

Journal of Freshwater Ecology



ISSN: 0270-5060 (Print) 2156-6941 (Online) Journal homepage: https://www.tandfonline.com/loi/tjfe20

Population dynamics of introduced flathead catfish in Lake Mitchell, South Dakota

David O. Lucchesi, Matthew D. Wagner, Tanner M. Stevens & B. D. S. Graeb

To cite this article: David O. Lucchesi, Matthew D. Wagner, Tanner M. Stevens & B. D. S. Graeb (2017) Population dynamics of introduced flathead catfish in Lake Mitchell, South Dakota, Journal of Freshwater Ecology, 32:1, 323-336, DOI: <u>10.1080/02705060.2017.1292963</u>

To link to this article: https://doi.org/10.1080/02705060.2017.1292963

9	© 2017 State of South Dakota, Department of Game, Fish and Parks. Published by Informa UK Limited, trading as Taylor & Francis Group
	Published online: 16 Mar 2017.
	Submit your article to this journal 🗹
ılıl	Article views: 1218
Q ^L	View related articles ☑
CrossMark	View Crossmark data ☑
4	Citing articles: 3 View citing articles 🗹



a OPEN ACCESS

Population dynamics of introduced flathead catfish in Lake Mitchell, South Dakota

David O. Lucchesi 📭, Matthew D. Wagner^b, Tanner M. Stevens^c and B. D. S. Graeb^d

^aSouth Dakota Department of Game, Fish and Parks, Sioux Falls, SD, USA; ^bMississippi Department of Wildlife, Fisheries, and Parks, Jackson, MS, USA; ^cMinnesota Department of Natural Resources, Little Falls, MN, USA; ^dDepartment of Natural Resource Management, South Dakota State University, Brookings, SD, USA

ABSTRACT

The introduction of flathead catfish (Pylodictis olivaris) into Lake Mitchell presented a unique opportunity to study the population dynamics of this species in a South Dakota impoundment. We collected flathead catfish using daytime, low-frequency electrofishing during June 2013, 2014, and 2015 and July, August and October of 2014 to examine population characteristics including abundance, recruitment, mortality, growth, condition, and diet. The flathead catfish population in Lake Mitchell was estimated at 1348 individuals (95% CI = 459-1455; density = 4.97/ha) in 2014 and 1197 individuals (95% CI = 931–1461; density = 4.42/ha) in 2015. Individuals from 11 year classes ranging from 1 to 13 years old were present. The population exhibited consistent recruitment, and annual mortality was estimated at 39%. Flathead catfish grew quickly exceeding stock length at age 3 and quality length at age 5; however, growth slowed in 2015. Similarly, condition of substock and stock-quality length fish declined in 2014 and 2015, respectively. The decline in growth and condition coincided with the recruitment of a large 2012 year class and may be an early indicator of intraspecific competition. Diets of Lake Mitchell flathead catfish primarily consisted of crayfish (Orconectes spp.) and fish with flathead catfish shifting to piscivory at approximately 400 mm. The high percentage of centrarchids in flathead catfish diets along with an increase in flathead catfish abundance coinciding with a decrease in bluegill abundance may indicate that flathead catfish are negatively impacting the bluegill population in Lake Mitchell.

ARTICLE HISTORY

Received 10 November 2016 Accepted 4 February 2017

KEYWORDS

Pylodictis olivaris; flathead catfish; population dynamics; predation; South Dakota

Introduction

Flathead catfish (*Pylodictis olivaris*) have been widely introduced because of their popularity with anglers (Jackson 1999; Kwak et al. 2006; Kaeser et al. 2011). They are renowned for their aggressive behavior, fighting ability when hooked, palatability, and potential to reach a large size (Jackson 1999). Flathead catfish have been stocked for purposes of recreational angling and as a predatory control of stunted or undesirable fish populations (Davis 1985; Haas et al. 2001; Vrtiska et al. 2003). Following introduction, populations often experience steady growth and can obtain a high abundance and biomass (Guier et al. 1981; Quinn 1988; Moser & Roberts 1999; Weller & Geihsler 1999; Kaeser et al. 2011). However, flathead catfish can impact native fish populations and their introduction into rivers in the eastern United States has resulted in the decline of native bullheads (Guier et al. 1981; Thomas 1993; Moser & Roberts 1999; Kaeser et al. 2011).

