in field trial is largely limited by its photosensitivity, water stickiness and high control costs [5, 6]. Therefore, developing novel, highly-efficient, and environmentally benign antiviral agents remains a daunting task in pesticide sciences.

Natural product-based compounds consistently display more advantages than traditional synthetic chemicals, such as low toxicity, easy decomposition, environmental friendliness and unique modes of action [7–10]. As important analogs of curcumin isolated from turmeric, penta-1,4-diene-3-ones are viewed as lead compounds due to their significant applications in the development of pharmaceuticals and agrochemicals [11–15]. In the last decade, series of penta-1,4-diene-3-one derivatives bearing pyrazole [16], quinazoline [17], quinazolin-4(3*H*)-one [18], 1,3,4-thiadiazole [19], 1,3,4-oxadiazole [20], emodin [21], glucopyranoside [22], oxime ether [23], oxime ester [15], pyridine [24], rutin [25], 4-thioquinazoline [26] and purine [27] moieties were reported for their fine antiviral activities against plant viruses (Fig. 1). However, the antiviral activities of these penta-1,4-diene-3-one derivatives were adequate but still not satisfactory.

Quinazolin-4(3*H*)-ones, important nitrogenous heterocycles that present extensive bioactivities including antibacterial [28], anti-inflammatory [29], antifungal [30], antimalarial [31] and anticancer [32] properties, were reported for their potent antiviral activities against plant viruses. Recently, Ma et al. evaluated antiviral activities of quinazolin-4(3*H*)-one derivatives containing penta-1,4-diene-3-one moiety, and found these compounds can

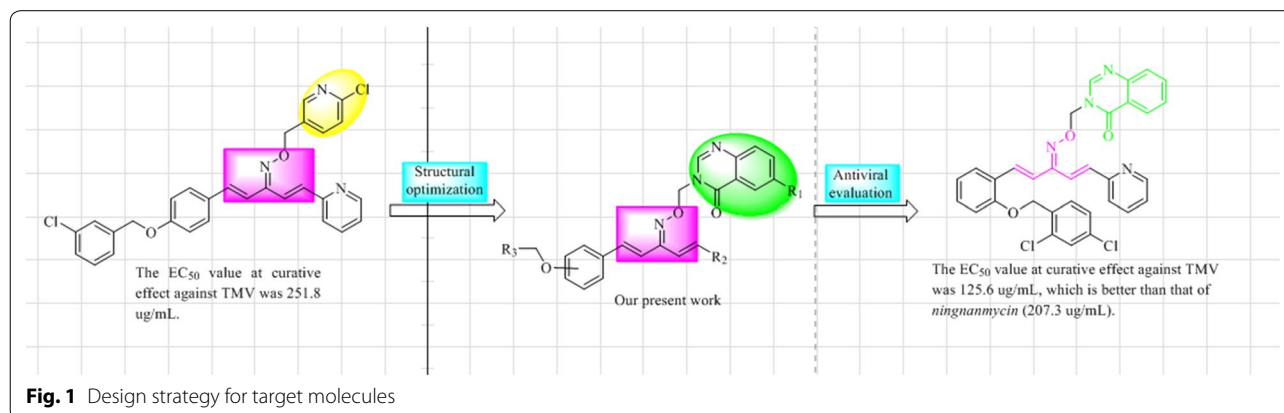
effectively control some plant viruses [18]. Chen et al. reported that cucumber mosaic virus could be inhibited effectively by some quinazolin-4(3*H*)-one derivatives containing malonate moiety [33].

To develop novel, highly-efficient and environmentally benign virucides, we introduced a quinazolin-4(3*H*)-one scaffold into penta-1,4-diene-3-one oxime ether derivatives, which might generate some novel curcumin derivatives with potent biological activities. Thus, 16 penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold were designed, synthesized and evaluated for their antiviral activity against TMV in vivo.

Results and discussion

Chemistry

The infrared spectrum (IR), ^1H nuclear magnetic resonance (^1H NMR), ^{13}C nuclear magnetic resonance (^{13}C NMR) and high resolution mass spectrum (HRMS) spectra of title compounds are shown in Additional file 1. IR spectra exhibit characteristic absorptions at 1650–1500 cm^{-1} , which indicate the presence of $-\text{C}=\text{N}-$ fragment. The signals at 1750–1650 and 1260–1210 cm^{-1} are attributed to the stretching frequencies of $-\text{C}=\text{O}$ and $-\text{C}-\text{O}-\text{N}=\text{groups}$, respectively. In ^1H NMR spectra, multiplet signals at δ 8.20–6.50 ppm show the presence of protons in olefinic bonds and aromatic nucleuses, and a singlet at δ 5.50–5.00 ppm reveals the presence of $-\text{CH}_2-$ groups. Absorption signals at δ 155–165 and 65–75 ppm in ^{13}C NMR spectra confirm the presences of $-\text{C}=\text{N}-$ and $-\text{CH}_2-$ groups, respectively. Noteworthy, the ^1H NMR and ^{13}C NMR spectra of title compounds also reveal that title compounds don't have chemical isomers, which are consist with a result described in our previous work [22]. In the HRMS spectra of target compounds show characteristic absorption signals of $[\text{M}+\text{H}]^+$ ions that is consistent with their molecular weight.



Antiviral activity screening of title compounds against TMV in vivo

Using *N. tabacum* L. leaves under the same age as the test subjects, the curative and protective activities against TMV in vivo at 500 µg/mL were evaluated by the half-leaf blight spot method [14, 18, 34, 35] and the obtained results were listed in Table 1. Preliminary bioassays indicated that the curative and protective effects of title compounds against TMV at 500 µg/mL ranged 31.7–59.1% and 39.4–69.8%, respectively. Among them, the curative effects of compounds **8c**, **8d**, **8e**, **8j** and **8k** against TMV were 55.9, 53.4, 54.6, 55.4 and 59.1%, respectively, which are better than that of ningnanmycin (51.8%). In addition, compounds **8c**, **8j** and **8k** exhibited significant protection effects against TMV at 500 µg/mL, with corresponding inhibitory rates of 69.8, 67.7 and 72.0%, respectively, which are superior to that of ningnanmycin (65.7%).

Based on the preliminary bioassays in Table 1, the curative activities of compounds **8c**, **8j** and **8k** against TMV at 500, 250, 125, 62.5 and 31.25 µg/mL were respectively tested to obtain the corresponding EC₅₀ values. As showed in Table 2, compounds **8c**, **8j** and **8k** showed excellent curative effects against TMV, with corresponding EC₅₀ values of 138.5, 132.9 and 125.6 µg/mL, respectively, which are better than that of ningnanmycin (207.3 µg/mL). These results indicated that this kind of penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold could be further studied as potential alternative templates in the search for novel antiviral agents.

Structure activity relationship (SAR) of title compounds against TMV

As indicated in Tables 1 and 2, most of penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold showed significant antiviral activities against TMV, and some structure–activity relationships can be analyzed and summarized. First, most of the target compounds bearing the same substituted fragment exhibited higher protective activity than curative activity against TMV. For example, the protective effects of compounds **8a**, **8b**, **8c**, **8e**, **8f**, **8g**, **8h**, **8i**, **8j**, **8k**, **8l**, **8m**, **8n**, **8o** and **8p** are 59.4, 56.4, 69.8, 60.1, 39.4, 55.4, 52.1, 47.7, 67.7, 72.0, 48.9, 54.4, 50.3, 65.1 and 53.2%, respectively, which were better than their curative effects (44.6, 40.1, 55.9, 54.6, 31.7, 33.4, 44.9, 45.3, 55.4, 59.1, 45.8, 50.5, 37.3, 53.5 and 37.4%, respectively). Second, the presence of a quinazolin-4(3*H*)-one fragment can effectively enhance the anti-TMV activities of target compounds. For example, the curative activities of compounds **8a**, **8b**, **8c**, **8d**, **8j** and **8k** (R₁=H) were 44.6, 40.1, 55.9, 53.4, 55.4 and 59.1%, respectively, which are better than that of compounds **8g**, **8f**, **8h**, **8i**, **8l** and **8m** (R₂=Cl; 33.4, 31.7, 44.9, 45.3, 45.8 and 50.5%, respectively). Third, antiviral bioassays reveals that the pyridine fragment, not thiophene fragment, was favorable for the anti-TMV activities of penta-1,4-diene-3-one oxime ether derivatives. For instances, the curative inhibition of compound **8n** was 37.3%, which was lower than that of compounds **8k** (54.6%). Furthermore, when the R₃ was substituted with 2-ClPh, 4-ClPh and

Table 1 Antiviral effects of title compounds against TMV in vivo at 500 µg/mL

Compound	R ₁	R ₂	R ₃	–O–	Curative activity(%) ^a	Protective activity(%) ^a
8a	H	Pyridin-2-yl	3-ClPh	4-O	44.6 ± 1.7	59.4 ± 1.6
8b	H	Pyridin-2-yl	3-MePh	4-O	40.1 ± 2.4	56.4 ± 1.3
8c	H	Pyridin-2-yl	2-ClPh	4-O	55.9 ± 2.9	69.8 ± 5.8
8d	H	Pyridin-2-yl	4-ClPh	4-O	53.4 ± 3.9	44.7 ± 3.2
8e	H	Pyridin-2-yl	2,4-di-ClPh	4-O	54.6 ± 4.5	60.1 ± 3.4
8f	Cl	Pyridin-2-yl	3-MePh	4-O	31.7 ± 3.8	39.4 ± 4.0
8g	Cl	Pyridin-2-yl	3-ClPh	4-O	33.4 ± 5.6	55.4 ± 4.9
8h	Cl	Pyridin-2-yl	2-ClPh	4-O	44.9 ± 4.7	52.1 ± 4.1
8i	Cl	Pyridin-2-yl	4-ClPh	4-O	45.3 ± 4.8	47.7 ± 6.3
8j	H	Pyridin-2-yl	2-ClPh	2-O	55.4 ± 4.3	67.7 ± 1.7
8k	H	Pyridin-2-yl	2,4-di-ClPh	2-O	59.1 ± 4.2	72.0 ± 3.9
8l	Cl	Pyridin-2-yl	2-ClPh	2-O	45.8 ± 6.1	48.9 ± 4.3
8m	Cl	Pyridin-2-yl	2,4-di-ClPh	2-O	50.5 ± 4.9	54.4 ± 3.9
8n	H	Thiophene-2-yl	2,4-di-ClPh	4-O	37.3 ± 6.1	50.3 ± 5.1
8o	H	Pyridin-3-yl	2-ClPh	4-O	53.5 ± 2.5	65.1 ± 6.2
8p	H	Pyridin-3-yl	3-MePh	4-O	37.4 ± 3.6	53.2 ± 3.5
Ningnanmycin ^b	–	–	–	–	51.8 ± 4.3	65.7 ± 2.9