In South Dakota, flathead catfish are native to the Missouri River and its major tributaries, where they provide popular fisheries in the lower portions of the Big Sioux and James rivers (Doorenbos et al. 1996; Doorenbos et al. 1999). In Lake Mitchell, South Dakota, flathead catfish were first sampled in 2007 and were presumed to have originated from an unauthorized introduction from the nearby James River. Nighttime electrofishing catches of 22 and 46 flathead catfish in 2011 and 2012, respectively (T. M. Stevens, Minnesota Department of Natural Resources, unpublished data), provided evidence of natural recruitment and an established population.

Information on the food habits of flathead catfish in reservoirs and their impact on the fish community is relatively scarce (Turner & Summerfelt 1970; Layher & Boles 1980; Haas et al. 2001; Vrtiska et al. 2003; Katt 2016b). The diets of smaller flathead catfish (<500 mm) commonly consist of invertebrates including crayfish (Orconectes spp.) and smaller fish, while larger individuals are primarily piscivorous. Highly abundant forage fish, such as gizzard shad (Dorosoma cepedianum), freshwater drum (Amplodinotus grunniens), and white perch (Morone americana) were the most common food items in flathead catfish in Great Plains reservoirs (Turner & Summerfelt 1970; Layher & Boles 1980; Vrtiska et al. 2003) while flathead catfish introduced into a Minnesota lake primarily consumed common carp (Cyprinius carpio) and black bullheads (Ameiurus melas) (Davis 1985). Flathead catfish appear to be opportunistic feeders typically consuming the most abundant and easily-accessible prey items (Layher & Boles 1980; Davis 1985; Haas et al. 2001; Vrtiska et al. 2003; Baumann & Kwak 2011).

Flathead catfish predation has been shown to reduce or nearly eliminate centrarchids in ponds (Hackney 1966; Swingle 1967) and rivers (Evans 1991; Thomas 1993). Herndon and Waters (2002) suggested that based on their high abundance and use of centrarchids as prey, flathead catfish would have an impact on the largemouth bass (Micropterus salmoides) and sunfish fishery in Lake Sutton, a cooling reservoir similar in size to Lake Mitchell. South Dakota Game, Fish and Parks (SDGFP) fisheries staff are concerned about the potential negative impact of flathead catfish predation upon bluegill (Lepomis macrochirus) and crappie (Pomoxis spp.) fisheries in Lake Mitchell. The objectives of this study were to describe the population dynamics (abundance, recruitment, growth, mortality, and condition), and quantify food habits of flathead catfish in Lake Mitchell, South Dakota.

Methods

Lake Mitchell, located in southeastern South Dakota (Davison County), is a 271-ha impoundment with a maximum depth of 8.8 m and mean depth of 3.7 m. The lake was constructed in 1928 by the City of Mitchell to serve as a domestic water supply and regional recreation center. The primary source of water is Firesteel Creek, which contains two main branches and drains a watershed of 93,081 ha extending 80 km above the lake. The lake is owned by the City of Mitchell and the fishery is managed by SDGFP. The City owns and maintains several public access areas and parks around the lake while the remainder of the lakeshore is privately owned and highly developed. About 50% of the shoreline is rocky habitat (mostly rip-rap). The predominant aquatic vegetation in the lake is sago pondweed (Potamogeton pectinatus); however, an invasive species, curlyleaf pondweed (Potamogeton crispus) has been present since 2012. Lake Mitchell contains a fish community with at least 15 species present. The most abundant game fish include bluegill, black crappie (Pomoxis nigromaculatis), white crappie (Pomoxis annularis), largemouth bass, channel catfish (Ictalurus punctatus), flathead catfish, and walleye (Sander vitreus). Common non-game fish species include common carp, bigmouth buffalo (Ictiobus cyprinellus), smallmouth buffalo (I. bubalus), freshwater drum, shorthead redhorse (Moxostoma macrolepidotum), white sucker (Catastomus commersoni), and green sunfish (*L. cyanellus*).

We collected flathead catfish using daytime low-frequency electrofishing (LFE) with a Smith-Root (7.5 GPP) electrofishing boat using low-pulse frequency (7.5-15 pulses/s) and direct current (2-3 A, 500 V). We used a chase boat to assist in netting surfacing fish in 2014 and 2015 (Gilliland 1988; Cunningham 1995), but not 2013. Electrofishing was completed during June of 2013, 2014,

and 2015, and July, August, September, and October of 2014. Total sampling time varied from 130 to 260 min per sampling event; electrofishing was done in 10-min intervals in 2014 and 2015 and 20-min intervals in 2013. Flathead catfish relative abundance was quantified as mean catch per hour of electrofishing (CPUE) of all sizes of fish and was compared between sampling events with a one-way analysis of variance (ANOVA). Tukey's honestly significant difference (HSD) test was employed to determine where differences existed when the ANOVA was significant.

We measured (total length, mm), weighed (g), removed adipose fins, and determined gender through external characteristics (with the exception of 2013) of all flathead catfish. We compared the proportion of males to females collected between months using chi-square analysis. We extracted pectoral spines from a subsample of 10 fish per 10-mm group for age and growth analysis. We prepared spines according to Koch and Quist (2007) and sectioned them at both the articulating process and the basal recess with a Buehler low-speed isomet saw (Buehler, Lake Bluff, Illinois, USA). We viewed spine sections with a stereo microscope (UNITRON Z850; Commack, New York, USA) using transmitted light through a darkfield attachment and recorded a digital image of each structure from each fish with a Motic 5.0 MP camera (Richmond, British Columbia, Canada) mounted on the trinocular body of the stereo microscope. All spines were aged independently by at least two experienced readers. Any discrepancies in assigned ages were resolved by consensus. The number of visible annuli, the distance from the focus to each annulus (mm), and the radial distance from the focus to the margin (mm) were determined and used to back-calculate length-at-age with the Fraser–Lee direct proportion method (Carlander 1982). Von Bertalanffy growth models

$$L_t = L_{\infty} [1 - e^{-k(t-t_0)}],$$

where L_t is the predicted total length at time t in years, L_{∞} is the theoretical maximum mean total length, e is the base of the natural logarithm, k is the growth coefficient, and t_0 is the hypothetical age at which fish length equals zero were fitted to mean length-at-age data for each population (Ricker 1975) using FAMS 1.64 (Fishery Analysis and Modeling Simulator; Slipke & Maceina 2014).

To test for differences in growth by gender and year, the length increment added in the last year was regressed as a function of initial length (for that year) and analysis of covariance (ANCOVA) was used to compare the regressions. All statistical tests were completed using SYSTAT 13 (SYSTAT Software Inc., Richmond, CA) with a significance level of 0.05.

We developed age-length keys using FAMS 1.64 (Slipke & Maceina 2014) for the 2013–2015 population samples, and then assigned ages to all individuals collected during electrofishing sampling events. The 2013–2015 age frequencies were pooled and \log_e (CPUE) was plotted against age for fish age 2 (the youngest age that appeared fully recruited to the sampling gear) and older in order to estimate instantaneous total mortality rate (Z). Total annual mortality (A) was derived from the formula $A = 1 - e^{-Z}$ (Ricker 1975).

We evaluated condition of flathead catfish using relative weight (W_r) . We tested differences in W_r values across years by length categories (Quinn 1989; substock, stock-quality, quality-preferred, preferred and longer) using a one-way ANOVA. In instances when the ANOVA was significant, Tukey's HSD test was used to determine where differences existed.

We used the recruitment variability index (RVI; Guy & Willis 1995) to evaluate variation in recruitment. The RVI employs presence/absence and magnitude of year classes and is sensitive to missing year classes. Values of RVI can range from -1 to 1 with values close to 1 indicating consistent recruitment.

We used single and multiple census mark-recapture models to estimate the flathead catfish population size with corresponding 95% confidence intervals (Ricker 1975). A Schnabel population estimate was obtained in 2014 from recaptures of fish marked during the three consecutive days of sampling. In 2015, the population was estimated using the Petersen method based on fish that were marked with an adipose fin-clip in 2014. Adipose-clipped marks applied in 2014 were easily identifiable in 2015.

326 D. O. LUCCHESI ET AL.

We collected up to 25 diets for each 100 mm length group in 2014 using either pulsed gastric lavage (Foster 1977; Waters et al. 2004) or manual extraction. Stomach contents were screened through a 710 μ m sieve, fixed in 10% formalin, preserved in 70% ethanol, identified to the lowest possible taxa, counted, and weighed. Stomach contents were further quantified by frequency of occurrence, percent composition by number, and percent composition by wet weight (Bowen 1996; Herndon & Waters 2002). We estimated the shift to piscivory by using logistic regression on the presence or absence of fish prey in diets across length.

Results

A total of 585 flathead catfish were collected during nine sampling events from June 2013 through June 2015. The percentage of male, female, and immature fish collected in 2014 and 2015 was 56%, 38%, and 6%, respectively. The proportion of males to females varied by month ($\chi^2 = 9.14$, df = 3, P = 0.027) with more males than females collected in June 2014, August 2014, and June 2015, and similar numbers collected in July 2014. Only three flathead catfish (two males and one female) were collected on 13 October 2014 and, therefore, it was not included in the analysis of sex ratio or mean CPUE.