^a Average of three replicates

^b Ningnanmycin was used as a comparison

Table 2 Curative effects of compounds 8c, 8j and 8k against TMV

Compound	Concentration ($\mu\text{g}/\text{mL}$) ^a	Curative activity (%) ^a	Toxic regression equation	r	EC ₅₀ ($\mu\text{g}/\text{mL}$)
8c	500	55.9 ± 2.9	$y = 0.3444x + 4.2222$	0.9860	138.5
	250	51.2 ± 5.5			
	125	48.8 ± 4.9			
	62.5	44.2 ± 5.6			
	31.25	38.9 ± 5.0			
8j	500	55.4 ± 4.3	$y = 0.3111x + 4.3085$	0.9746	132.9
	250	52.2 ± 5.1			
	125	48.7 ± 5.3			
	62.5	46.1 ± 5.6			
	31.25	39.9 ± 4.8			
8k	500	59.1 ± 4.2	$y = 0.4154x + 4.1258$	0.9836	125.6
	250	56.1 ± 5.6			
	125	50.0 ± 3.8			
	62.5	43.6 ± 3.5			
	31.25	40.6 ± 5.1			
Ningnamycin ^b	500	51.6 ± 4.3	$y = 0.5475x + 3.4987$	0.9761	207.3
	250	41.2 ± 3.8			
	125	33.3 ± 2.6			
	62.5	30.3 ± 4.6			
	31.25	26.2 ± 3.4			

^a Average of three replicates

^b Ningnanmycin was used as a comparison

2,4-di-ClPh groups, the corresponding compounds **8c**, **8j** and **8k** showed better curative effects against TMV, with the EC₅₀ values of 138.5, 132.9 and 125.6 $\mu\text{g}/\text{mL}$, respectively, which are better than that of ningnanmycin (207.3 $\mu\text{g}/\text{mL}$).

The expression and purification of TMV CP

pET28a-TMV CP vector were constructed and preserved in our lab [36, 37]. TMV CP genes were over-expressed in pET28a-TMV CP vector when the final concentration was increased to 0.8 mM isopropoyl- β -D-galactopyranoside (IPTG) and the solution was left overnight at 16 °C. More than 90% of the TMV CP protein was eluted. Then, the dealt proteins were loaded in desalting column, in a buffer containing 10 mM PB and 100 mM sodium chloride pH 7.2, the retention time of the TMV CP protein was determined with 19.5 kDa with His-tags.

The binding studies of TMV CP and 8k

We studied the interactions between **8k** and TMV CP using Monolith NT. 115 (Nano Temper Technologies, Germany) [38, 39]. The results showed that the **8k** bound to TMV CP with a dissociation constant (Kd) of $0.97 \pm 0.69 \mu\text{M}$. An affinity between **8k** and TMV CP was

far greater than the affinity between ningnanmycin (Control) and TMV CP. Base on anti-TMV activities and MST results. We speculate TMV CP protein was a potential target of **8k**. The affinity curves are shown in Fig. 2.

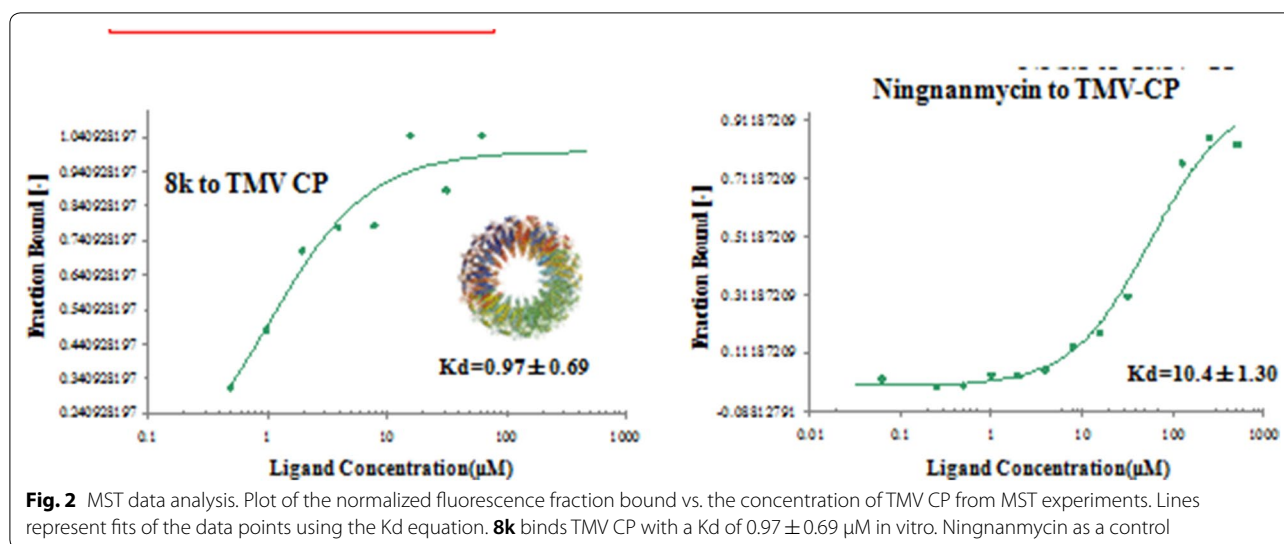
Molecular docking of 8k and TMV CP

To identify the **8k** recognition sites in TMV CP, we performed molecular docking using the Gold method with 200 cycles. After optimization, we found that Arg90 in the TMV CP shared one hydrogen bonds with **8k**, C=O–H–N = 3.24 Å (Fig. 3c). In our previously reported, Arg90 was an important residue for TMV RNA binding [38, 39]. These analyses indicated that Arg90 was the key residue to infect the activity of **8k**.

Methods and materials

Chemistry

Melting points of synthesized compounds were determined under an XT-4 binocular microscope (Beijing Tech Instrument Co., China) and were not corrected. The IR were recorded from KBr disks using a Bruker VECTOR 22 spectrometer (Bruker, USA). NMR spectra were obtained on a JEOL-ECX500 NMR spectrometer (JEOL, Japan) at room temperature using tetramethylsilane as an internal standard. Reaction was monitored by



thin-layer chromatography (TLC) on silica gel GF₂₄₅ (400 mesh). Mass spectral studies were performed on a quadrupole/electrostatic field orbitrap mass spectrometer (Thermo Scientific, USA). The micro thermophoresis of the compound and TMV CP was determined by a micro thermophoresis instrument (NanoTemper Technologies GmbH, Germany); the fluorescence spectroscopy of the compound interacting with TMV CP was determined by FluoroMax-4 fluorescence spectrometer (HORIBA Scientific, France). All reagents and reactants were purchased from commercial suppliers and were analytical grade or chemically pure. The synthetic route to penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold was shown in Scheme 1. Intermediates **2** and **7** were prepared according to the reported methods [16, 31].

Expression and purification of TMV CP

According to previous procedure described in the literature [36], The pET28a-TMV CP as a expression vector, which was stored at -80 °C in our lab containing the full-length TMV CP gene. After overnight culture, the *Escherichia coli* strain BL21(DE3) containing the plasmid pET28a-TMV CP freshly transformed, and was transferred to 1 L Luria broth. At 37 °C in Luria-Bertani medium, the cells were grown which were supplemented with 50 $\mu\text{g}/\text{mL}$ kanamycin and shaken at 200 rpm until the OD₆₀₀ was 0.8. At 16 °C, using 0.8 mM IPTG inducing the protein expression overnight, then centrifugation to obtained the cells, and stored at -80 °C. When analyzed, the cells were resuspended in lysis buffer (500 mM NaCl, 20 mM PB, 5 mM β -mercaptoethanol, 30 mM imidazole and 5% glycerol, pH 7.2) and then use sonication to lysed

at 4 °C. By centrifugation at 12,000g for 30 min at 4 °C, the lysate was clarified, the soluble supernatants were loaded onto a 5 mL Ni-NTA column (GE Healthcare, USA), and using a linear gradient of 30–350 mM imidazole (pH 7.2) to eluting the protein. using a desalting column (GE Healthcare, USA) attached to an AKTA purifier protein liquid chromatography system (GE Healthcare, USA), the crude protein was performed at 4 °C, and the fractions containing target protein with His-tags were pooled, we use ultrafiltration (10 kDa cut-off) concentrated to a suitable concentration. Using a Genequant100 (GE Healthcare, USA) to determined the dealt protein concentration, stored at -80 °C until further analysis.