Mean CPUE differed between sampling trips ($F_{3,72} = 4.21$, P = 0.008). The mean CPUE on 09 June 2014 (48.8/h) was greater than mean CPUEs for sampling events in July and August (Tukey's HSD, Figure 1). The second highest mean CPUE occurred on the 09 June 2015 sampling event; however, it was not significantly different from any of the others.

In 2015, we recaptured 53 (14.8%) of the 359 fish marked in 2014. The number of flathead catfish in Lake Mitchell was estimated at 1348 individuals (95% CI = 459-1455; density = 4.97/ha) in 2014 and 1197 individuals (95% CI = 931-1461; density = 4.42/ha) in 2015.

The estimated ages of flathead catfish ranged from 1 to 13 years and individuals were collected from 11 year classes (Figure 2). The large 2012 year class comprised 62%, 58% and 49% of the total sample in 2013, 2014, and 2015, respectively. Total annual mortality of fish age 2 and older was estimated at 39%. The population exhibited consistent recruitment with RVI values ranging from 0.45 in 2013 to 0.77 in 2015. There were no missing year classes between 2006 and 2015.

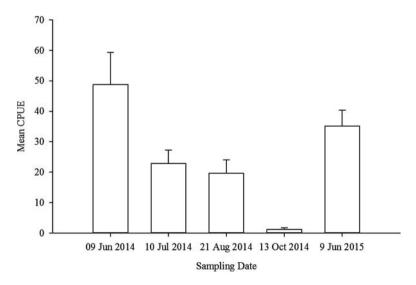


Figure 1. Mean catch per hour (CPUE) of flathead catfish sampled from Lake Mitchell, South Dakota, 2014–2015. Only three flathead catfish were collected on 13 October 2014, so it was excluded from the analysis. Vertical bars represent \pm one SE.

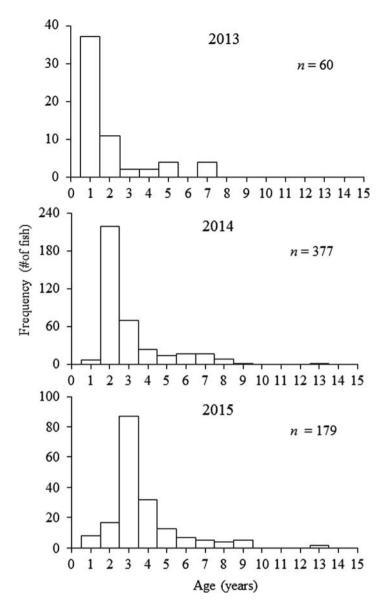


Figure 2. Age frequency distribution of flathead catfish collected from Lake Mitchell, South Dakota, 2013–2015.

Flathead catfish grew quickly exceeding stock length at age 3 and quality length at age 5 (Table 1). Incremental growth was similar in 2013 and 2014 (ANCOVA: slope test, $F_{1,153} = 0.56$, P = 0.453; intercept test, $F_{1,154} = 1.74$, P = 0.190), but was slower in 2015 than in 2013 (slope test, $F_{1,116} = 27.78$, P < 0.001) or 2014 (slope test, $F_{1,209} = 18.13$, P < 0.001). Incremental growth was similar for males and females in 2014 (slope test, $F_{1,111} = 1.65$, P = 0.201; intercept test, $F_{1,112} = 3.66$, P = 0.058), but slower for males in 2015 (slope test, $F_{1,83} = 12.74$, P = 0.001).

Von Bertalanffy growth models fit the data well for both males and females ($r^2 = 0.99$; Table 2). Back-calculated and von Bertalanffy modeled growth rates were very similar for males and females from age 1 through age 8 (Figure 3). Modeled growth for males and females began to diverge after age 8 because no fish exceeding age 9 were available for inclusion in the male growth model while the female growth model included two age 13 individuals (Table 1).

Table 1. Mean (± 95% C.I.) back-calculated length-at-age (mm) and number of fish at each estimated age for male (M) and female
(F) flathead catfish collected from Lake Mitchell, South Dakota, 2013–2015.