Interaction studies between **8k** and TMV CP

According to previously classic method, the binding was calculated for MST Monolith NT. 115 (Nano Temper Technologies, Germany) [38, 39]. Using a NT-647 dye (Nano Temper Technologies, Germany) incubating 0.5 μM of purified recombinant proteins with A range of ligands from 0 to 5 μM were for 5 min, and using the final concentration of 20 nM to the thermophoresis experiment. The selected compounds in DMSO were made a 16 point dilution series. Each compound dilution series was subsequently transferred to protein solutions (10 mM Tris-HCl and 100 mM sodium chloride pH 7.5, 0.05% Tween-20). The labeled TMV CP with each dilution point (1:1 mix) were incubation 15 min at room temperature, then the standard capillaries (NanoTemper Technologies, Germany) were filling into the samples. Under a setting of 20% LED and 40% IR laser, using the Monolith NT.115 microscale thermophoresis system (NanoTemper Technologies,

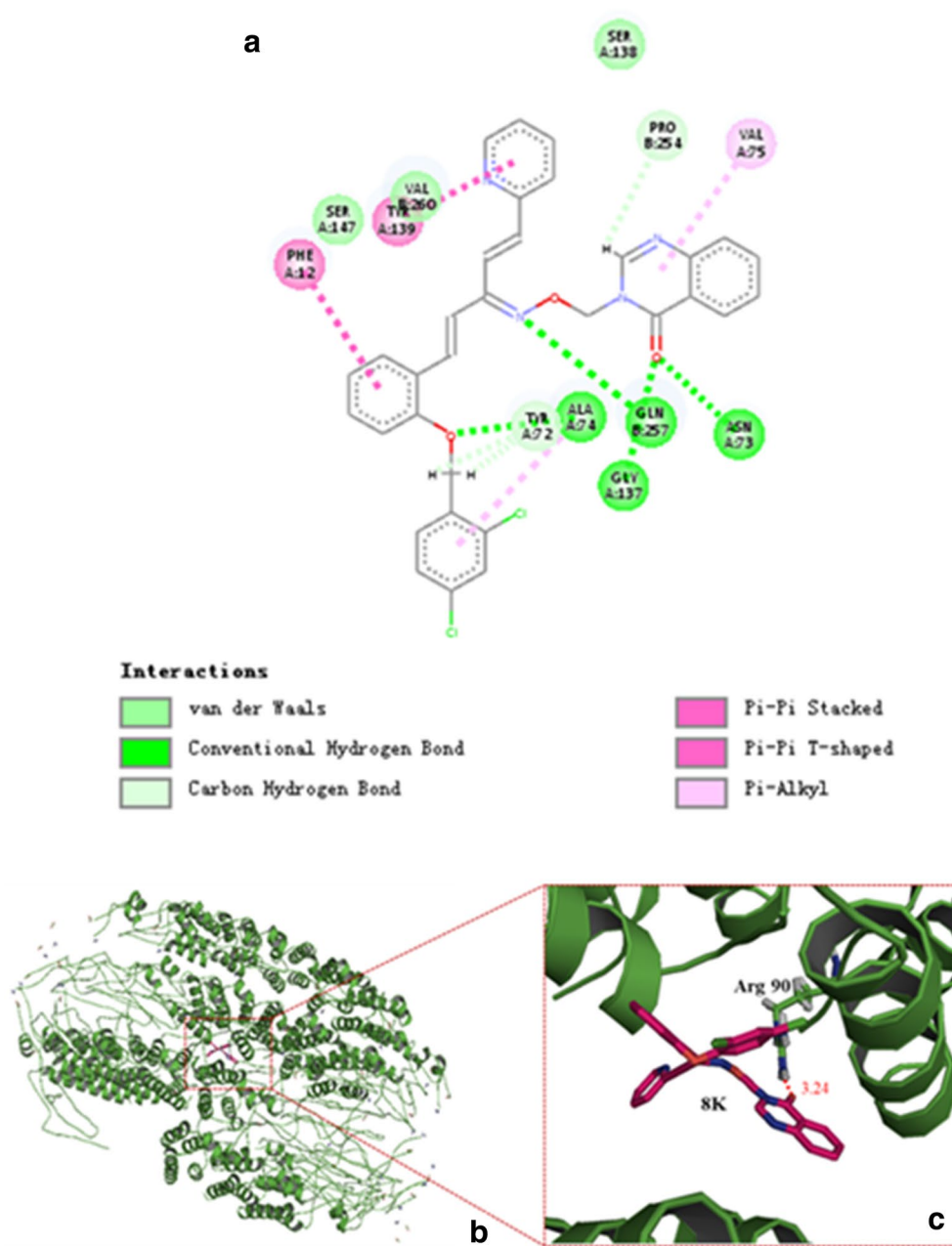
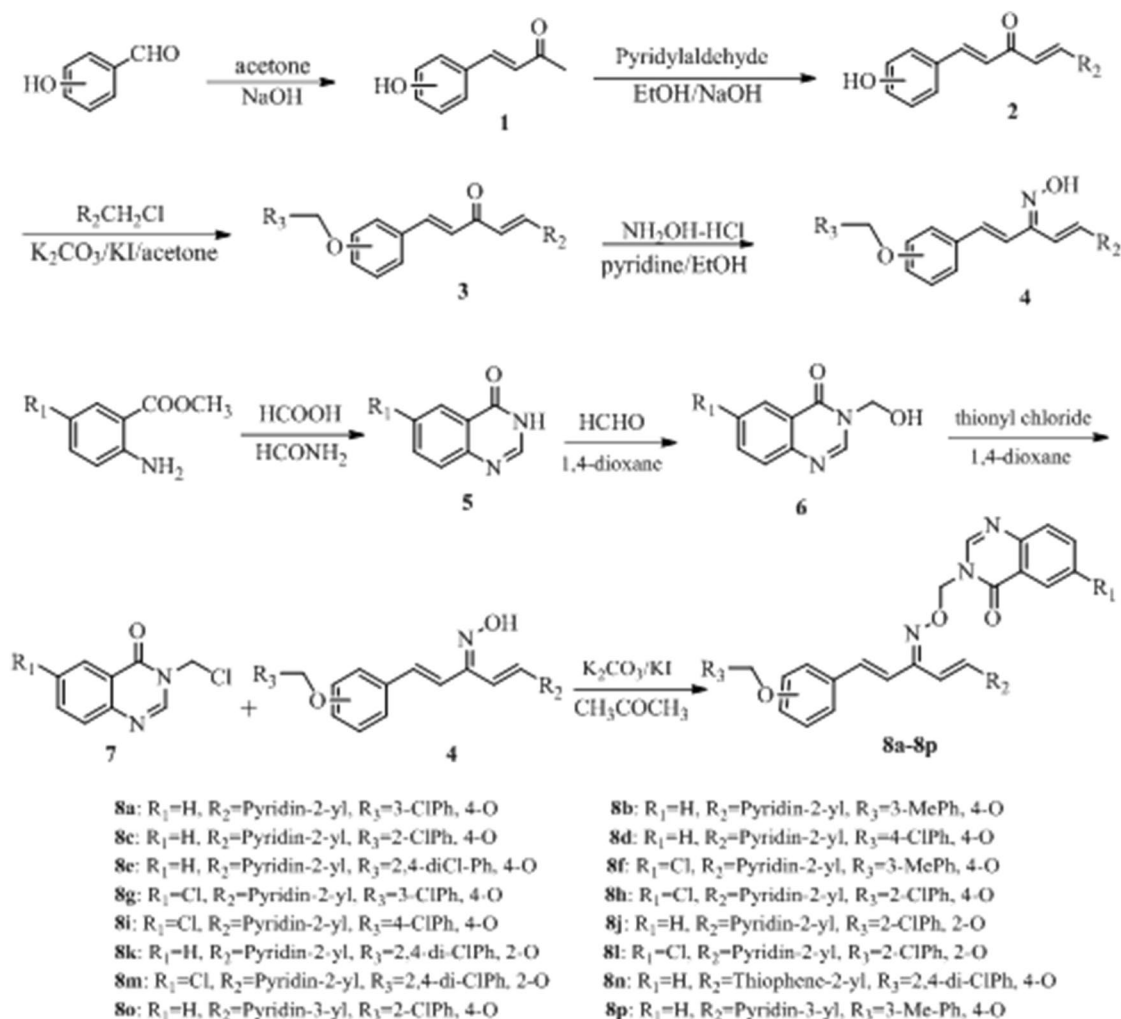


Fig. 3 The 2D diagram **a** was drawn by Discovery Studio 4.5. Docking analysis of the interactions between **8k** and TMV CP. **b** The binding-sites between **8k** and TMV CP; **c** was a partial enlarged detail of (**b**), one binding-site, C=O-H-N = 3.24 Å, were found in the representative conformation. The stick model stands for **8k** and Arg90

Germany) to measure. Laser on time was set at 30 s, and laser-off time was set at 5 s. Using the mass action equation in the Nano Temper software, the K_d values were calculated from the duplicate reads, each experiments average of three replicates.

General synthesis procedure for intermediate 4

According to our previous synthesis procedure described in the literature [24], intermediates 4 were synthesized with minor modification. A solution of intermediate 3 (5.00 mmol), pyridine (13 mL) and hydroxylamine hydrochloride (1.73 mmol) in ethanol (25 mL) was stirred at



Scheme 1 Synthetic route to title compounds **8a–8p**

room temperature for 13 h. Then, a white solid appeared in the reaction mixture was filtered and washed with ethanol (50 mL) to obtain intermediates **4**. Physical properties and related NMR data of intermediates **4** are listed in Additional file 1.

General synthesis procedures for title compounds **8**

A solution of intermediates **4** (1.50 mmol), intermediates **7** (1.80 mmol) and K_2CO_3 (3.00 mmol) in acetonitrile (30 mL) was stirred under reflux for 4–5 h. Then, the mixture was filtered, the solvent was removed under vacuum, and the crude product was then separated by column chromatography with petroleum ether/ethyl acetate (V:V = 1:1) to obtain the title compounds **8a–8p**.

3-((((1*E*, 3*Z*, 4*E*)-1-(4-((3-chlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino

oxy)methyl)quinazolin-4(3*H*)-one (**8a**) White solid, m.p. 148–150 °C; yield: 36%; IR (KBr, cm^{-1}): 3063, 2924, 1749, 1728, 1627, 1600, 1506, 1224, 1172, 1022, 958; 1H NMR (500 MHz, $DMSO-d_6$) δ 8.56 (d, $J=4.0$ Hz, 1H, Py-6-H), 8.54–8.51 (m, 1H, Qu-2-H), 8.16 (dd, $J=8.0$, 1.3 Hz, 1H, Qu-5-H), 7.85–7.80 (m, 1H, Qu-7-H), 7.78 (td, $J=7.7$, 1.8 Hz, 1H, Ar(4-O)-2-H), 7.66 (d, $J=8.0$ Hz, 1H, Ar(4-O)-6-H), 7.61 (d, $J=7.8$ Hz, 1H, Qu-8-H), 7.59 (s, 1H, Qu-6-H), 7.57 (s, 1H, Py-2-H), 7.54 (t, $J=7.4$ Hz, 1H, Ar(3-Cl)-2-H), 7.49 (s, 1H, Py-4-H), 7.39 (d, $J=3.2$ Hz, 1H, Py-5-H), 7.39 (s, 1H, Ar(3-Cl)-5-H), 7.37 (d, $J=2.2$ Hz, 1H, Ar(3-Cl)-6-H), 7.35 (d, $J=4.1$ Hz, 1H, Ar(4-O)-3-H), 7.29 (dd, $J=7.4$, 5.3 Hz, 1H, Ar(4-O)-5-H), 7.20 (dd, $J=18.6$, 13.6 Hz, 2H, Ar-CH=), 7.10–7.05 (m, 1H, Py-C=CH), 7.03 (s, 1H, Py-CH=), 7.01 (s, 1H, Ar-C=CH), 5.99 (s, 2H, CH_2), 5.14 (s, 2H, CH_2); ^{13}C NMR (125 MHz, $DMSO-d_6$) δ 160.5, 159.7, 155.8,

154.3, 150.2, 148.6, 148.3, 139.9, 138.7, 137.5, 135.5, 135.1, 133.7, 130.9, 129.8, 129.0, 128.4, 127.9, 127.9, 126.9, 126.8, 125.3, 124.0, 123.8, 122.1, 115.7, 114.4, 77.8, 68.9; HRMS calcd for $C_{32}H_{25}ClN_4O_3$ $[M+H]^+$: 549.1688, found 549.1683.

3-((((1E,3Z,4E)-1-(4-((3-methylbenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8b) White solid, m.p. 155–157 °C; yield: 41%; IR (KBr, cm^{-1}): 2931, 1747, 1732, 1681, 1593, 1506, 1247, 1172, 1091, 1012, 748; 1H NMR (500 MHz, $CDCl_3$) δ 8.61 (d, $J=4.5$ Hz, 1H, Py-6-H), 8.39 (s, 1H, Qu-2-H), 8.34 (d, $J=7.9$ Hz, 1H, Qu-5-H), 7.78–7.73 (m, 1H, Qu-7-H), 7.72–7.65 (m, 2H, Ar(4-O)-2,6-2H), 7.52 (dd, $J=13.4, 7.4$ Hz, 2H, Ar(2-Cl)-3-H, Qu-8-H), 7.47 (d, $J=8.7$ Hz, 3H, Qu-6-H, Py-3,4-2H), 7.41 (d, $J=6.1$ Hz, 1H, Py-5-H), 7.38 (s, 1H, Ar(2-Cl)-6-H), 7.36 (d, $J=7.9$ Hz, 1H, Ar(2-Cl)-4-H), 7.28 (d, $J=6.2$ Hz, 1H, Ar(2-Cl)-5-H), 7.19 (t, $J=6.0$ Hz, 2H, Ar(4-O)-3,5-2H), 7.16 (s, 1H, Py-CH=), 7.09 (d, $J=16.6$ Hz, 1H, Py-C=CH), 6.96 (d, $J=8.5$ Hz, 2H, Ar-CH=, Ar-C=CH), 6.02 (s, 2H, CH_2), 5.18 (s, 2H, CH_2); ^{13}C NMR (125 MHz, $CDCl_3$) δ 161.0, 159.6, 156.2, 154.4, 149.9, 148.2, 146.9, 138.3, 136.8, 134.7, 134.4, 132.7, 129.5, 129.3, 129.2, 129.1, 128.9, 127.7, 127.4, 127.2, 127.1, 125.2, 123.3, 123.1, 122.3, 115.2, 114.8, 77.5, 67.2; HRMS calcd for $C_{32}H_{25}ClN_4O_3$ $[M+H]^+$: 529.2234, found 529.2233.

3-((((1E,3Z,4E)-1-(4-((2-chlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8c) White solid, m.p. 135–137 °C; yield: 33%; IR (KBr, cm^{-1}): 3061, 2927, 1732, 1716, 1600, 1506, 1454, 1284, 1234, 1074, 1028, 752; 1H NMR (500 MHz, $CDCl_3$) δ 8.62 (s, 1H, Py-6-H), 8.39 (d, $J=3.3$ Hz, 1H, Qu-2-H), 8.35 (dd, $J=7.9, 1.9$ Hz, 1H, Qu-5-H), 7.75 (dd, $J=10.9, 4.0$ Hz, 1H, Qu-7-H), 7.73–7.69 (m, 1H, Ar(4-O)-2-H), 7.67 (dd, $J=6.7, 2.5$ Hz, 1H, Ar(4-O)-6-H), 7.53–7.48 (m, 1H, Qu-8-H), 7.47 (d, $J=3.0$ Hz, 1H, Qu-6-H), 7.45 (d, $J=3.1$ Hz, 1H, Py-3-H), 7.40 (dd, $J=15.9, 3.4$ Hz, 1H, Ar(3- CH_3)-5-H), 7.36 (d, $J=7.7$ Hz, 1H, Py-5-H), 7.28 (dd, $J=7.4, 3.3$ Hz, 1H, Py-4-H), 7.25 (d, $J=3.6$ Hz, 2H, Ar(3- CH_3)-4,6-2H), 7.22 (d, $J=8.7$ Hz, 1H, Ar(3- CH_3)-2-H), 7.20 (d, $J=1.8$ Hz, 1H, Ar(4-O)-3-H), 7.16 (d, $J=3.3$ Hz, 1H, Ar(4-O)-5-H), 7.10 (dd, $J=17.2, 14.2$ Hz, 2H, Ar-CH=, Py-C=CH), 6.96 (d, $J=3.1$ Hz, 1H, Ar-C=CH), 6.94 (d, $J=3.1$ Hz, 1H, Py-CH=), 6.02 (d, $J=3.1$ Hz, 2H, CH_2), 5.04 (d, $J=2.6$ Hz, 2H, CH_2), 2.37 (d, $J=2.7$ Hz, 3H, CH_3); ^{13}C NMR (125 MHz, $CDCl_3$) δ 161.0, 160.0, 156.2, 154.4, 149.9, 148.2, 146.9, 138.5, 138.3, 136.8, 136.6, 134.7, 134.6, 129.2, 129.0, 128.8, 128.6, 128.4, 127.7, 127.4, 127.2, 125.1, 124.7, 123.3, 123.2, 122.3, 115.2, 114.6, 77.5, 70.2, 21.5; HRMS calcd for $C_{33}H_{28}N_4O_3$ $[M+H]^+$: 549.1687, found 549.1696.

3-((((1E,3Z,4E)-1-(4-((4-chlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8d) White solid, m.p. 147–149 °C; yield: 27%; IR (KBr, cm^{-1}): 3061, 2941, 1747, 1732, 1614, 1487, 1454, 1282, 1232, 1107, 1028, 916, 748; 1H NMR (500 MHz, $CDCl_3$) δ 8.61 (d, $J=4.2$ Hz, 1H, Py-6-H), 8.39 (s, 1H, Qu-2-H), 8.34 (d, $J=7.1$ Hz, 1H, Qu-5-H), 7.75 (dd, $J=11.0, 4.3$ Hz, 1H, Qu-7-H), 7.71 (s, 1H, Ar(4-O)-2-H), 7.70–7.68 (m, 1H, Ar(4-O)-6-H), 7.68–7.65 (m, 1H, Qu-8-H), 7.50 (t, $J=7.5$ Hz, 1H, Qu-6-H), 7.46 (d, $J=8.7$ Hz, 2H, Py-3,4-2H), 7.40 (d, $J=15.8$ Hz, 1H, Py-5-H), 7.37 (s, 1H, Ar(4-Cl)-3-H), 7.25 (s, 1H, Ar(4-Cl)-5-H), 7.21 (d, $J=4.8$ Hz, 1H, Ar(4-Cl)-2-H), 7.19 (s, 1H, Ar(4-Cl)-6-H), 7.19–7.18 (m, 1H, Py-C=CH), 7.16 (s, 1H, Ar(4-O)-3-H), 7.16 (s, 1H, Ar(4-O)-5-H), 7.08 (d, $J=16.6$ Hz, 1H, Ar-CH=), 6.93 (d, $J=8.7$ Hz, 2H, Py-CH=, Ar-C=CH), 6.02 (s, 2H, CH_2), 5.02 (d, $J=16.2$ Hz, 2H, CH_2); ^{13}C NMR (125 MHz, $CDCl_3$) δ 161.1, 159.6, 156.1, 154.4, 149.9, 148.2, 146.9, 138.2, 136.8, 135.2, 134.7, 134.0, 129.3, 129.1, 128.9, 128.9, 127.7, 127.4, 127.2, 125.1, 123.3, 123.2, 122.3, 115.2, 114.8, 77.5, 69.3; HRMS calcd for $C_{32}H_{25}ClN_4O_3$ $[M+H]^+$: 549.1687, found 549.1691.