	2014				2015				
	Male		Female		Male		Female		
Age	n	Mean	n	Mean	N	Mean	n	Mean	
1	77	189 ± 12	38	180 ± 19	14	167 ± 21	19	181 ± 19	
2	77	302 ± 13	38	306 ± 18	14	285 ± 31	19	296 ± 20	
3	34	380 ± 24	21	390 ± 26	13	367 ± 51	17	402 ± 30	
4	21	451 ± 35	16	441 ± 22	8	470 ± 81	15	476 ± 41	
5	16	531 ± 76	11	529 ± 32	5	567 ± 113	11	552 ± 51	
6	15	590 ± 54	8	608 ± 63	4	662 ± 156	8	639 ± 75	
7	8	651 ± 105	3	646 ± 99	4	718 ± 160	6	720 ± 62	
8	2	776	3	687 ± 100	3	722 ± 232	5	779 ± 53	
9					1	840	4	805 ± 64	
13			1	968			1	969	

Table 2. Flathead catfish age range and growth parameters (95% confidence limits in parentheses) of von Bertalanffy models fitted to mean total length at age data pooled for 2014 and 2015 for males and females collected from Lake Mitchell, South Dakota (L_{∞} is the theoretical maximum mean total length, k is the growth coefficient, and t_0 is the hypothetical age at which fish length = 0, and r^2 is the coefficient of determination for model fit with respect to all age classes).

	Si	ex	
Parameter	Males	Females	
Age range (years)	1–9	1-13	
L_{∞} (mm)	2758	1205	
k	0.035	0.111	
t_{o}	−1.129	-0.505	
r ²	0.99	0.99	

Relative weights differed between years for substock ($F_{2.307} = 8.05$, P < 0.001) and stock-quality $(F_{2,224} = 6.82, P = 0.001)$ length categories, but not for quality-preferred $(F_{2,65} = 0.64, P = 0.532)$ and preferred ($F_{2,22} = 0.51$, P = 0.609) length categories (Table 3). Condition of substock fish was significantly lower in 2014 than 2013, and the stock-quality length fish exhibited significantly lower

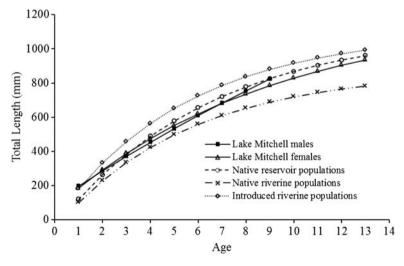


Figure 3. A comparison of von Bertalanffy growth curves for male and female flathead catfish collected from Lake Mitchell in 2014 and 2015 and for native riverine, introduced riverine and native reservoir populations in the USA modeled from length-at-age data summarized by Kwak et al. (2006).

Table 3. Mean relative weight (W_n , SE in parentheses) by length category [substock (SS, > 35 cm), stock-quality (SQ, 35–50 cm), quality-preferred (QP, 51–70 cm) and preferred ($P_n > 70$ cm)] for each year for flathead catfish from Lake Mitchell, 2013–2015. Mean W_n values denoted by different letters (a-c for years, w-z for size categories) were significantly different at $p \le 0.05$. Differences were not tested for the categories of QP and P in 2013 because sample size was too small (n < 5).

		Lengt	h category	
Year	SS	SQ	QP	Р
2013	97.8 (1.5) ^{a,x}	85.1 (2.4) ^{a,y}	80.9 (0.4)	92.5 (3.5)
2014	90.4 (1.1) ^{b,w}	85.4 (0.9) ^{a,x}	83.9 (1.5) ^{<i>a,x</i>}	94.6 (7.2) ^{a,w,x}
2015	86.4 (1.2) ^{b,w}	81.1 (0.7) ^{b,x}	86.7 (2.6) ^{a,w,y}	100.9 (2.6) ^{a,z}

Table 4. Stomach contents of three sizes of flathead catfish collected in 2014 from Lake Mitchell, South Dakota quantified by frequency of occurrence (FO), percent composition by number (PN), and percent composition by wet weight (PW).

	<300 mm ($n = 30$)		300–600 mm (<i>n</i> = 77)			>600 mm ($n = 7$)			
Food item	FO	PN	PW	FO	PN	PW	FO	PN	PW
Centrarchidae	17	14	33	42	43	77	86	86	97
Ictaluridae	7	6	4	3	2	trace	0	0	0
Unidentified fish	7	5	1	7	5	1	14	14	3
All teleostei	27	25	38	49	50	78	100	100	100
Orconectes spp.	67	67	62	55	50	22	0	0	0
Chironomidae	10	8	trace	0	0	0	0	0	0

condition in 2015 than 2014. We also detected significant differences in mean relative weight between length categories within years (Table 3). Mean relative weight differed between the substock and stock-quality length categories in all three years.