3-((((1E,3Z,4E)-1-(4-((2,4-dichlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8e) White solid, m.p. 174–176 °C; yield: 24%; IR (KBr, cm^{-1}): 3065, 2871, 1734, 1716, 1602, 1521, 1489, 1456, 1344, 1249, 1172, 1062, 1012, 932, 752; 1H NMR (500 MHz, $CDCl_3$) δ 8.61 (d, $J=4.1$ Hz, 1H, Py-6-H), 8.39 (s, 1H, Qu-2-H), 8.34 (dd, $J=8.0, 1.1$ Hz, 1H, Qu-5-H), 7.78–7.73 (m, 1H, Qu-7-H), 7.72–7.65 (m, 2H, Ar(4-O)-2,6-2H), 7.52–7.49 (m, 1H, Ar(2,4-2Cl)-3-H), 7.49–7.45 (m, 3H, Qu-6,8-2H, Py-3-H), 7.41 (t, $J=9.1$ Hz, 2H, Py-4,5-2H), 7.36 (d, $J=7.9$ Hz, 1H, Ar(2,4-2Cl)-5-H), 7.27 (dd, $J=8.3, 2.0$ Hz, 1H, Ar(2,4-2Cl)-6-H), 7.22–7.15 (m, 3H, Py-CH=, Ar(4-O)-3,5-2H), 7.08 (d, $J=16.7$ Hz, 1H, Ar-C=CH), 6.95 (t, $J=5.7$ Hz, 2H, Ar-CH=, Py-C=CH), 6.02 (s, 2H, CH_2), 5.13 (s, 2H, CH_2); ^{13}C NMR (125 MHz, $CDCl_3$) δ 161.0, 159.3, 156.1, 154.4, 149.9, 148.2, 146.9, 138.1, 136.8, 134.7, 134.3, 133.3, 133.1, 129.7, 129.4, 129.3, 127.7, 127.4, 127.2, 125.1, 123.3, 123.2, 122.3, 115.2, 115.0, 77.5, 66.7. HRMS calcd for $C_{32}H_{25}ClN_4O_3$ $[M+H]^+$: 583.1298, found 583.1297.

6-chloro-3-((((1E,3Z,4E)-1-(4-((3-methylbenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8f) White solid, m.p. 168–170 °C; yield: 34%; IR (KBr, cm^{-1}): 1741, 1600, 1506, 1247, 1172, 1089, 1066, 1012, 960, 821, 748; 1H NMR (500 MHz, CD_3COCD_3) δ 8.61 (d, $J=4.6$ Hz, 1H, Py-6-H), 8.37 (s, 1H, Qu-2-H), 8.30 (d, $J=2.3$ Hz, 1H,

Qu-5-H), 7.70–7.63 (m, 3H, Ar(4-O)-2,6-2H, Qu-7-H), 7.46 (t, $J=5.7$ Hz, 2H, Ar(3-CH₃)-5-H, Py-3-H), 7.39 (d, $J=15.7$ Hz, 1H, Py-4-H), 7.35 (d, $J=7.9$ Hz, 1H, Py-5-H), 7.28 (d, $J=7.5$ Hz, 1H, Ar(3-CH₃)-6-H), 7.24 (s, 1H, Qu-8-H), 7.21 (dd, $J=7.8, 4.1$ Hz, 2H, Py-C=CH, Ar(3-CH₃)-4-H), 7.19–7.12 (m, 3H, Ar(3-CH₃)-2-H, Ar(4-O)-3,5-2H), 7.08 (d, $J=16.6$ Hz, 1H, Ar-CH=), 6.95 (d, $J=8.8$ Hz, 2H, Py-CH=, Ar-C=CH), 6.00 (s, 2H, CH₂), 5.04 (s, 2H, CH₂), 2.37 (s, 3H, CH₃); ¹³C NMR (125 MHz, CD₃COCD₃) δ 160.1, 160.0, 156.4, 154.4, 149.9, 147.0, 146.7, 138.5, 138.5, 136.8, 136.6, 135.1, 134.8, 133.3, 129.4, 129.2, 129.0, 128.7, 128.6, 128.3, 126.6, 125.1, 124.7, 123.4, 123.3, 123.2, 115.2, 114.5, 77.6, 70.2, 21.5; HRMS calcd for C₃₃H₂₇ClN₄O₃ [M+H]⁺: 563.1844, found 563.1844.

6-Chloro-3-((((1E,3Z,4E)-1-(4-((3-chlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8g) White solid, m.p. 126–128 °C; yield: 26%; IR (KBr, cm⁻¹): 3050, 2954, 1734, 1610, 1508, 1487, 1456, 1284, 1234, 1174, 1109, 1029, 916, 823, 752; ¹H NMR (500 MHz, CDCl₃) δ 8.61 (d, $J=3.9$ Hz, 1H, Py-6-H), 8.37 (s, 1H, Qu-2-H), 8.29 (d, $J=2.3$ Hz, 1H, Qu-5-H), 7.70–7.67 (m, 2H, Ar(4-O)-2,6-2H), 7.65 (dd, $J=9.8, 5.3$ Hz, 1H, Qu-7-H), 7.46 (d, $J=8.8$ Hz, 2H, Py-3-H, Ar(3-Cl)-2-H), 7.43 (s, 1H, Py-4-H), 7.39 (d, $J=15.9$ Hz, 1H, Py-5-H), 7.35 (d, $J=7.8$ Hz, 1H, Ar(3-Cl)-5-H), 7.30 (t, $J=2.5$ Hz, 3H, Ar(3-Cl)-4,6-2H, Qu-8-H), 7.22–7.19 (m, 1H, Py-C=CH), 7.16 (dd, $J=16.2, 4.4$ Hz, 2H, Ar(4-O)-3,5-2H), 7.08 (d, $J=16.7$ Hz, 1H, Ar-CH=), 6.96–6.91 (m, 2H, Ar-C=CH, Py-CH=), 6.00 (s, 2H, CH₂), 5.05 (s, 2H, CH₂); ¹³C NMR (125 MHz, CDCl₃) δ 160.0, 159.6, 156.4, 154.4, 149.9, 147.0, 146.7, 138.8, 138.4, 136.8, 135.1, 134.9, 134.7, 133.3, 130.0, 129.4, 129.3, 129.0, 128.3, 127.5, 126.6, 125.5, 125.1, 123.4, 123.3, 123.2, 115.2, 114.7, 77.6, 69.3; HRMS calcd for C₃₂H₂₅Cl₂N₄O₃ [M+H]⁺: 583.1298, found 583.1298.

6-Chloro-3-((((1E,3Z,4E)-1-(4-((2-chlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8h) White solid, m.p. 214–216 °C; yield: 21%; IR (KBr, cm⁻¹): 2922, 2852, 1747, 1597, 1521, 1452, 1346, 1244, 1058, 923, 837, 736, 700; ¹H NMR (500 MHz, CDCl₃) δ 8.62 (d, $J=3.7$ Hz, 1H, Py-6-H), 8.37 (s, 1H, Qu-2-H), 8.30 (d, $J=2.3$ Hz, 1H, Qu-5-H), 7.70–7.63 (m, 3H, Ar(4-O)-2,6-2H, Ar(2-Cl)-3-H), 7.53 (dd, $J=7.1, 2.1$ Hz, 1H, Qu-7-H), 7.47 (d, $J=8.7$ Hz, 2H, Py-3,4-2H), 7.42–7.37 (m, 2H, Py-5-H, Ar(2-Cl)-6-H), 7.35 (d, $J=7.8$ Hz, 1H, Qu-8-H), 7.31–7.26 (m, 2H, Ar(2-Cl)-4,5-2H), 7.21 (dd, $J=6.2, 1.9$ Hz, 1H, Py-C=CH), 7.17 (dd, $J=16.3, 3.6$ Hz, 2H, Ar(4-O)-3,5-2H),

7.09 (d, $J=16.6$ Hz, 1H, Ar-CH=), 6.97 (d, $J=8.7$ Hz, 2H, Ar-C=CH, Py-CH=), 6.00 (s, 2H, CH₂), 5.19 (s, 2H, CH₂); ¹³C NMR (125 MHz, CDCl₃) δ 160.0, 159.7, 156.4, 154.4, 149.9, 147.0, 146.7, 138.4, 136.8, 135.1, 134.9, 134.4, 133.3, 132.7, 129.5, 129.4, 129.3, 129.2, 129.0, 128.9, 127.1, 126.6, 125.1, 123.4, 123.3, 123.2, 115.2, 114.7, 77.6, 67.3; HRMS calcd for C₃₂H₂₄Cl₂N₄O₃ [M+H]⁺: 583.1298, found 583.1306.

6-Chloro-3-((((1E,3Z,4E)-1-(4-((4-chlorobenzyl)oxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8i) White solid, m.p. 172–174 °C; yield: 18%; IR (KBr, cm⁻¹): 1718, 1633, 1598, 1568, 1448, 1238, 1170, 1064, 1012, 914, 823, 713; ¹H NMR (500 MHz, CDCl₃) δ 8.61 (d, $J=4.0$ Hz, 1H, Py-6-H), 8.37 (s, 1H, Qu-2-H), 8.29 (d, $J=2.3$ Hz, 1H, Qu-5-H), 7.69 (dd, $J=5.0, 2.8$ Hz, 1H, Ar(4-O)-2-H), 7.67 (d, $J=2.4$ Hz, 1H, Ar(4-O)-6-H), 7.65 (dd, $J=10.3, 5.2$ Hz, 1H, Qu-7-H), 7.48–7.44 (m, 2H, Py-3,4-2H), 7.41–7.37 (m, 1H, Py-5-H), 7.37–7.34 (m, 5H, Ar(4-Cl)-2,3,4,5-4H, Qu-8-H), 7.22–7.16 (m, 2H, Ar(4-O)-3,5-2H), 7.15 (d, $J=5.5$ Hz, 1H, Ar-CH=), 7.08 (d, $J=16.7$ Hz, 1H, Py-C=CH), 6.95–6.91 (m, 2H, Ar-C=CH, Py-CH=), 6.00 (s, 2H, CH₂), 5.04 (s, 2H, CH₂); ¹³C NMR (125 MHz, CDCl₃) δ 160.0, 159.7, 156.4, 154.4, 149.9, 147.0, 146.7, 138.4, 136.8, 135.2, 135.1, 134.8, 134.0, 133.3, 129.4, 129.3, 129.0, 128.9, 128.8, 126.6, 125.1, 123.4, 123.3, 123.2, 115.2, 114.7, 77.6, 69.3; HRMS calcd for C₃₂H₂₅Cl₂N₄O₃ [M+H]⁺: 583.1225, found 583.1359.

3-((((1E,3Z,4E)-1-(2-(2-chlorophenoxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxymethyl)quinazolin-4(3H)-one (8j) White solid, m.p. 95–97 °C; yield: 36%; IR (KBr, cm⁻¹): 2924, 1734, 1716, 1583, 1558, 1489, 1247, 1172, 1076, 1012, 748; ¹H NMR (500 MHz, CDCl₃) δ 8.60 (d, $J=4.4$ Hz, 1H, Py-6-H), 8.38 (s, 1H, Qu-2-H), 8.34 (d, $J=8.0$ Hz, 1H, Qu-5-H), 7.75 (dd, $J=11.0, 4.1$ Hz, 1H, Qu-7-H), 7.70 (d, $J=8.0$ Hz, 1H, Ar(2-Cl)-3-H), 7.67–7.60 (m, 3H, Ar(2-O)-6-H, Qu-6,8-2H), 7.52–7.46 (m, 2H, Py-3,4-2H), 7.43 (d, $J=15.9$ Hz, 1H, Py-5-H), 7.32 (d, $J=8.8$ Hz, 1H, Ar(2-Cl)-6-H), 7.31–7.24 (m, 3H, Ar(2-Cl)-4,5-2H, Ar(2-O)-4-H), 7.22–7.14 (m, 3H, Py-C=CH, Ar-CH=, Ar(2-O)-5-H), 7.08 (t, $J=7.5$ Hz, 1H, Ar-C=CH), 7.00 (t, $J=7.5$ Hz, 1H, Ar(2-O)-3-H), 6.93 (d, $J=8.2$ Hz, 1H, Py-CH=), 6.01 (s, 2H, CH₂), 5.19 (s, 2H, CH₂); ¹³C NMR (125 MHz, CDCl₃) δ 161.0, 156.8, 156.5, 154.6, 149.9, 148.2, 146.9, 136.6, 135.1, 134.7, 134.5, 133.7, 132.5, 130.7, 129.4, 128.9, 128.6, 127.7, 127.7, 127.4, 127.2, 127.1, 125.6, 125.4, 123.1, 123.0, 122.3, 121.5, 117.1, 112.7, 77.5, 67.6; HRMS calcd for C₃₂H₂₅ClN₄O₃ [M+H]⁺: 549.1687, found 549.1684.

3-((((1E,3Z,4E)-1-(2-(2,4-dichlorophenoxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxy)methylquinazolin-4(3H)-one (**8k**) White solid, m.p. 194–196 °C; yield: 44%; IR (KBr, cm^{-1}): 3042, 2923, 1734, 1608, 1508, 1454, 1282, 1234, 1172, 1105, 1031, 918, 748; ^1H NMR (500 MHz, CDCl_3) δ 8.63–8.59 (m, 1H, Py-6-H), 8.38 (s, 1H, Qu-2-H), 8.34 (dd, $J=8.0, 1.2$ Hz, 1H, Qu-5-H), 7.76 (m, 1H, Qu-7-H), 7.72–7.69 (m, 1H, Ar(2-O)-6-H), 7.69–7.63 (m, 2H, Ar(2,4-2Cl)-3-H, Qu-8-H), 7.61–7.56 (m, 1H, Qu-6-H), 7.50 (ddd, $J=8.1, 7.1, 1.2$ Hz, 1H, Py-3-H), 7.45–7.39 (m, 2H, Py-4,5-2H), 7.35 (d, $J=2.1$ Hz, 1H, Ar(2-O)-4-H), 7.29 (dd, $J=12.4, 4.4$ Hz, 3H, Ar(2,4-2Cl)-5,6-2H, Ar-CH=), 7.22 (m, 1H, Ar(2-O)-5-H), 7.18 (d, $J=15.9$ Hz, 1H, Py-C=CH), 7.05 (dd, $J=8.3, 2.1$ Hz, 1H, Ar-C=CH), 7.01 (t, $J=7.5$ Hz, 1H, Ar(2-O)-3-H), 6.91 (d, $J=8.2$ Hz, 1H, Py-CH=), 6.02 (s, 2H, CH_2), 5.14 (s, 2H, CH_2); ^{13}C NMR (125 MHz, CDCl_3) δ 161.0, 156.7, 156.2, 154.5, 149.9, 148.2, 146.9, 136.7, 135.2, 134.7, 134.1, 133.5, 133.2, 133.1, 130.7, 129.6, 129.2, 127.7, 127.7, 127.5, 127.4, 127.2, 125.5, 125.4, 123.2, 123.1, 122.3, 121.7, 117.1, 112.7, 77.5, 67.1; HRMS calcd for $\text{C}_{32}\text{H}_{25}\text{Cl}_2\text{N}_4\text{O}_3$ $[\text{M}+\text{H}]^+$: 583.1298, found 583.1300.

6-Chloro-3-((((1E,3Z,4E)-1-(2-(2-chlorophenoxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxy)methylquinazolin-4(3H)-one (**8l**) White solid, m.p. 153–155 °C; yield: 23%; IR (KBr, cm^{-1}): 2940, 2830, 1734, 1716, 1600, 1519, 1489, 1456, 1346, 1284, 1259, 1107, 1083, 1029, 802, 736; ^1H NMR (500 MHz, CDCl_3) δ 8.61 (d, $J=4.4$ Hz, 1H, Py-6-H), 8.36 (s, 1H, Qu-2-H), 8.29 (d, $J=2.2$ Hz, 1H, Qu-5-H), 7.67 (dt, $J=4.5, 2.2$ Hz, 1H, Ar(2-Cl)-3-H), 7.65 (s, 2H, Ar(2-O)-6-H, Qu-7-H), 7.63 (s, 1H, Py-3-H), 7.60 (s, 1H, Py-4-H), 7.48 (d, $J=7.5$ Hz, 1H, Py-5-H), 7.42 (d, $J=15.9$ Hz, 1H, Ar(2-Cl)-6-H), 7.33 (d, $J=7.9$ Hz, 1H, Qu-8-H), 7.30 (dd, $J=6.0, 3.5$ Hz, 2H, Ar(2-Cl)-5-H, Ar(2-Cl)-4-H), 7.27 (d, $J=3.6$ Hz, 1H, Ar(2-O)-4-H), 7.21–7.15 (m, 3H, Ar-CH=, Ar(2-O)-5-H, Ar(2-O)-3-H), 7.08 (t, $J=7.3$ Hz, 1H, Py-C=CH), 7.00 (t, $J=7.5$ Hz, 1H, Ar-C=CH), 6.94 (d, $J=8.3$ Hz, 1H, Py-CH=), 5.99 (s, 2H, CH_2), 5.19 (s, 2H, CH_2); ^{13}C NMR (125 MHz, CDCl_3) δ 160.0, 157.0, 156.5, 154.5, 149.9, 147.0, 146.7, 136.7, 135.3, 135.1, 134.5, 133.9, 133.3, 132.5, 130.8, 129.4, 129.3, 129.0, 128.6, 127.7, 127.1, 126.6, 125.5, 125.3, 123.4, 123.1, 123.1, 121.5, 117.0, 112.8, 77.5, 67.7; HRMS calcd for $\text{C}_{32}\text{H}_{25}\text{Cl}_2\text{N}_4\text{O}_3$ $[\text{M}+\text{H}]^+$: 583.1298, found 583.1285.

6-Chloro-3-((((1E,3Z,4E)-1-(2-(2,4-dichlorophenoxy)phenyl)-5-(pyridin-2-yl)penta-1,4-dien-3-ylidene)amino)oxy)methylquinazolin-4(3H)-one (**8m**) White solid, m.p. 113–115 °C; yield: 32%; IR (KBr, cm^{-1}): 2927, 1747, 1519, 1456, 1346, 1261, 1091, 1076, 1029, 800; ^1H NMR

(500 MHz, CDCl_3) δ 8.61 (d, $J=3.9$ Hz, 1H, Py-6-H), 8.37 (d, $J=6.1$ Hz, 1H, Qu-2-H), 8.29 (t, $J=2.3$ Hz, 1H, Qu-5-H), 7.68 (m, 2H, Ar(2,4-2Cl)-3-H, Ar(2-O)-6-H), 7.65 (d, $J=8.6$ Hz, 1H, Qu-7-H), 7.47 (dt, $J=4.8, 2.1$ Hz, 3H, Py-3,4,5-3H), 7.40 (dd, $J=14.7, 9.0$ Hz, 2H, Ar(2,4-2Cl)-5,6-2H), 7.36 (d, $J=7.8$ Hz, 1H, Qu-8-H), 7.27 (dd, $J=8.3, 2.1$ Hz, 1H, Ar(2-O)-4-H), 7.22–7.20 (m, 1H, Ar-CH=), 7.19 (t, $J=2.5$ Hz, 1H, Ar(2-O)-5-H), 7.15 (d, $J=3.7$ Hz, 1H, Ar(2-O)-3-H), 7.09 (d, $J=16.7$ Hz, 1H, Py-C=CH), 6.97–6.93 (m, 2H, Py-CH=, Ar-C=CH), 6.00 (s, 2H, CH_2), 5.13 (s, 2H, CH_2); ^{13}C NMR (125 MHz, CDCl_3) δ 160.0, 159.4, 156.3, 154.4, 149.9, 147.0, 146.7, 138.3, 136.8, 135.1, 134.9, 134.4, 133.3, 133.3, 133.1, 129.7, 129.4, 129.3, 129.2, 127.5, 126.6, 125.1, 123.4, 123.3, 123.2, 115.2, 114.9, 77.6, 66.7; HRMS calcd for $\text{C}_{32}\text{H}_{24}\text{Cl}_3\text{N}_4\text{O}_3$ $[\text{M}+\text{H}]^+$: 617.0908, found 617.0909.

3-((((1E,3Z,4E)-1-(4-((2,4-dichlorobenzyl)oxy)phenyl)-5-(thiophen-2-yl)penta-1,4-dien-3-ylidene)amino)oxy)methylquinazolin-4(3H)-one (**8n**) White solid, m.p. 84–86 °C; yield: 23%; IR (KBr, cm^{-1}): 3062, 2944, 1716, 1600, 1558, 1503, 1456, 1284, 1228, 1174, 750; ^1H NMR (500 MHz, CD_3COCD_3) δ 8.46 (s, 1H, Qu-2-H), 8.22 (dd, $J=8.0, 1.4$ Hz, 1H, Qu-5-H), 7.82–7.77 (m, 1H, Qu-7-H), 7.64 (dd, $J=14.9, 8.2$ Hz, 2H, Ar(2,4-2Cl)-3-H, Qu-8-H), 7.60–7.57 (m, 2H, Ar(4-O)-2,6-2H), 7.53 (dd, $J=13.4, 4.5$ Hz, 2H, Qu-6-H, Thiophene-5-H), 7.45–7.36 (m, 3H, Ar(2,4-2Cl)-5,6-2H, Thiophene-3-H), 7.25 (d, $J=3.4$ Hz, 1H, Thiophene-4-H), 7.19 (q, $J=16.8$ Hz, 2H, Ar(4-O)-3,5-2H), 7.09–7.02 (m, 3H, Ar-CH=, Thiophene-CH=, Thiophene-C=CH), 6.72 (d, $J=16.0$ Hz, 1H, Ar-C=CH), 6.04 (s, 2H, CH_2), 5.20 (s, 2H, CH_2); ^{13}C NMR (125 MHz, CD_3COCD_3) δ 161.0, 159.3, 156.4, 148.2, 146.9, 141.6, 137.9, 134.8, 134.4, 133.3, 133.1, 129.7, 129.4, 129.2, 129.0, 128.3, 127.9, 127.7, 127.5, 127.2, 126.2, 122.3, 120.5, 115.2, 115.2, 77.2, 66.7; HRMS calcd for $\text{C}_{31}\text{H}_{23}\text{Cl}_2\text{N}_3\text{O}_3\text{S}$ $[\text{M}+\text{H}]^+$: 588.0909, found 588.0907.

3-((((1E,3Z,4E)-1-(4-((2-chlorobenzyl)oxy)phenyl)-5-(pyridin-3-yl)penta-1,4-dien-3-ylidene)amino)oxy)methylquinazolin-4(3H)-one (**8o**) White solid, m.p. 94–96 °C; yield: 25%; IR (KBr, cm^{-1}): 2924, 1741, 1683, 1591, 1282, 1251, 1091, 1078, 1014, 918, 852, 761; ^1H NMR (500 MHz, CDCl_3) δ 8.83 (s, 1H, Py-2-H), 8.63 (d, $J=4.6$ Hz, 1H, Py-6-H), 8.57 (s, 1H, Qu-2-H), 8.39 (d, $J=7.9$ Hz, 1H, Qu-5-H), 8.07 (d, $J=7.9$ Hz, 1H, Py-4-H), 8.00 (s, 1H, Qu-7-H), 7.92 (t, $J=7.5$ Hz, 1H, Ar(4-O)-2-H), 7.81 (d, $J=8.1$ Hz, 1H, Ar(4-O)-6-H), 7.70–7.61 (m, 5H, Ar(2-Cl)-3,6-2H, Qu-6,8-2H, Py-5-H), 7.57–7.52 (m, 1H, Ar(2-Cl)-4-H), 7.49–7.42 (m, 3H, Ar(2-Cl)-5-H, Ar(4-O)-3,5-2H), 7.29 (s, 1H, Py-C=CH), 7.17–7.09 (m, 3H, Ar-CH=, Py-CH=, Ar-C=CH), 6.15 (s, 2H, CH_2),

5.31 (s, 2H, CH₂); ¹³C NMR (125 MHz, CD₃COCD₃) δ 166.0, 160.8, 160.6, 154.5, 154.1, 151.6, 150.0, 140.5, 138.1, 137.1, 137.0, 136.8, 134.0, 129.7, 128.6, 128.6, 128.5, 128.3, 127.9, 127.7, 124.8, 124.1, 123.8, 123.6, 123.3, 115.3, 115.1, 114.1, 78.4, 69.8; HRMS calcd for C₃₂H₂₅ClN₄O₃ [M+H]⁺: 549.1687, found 549.1691.

3-((((1*E*, 3*Z*, 4*E*)-1-(4-((3-*methylbenzyl*)oxy)phenyl)-5-(pyridin-3-yl)penta-1,4-dien-3-ylidene)amino)oxymethylquinazolin-4(3*H*)-one (**8p**) White solid, m.p. 154–156 °C; yield: 29%; IR (KBr, cm⁻¹): 3065, 2947, 1741, 1732, 1600, 1506, 1456, 1240, 1172, 1053, 1022, 918, 825, 698; ¹H NMR (500 MHz, CDCl₃) δ 8.64–8.60 (m, 1H, Py-2-H), 8.41 (s, 1H, Py-6-H), 8.20 (dd, *J*=7.9, 0.6 Hz, 1H, Qu-2-H), 7.68 (td, *J*=7.7, 1.8 Hz, 1H, Qu-5-H), 7.62–7.58 (m, 1H, Py-4-H), 7.46 (d, *J*=8.7 Hz, 2H, Ar(4-O)-2,6-2H), 7.43–7.34 (m, 3H, Qu-6,7,8-3H), 7.27 (t, *J*=7.5 Hz, 1H, Ar(3-CH₃)-5-H), 7.25–7.19 (m, 4H, Ar(3-CH₃)-2,4,6-3H, Py-5-H), 7.19–7.17 (m, 1H, Ar-CH=), 7.17–7.12 (m, 2H, Ar(4-O)-3,5-2H), 7.08 (d, *J*=16.7 Hz, 1H, Py-C=CH), 6.95 (d, *J*=8.8 Hz, 2H, Py-CH=, Ar-C=CH), 6.02 (s, 2H, CH₂), 5.04 (s, 2H, CH₂), 2.37 (s, 3H, CH₃); ¹³C NMR (125 MHz, CDCl₃) δ 161.4, 160.0, 156.1, 154.5, 149.9, 146.8, 145.6, 138.5, 138.3, 136.8, 136.6, 136.1, 135.4, 134.7, 129.2, 129.0, 128.8, 128.6, 128.4, 127.0, 125.3, 124.9, 124.7, 123.2, 123.1, 122.3, 115.2, 114., 77.5, 70.2, 17.6; HRMS calcd for C₃₃H₂₈N₄O₃ [M+H]⁺: 529.2234, found 529.2235.

Conclusions

In summary, aiming to develop novel, highly-efficient and environmentally benign agrochemicals, we introduced a quinazolin-4(3*H*)-one scaffold into penta-1,4-diene-3-one oxime ether to synthesis 16 curcumin derivatives, and their antiviral activity against TMV *in vivo* were evaluated. Antiviral bioassays showed that some compounds exhibited remarkable antiviral activity against TMV. In particular, compounds **8c**, **8j** and **8k** possessed appreciable curative activities against TMV *in vivo*, with the EC₅₀ values of 138.5, 132.9 and 125.6 µg/mL, respectively, which are better than ningnanmycin (207.3 µg/mL). Further studies on the MST and molecular docking experiments of **8k** interaction with TMV CP showed that **8k** bound Arg90 in the TMV CP protein. Arg90 was an important residue to bind TMV RNA. Thus, we speculate that **8k** inhibited the virulence of TMV by binding Arg90 in TMV CP. Given the above results, this kind of penta-1,4-diene-3-one oxime ether derivatives containing a quinazolin-4(3*H*)-one scaffold could be further studied as potential alternative templates in the search for novel antiviral agents.

Additional file

Additional file 1. The data of intermediates **1**, **2**, **3**, **4**, **6**, **7**; all the copies of ¹H NMR, ¹³C NMR, ³¹P NMR and HRMS for the title compounds; activity confirmed of compounds **8k** were presented in additional information.

Abbreviations

TMV: tobacco mosaic virus; MST: microscale thermophoresis experiments; TMV CP: TMV coat protein; IR: infrared spectrum; ¹H NMR: ¹H nuclear magnetic resonance; ¹³C NMR: ¹³C nuclear magnetic resonance; HRMS: high resolution mass spectrum.

Authors' contributions

The current study is an outcome of constructive discussion with WX. LC, XW, CZ and XT carry out their synthesis and characterization experiments; LC, XW, RX, TG and XL performed the antiviral and antibacterial activities; XW, LC, CZ, RX and XT carried out the ¹H NMR, ¹³C NMR, IR and HRMS spectral analyses; LC and XW were involved in the drafting of the manuscript and revising the manuscript. All authors read and approved the final manuscript.

Author details

¹ State Key Laboratory Breeding Base of Green Pesticide and Agricultural Bioengineering, Key Laboratory of Green Pesticide and Agricultural Bioengineering, Ministry of Education, Guizhou University, Guiyang 550025, China. ² College of Sciences, Nanjing Agricultural University, Nanjing 210095, China.

Acknowledgements

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

We have presented all our main data in the form of tables and figures. Meanwhile, all the copies of IR, ¹H NMR, ¹³C NMR and HRMS for the title compounds were presented in the Support Information. The datasets supporting the conclusions of the article are included within the article and its additional file.

Funding

The authors gratefully acknowledge grants from the National Key Research and Development Program of China (No. 2017YFD0200506), the National Nature Science Foundation of China (Nos. 21462012 and 21502032) and the Special Fund for Outstanding Scientific and Technological Candidates of Guizhou Province (No. 201535).

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 27 October 2018 Accepted: 2 March 2019

Published online: 25 March 2019

References

- Bos L (2000) 100 years of virology: from vitalism via molecular biology to genetic engineering. Trends Microbiol 8:82–87
- Reichman M, Devash Y, Suhadolnik RJ, Sela I (1983) Human leukocyte interferon and the antiviral factor (AVF) from virus-infected plants stimulate plant tissues to produce nucleotides with antiviral activity. Virology 128:240–244
- Li TX, Zhang J, Pan JK, Wu ZX, Hu DY, Song BA (2017) Design, synthesis, and antiviral activities of 1,5-benzothiazepine derivatives containing pyridine moiety. Eur J Med Chem 125:657–662
- Chen MH, Chen Z, Song BA, Bhadury PS, Yang S, Cai XJ, Hu DY, Xue W, Zeng S (2009) Synthesis and antiviral activities of chiral thiourea derivatives containing an a-aminophosphonate moiety. J Agric Food Chem 57:1383–1388

5. Gan XH, Hu DY, Wang YJ, Yu L, Song BA (2017) Novel trans-ferulic acid-derivatives containing a chalcone moiety as potential activator for plant resistance induction. *J Agric Food Chem* 65:4367–4377
6. Wang ZW, Wang L, Ma S, Liu YX, Wang LZ, Wang QM (2012) Design, synthesis, antiviral activity, and SARs of 14-aminophenanthroindolizidines. *J Agric Food Chem* 60:5825–5831
7. Qian XH, Lee PW, Cao S (2010) China: forward to the green pesticides via a basic research program. *J Agric Food Chem* 58:2613–2623
8. Leonard GC, Stephen OD (2007) Natural products that have been used commercially as crop protection agents. *Pest Manag Sci* 63:524–554
9. Venkateshwarlu R, Chinnababu B, Ramulu U, Reddy KP, Reddy MD, Sowjanya P, Rao PV, Aravind S (2017) Synthesis and biological evaluation of (–)—kunstleramide and its derivatives. *Med Chem Commun* 8:394–404
10. Reddy MD, Kobori H, Mori T, Wu J, Kawagishi H, Watkins EB (2017) Gram-scale, stereoselective synthesis and biological evaluation of (+)—armilariol C. *J Nat Prod* 80:2561–2565
11. Luo H, Yang SJ, Cai YQ, Peng ZJ, Liu T (2014) Synthesis and biological evaluation of novel 6-chloro-quinazolin derivatives as potential antitumor agents. *Eur J Med Chem* 84:746–752
12. Liu ZG, Tang LG, Zou P, Zhang YL, Wang Z, Fang QL, Jiang LL, Chen GZ, Xu Z, Zhang HJ, Liang G (2014) Synthesis and biological evaluation of allylated and prenylated mon-carbonyl analogs of curcumin as anti-inflammatory agents. *Eur J Med Chem* 74:671–682
13. Quincoces Suarez JA, Rando DG, Santos RP, Goncalves RP, Ferreira E, Carvalho JED, Kohn L, Maria DA, Faiao-Flores F, Michalik D, Marcucci MC, Vogel C (2010) New antitumoral agents I: in vitro anticancer activity and in vivo acute toxicity of synthetic 1,5-bis(4-hydroxy-3-methoxyphenyl)-1,4-pentadien-3-one and derivatives. *Bioorg Med Chem* 18:6275–6281
14. Li SB, Hu DY, Song BA, Yang S, Jin LH, Xue W, Zeng S, Wang J, Chen Z, Lu P, Zhou X, Fan LE (2008) Synthesis and antifungal activity of novel 1,5-diphenyl-1,4-pentadien-3-one oxime esters. *Chin J Org Chem* 28:311–316
15. Xia LJ, Chen LJ, Wang XB, Du XJ, Zheng ZY, Xue W (2017) Synthesis and biological activity of penta-1,4-diene-3-one oxime ester derivatives. *Fine Chem* 34:834–840
16. Wang ZN, Hu DY, Song BA, Yang S, Jin LH, Xue W (2009) Synthesis and biological activity of 1,5-bis(substituted pyrazol-4-yl)-1,4-pentadien-3-one derivatives. *Chin J Org Chem* 29:1412–1418
17. Luo H, Liu JJ, Jin LH, Hu DY, Chen Z, Yang S, Wu J, Song BA (2013) Synthesis and antiviral bioactivity of novel (1E,4E)-1-aryl-5-(2-(quinazolin-4-yloxy)phenyl)-1,4-pentadien-3-one derivatives. *Eur J Med Chem* 3:662–669
18. Ma J, Li P, Li XY, Shi Q, Wan Z, Hu DY, Jin LH, Song BA (2014) Synthesis and antiviral bioactivity of novel 3-((2-((1E,4E)-3-Oxo-5-aryl-penta-1,4-dien-1-yl)phenoxy)-methyl)-4(3H)-quinazolinone derivatives. *J Agric Food Chem* 62:8928–8934
19. Yu L, Gan XH, Zhou DG, He F, Zeng S, Hu DY (2017) 1,4-Pentadien-3-one derivatives containing a 1,3,4-thiadiazole moiety. *Molecules* 22:658
20. Gan XH, Hu DY, Li P, Wu J, Chen XW, Xue W, Song BA (2016) Design, synthesis, antiviral activity and three-dimensional quantitative structure-activity relationship study of novel 1,4-pentadien-3-one derivatives containing the 1,3,4-oxadiazole moiety. *Pest Manag Sci* 72:534–543
21. Wu J, Zhu YY, Zhao YH, Shan WL, Hu DY, Chen JX, Liu DY, Li XY, Yang S (2016) Synthesis and antiviral activities of novel 1,4-pentadien-3-one derivatives bearing an emodin moiety. *Chin Chem Lett* 27:948–952
22. Chen MH, Hu DY, Li XY, Yang S, Zhang WY, Li P, Song BA (2015) Antiviral activity and interaction mechanisms study of novel glucopyranoside derivatives. *Bioorg Med Chem Lett* 25:3840–3844
23. Chen Y, Wang ZB, Zhang X, Xia LJ, Gong HY, Zhao HJ, Xue W (2014) Synthesis and biological activity of oxime ethers of curcumin derivatives. *Chin J Org Chem* 34:1662–1668
24. Wang XB, Chen MH, Li Q, Zhang JP, Ruan XH, Xie Y, Xue W (2017) Synthesis and antiviral activities of novel penta-1,4-diene-3-one oxime derivatives bearing a pyridine moiety. *Chem Pap* 71:1225–1233
25. Han Y, Ding Y, Xie DD, Hu DY, Li P, Li XY, Xue W, Jin LH, Song BA (2015) Design, synthesis, and antiviral activity of novel rutin derivatives containing 1,4-pentadien-3-one moiety. *Eur J Med Chem* 92:732–737
26. Long CW, Li P, Chen MH, Dong LR, Hu DY, Song BA (2015) Synthesis, anti-tobacco mosaic virus and cucumber mosaic virus activity, and 3D-QSAR study of novel 1,4-pentadien-3-one derivatives containing 4-thioquinazolinone moiety. *Eur J Med Chem* 102:639–647
27. Wu F, Li P, Hu DY, Song BA (2016) Design, synthesis, and antiviral activity of novel purine derivatives containing 1,4-pentadien-3-one moiety. *Res Chem Intermed* 42:7153–7168
28. Zhu SR, Wang J, Chandrashekar G, Smith E, Liu XJ, Zhang YS (2010) Synthesis and evaluation of 4-quinazolinone compounds as potential antimalarial agents. *Eur J Med Chem* 45:3864–3869
29. Mohamed MS, Kamei MM, Kassem EM, Abotaleb N, Nofal SM, Ahmed MF (2009) Novel 3-(p-substituted phenyl)-6-bromo-4(3H)-quinazolinone derivatives of promising anti-inflammatory and analgesic properties. *Drug Res* 66:487–500
30. Wang X, Li P, Li ZN, Yin J, He M, Xue W, Chen ZW, Song BA (2013) Synthesis and bioactivity evaluation of novel arylimines containing a 3-aminoethyl-2-((p-trifluoromethoxy) aniline)-4(3H)-quinazolinone moiety. *J Agric Food Chem* 61:9575–9582
31. Kumar KS, Ganguly S, Veerasamy R, De Clercq E (2010) Synthesis, antiviral activity and cytotoxicity evaluation of schiff bases of some 2-phenylquinazolinone-4(3H)-ones. *Eur J Med Chem* 45:5474–5479
32. Abdel-Hamid SG (1997) Synthesis of some new heterocyclic systems bearing 2-phenyl-6-iodo-4(3H)-quinazolinone-3-yl moiety as antibacterial agents. *J Indian Chem Soc* 74:613–618
33. Chen MH, Li P, Hu DY, Zeng S, Li TX, Jin LH, Xue W, Song BA (2016) Synthesis, antiviral activity, 3D-QSAR, and interaction mechanisms study of novel malonate derivatives containing quinazolin-4(3H)-one moiety. *Bioorg Med Chem Lett* 26:168–173
34. Zhong XM, Wang XB, Chen LJ, Ruan XH, Li Q, Zhang JP, Chen Z, Xue W (2017) Synthesis and biological activity of myricetin derivatives containing 1,3,4-thiadiazole scaffold. *Chem Cent J* 11:106
35. Jin Y, Song BA, Hu DY, Li XY, Bhadury PS, Wang ZC, Yang S (2011) Inorganic base-catalyzed formation of antivirally active N-substituted benzamides from a-amido sulfones and N-nucleophile. *Chem Cent J* 5:21
36. Li XY, Song BA, Chen X, Wang ZC, Zeng MJ, Yu DD, Hu DY, Chen Z, Jin LH, Yang S, Yang CG, Chen BE (2013) Crystal structure of a four-layer aggregate of engineered TMV CP implies the importance of terminal residues for oligomer assembly. *PLoS ONE* 8:e77717
37. Li XY, Hao GF, Wang QM, Chen Z, Ding Y, Lu Y, Hu DY, Song BA (2017) Ningnanmycin inhibits tobacco mosaic virus virulence by binding directly to its coat protein discs. *Oncotarget* 8:82446–82458
38. Li XY, Liu J, Yang X, Ding Y, Wu J, Hu DY, Song BA (2015) Studies of binding interactions between Dufulin and southern rice black-streaked dwarf virus P9-1. *Bioorg Med Chem* 23:3629–3637
39. Wienken CJ, Baaske P, Rothbauer U, Braun D, Duhr S (2010) Protein-binding assays in biological liquids using microscale thermophoresis. *Nat Commun* 1:100

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