A total of 114 diets were collected from flathead catfish; 30 from fish less than 300 mm, 77 from fish 300–600 mm, and 7 from fish over 600 mm (Table 4). Crayfish comprised approximately 67% and 62% of the diets of smaller fish (>300 mm) by number and weight, respectively. Stomachs of 300–600 mm fish contained nearly equal numbers of crayfish and fish; however, fish comprised a substantially higher proportion of the diet by weight (Table 4). Larger flathead catfish (>600 mm) had only fish their diets. Centrarchidae comprised 77% and 97% by weight of fish in diets of 300–600 mm and >600 mm flathead catfish, respectively. Ictaluridae in the diet were all smaller flathead catfish. Chironomidae were found in the diet of three smaller (<190 mm) individuals. The proportion of fish to crayfish in the diet was similar between the months of July and August (Figure 4). Flathead catfish in Lake Mitchell shifted to piscivory at approximately 400 mm. The logistic regression fit the data well (null deviance = 157.48, df = 113; residual deviance = 119.15, df = 112; Figure 5) and few fish over 400 mm had non-fish prey in their diets.

Discussion

Low-frequency electrofishing is the most common gear used to sample flathead catfish (Brown 2009) and a chase boat is often employed when sampling lotic systems (Cunningham 2004). Low-frequency electrofishing appeared to be an effective method for capturing flathead catfish on Lake Mitchell although catch rates commonly fell below the range (38.5–58.0 fish/h) reported for Oklahoma reservoirs (Gilliland 1988; Cunningham 1995, 2000). The relative abundance of flathead catfish in Lake Mitchell was similar to the nearby James River, but higher than that reported for the Big Sioux River in South Dakota (Arterburn 2001). Electrofishing catches for lentic systems on average are higher than for lotic systems (Bodine et al. 2013) and a chase boat was not employed to do the river sampling (Arterburn 2001).

We were most effective at collecting flathead catfish in the month of June. Maximum catch rates in lotic systems have been achieved during summer months, when water temperatures exceed 20 °C and water levels were low (Quinn 1986; Travnichek 2011); however, sampling conditions relative to efficiency have not been evaluated for lentic habitats (Bodine et al. 2013). Although we

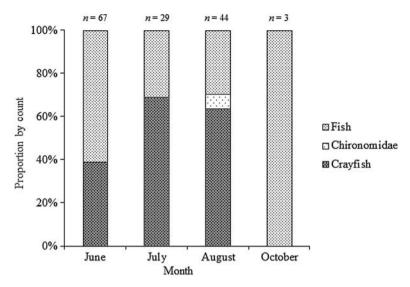


Figure 4. Proportion by number of food items in the stomachs of flathead catfish collected from Lake Mitchell, South Dakota, in 2014.

did not quantify catch by habitat type, a large proportion of the flathead catfish were captured near rocky habitat. Vrtiska et al. (2003) and Katt (2016b) also observed a preference for rocky habitat by flathead catfish in Branched Oak Reservoir, Nebraska. Layher and Boles (1980) caught flathead catfish along the rocky rip-rap of the causeway at Milford Reservoir, Kansas, in July and August and attributed their high abundance at that location to use of rip-rap as spawning, feeding and resting habitat. However, unlike Lake Mitchell, Branched Oak and Milford Reservoir flathead catfish, especially larger fish, were also common in woody habitat.

Estimates of flathead catfish density in lentic waters are scarce, and thus, little is available for comparison with Lake Mitchell. Recent research in Nebraska impoundments yielded estimates of flathead catfish density of 3.4/ha in Pawnee Reservoir, Nebraska (Katt 2016a) and 7.4/ha in

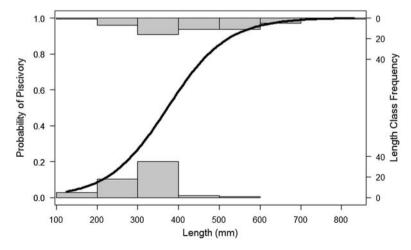


Figure 5. Probability of piscivorous feeding across length classes of flathead catfish in Lake Mitchell, SD, as estimated by logistic regression of diets (presence or absence of fish prey). Frequency of individuals within each size class is shown for piscivorous diets (top) and non-fish diets (bottom).

Branched Oak Reservoir (Katt 2016b). Density in Branched Oak Reservoir was higher than in Lake Mitchell, while Pawnee Reservoir was similar. Density of flathead catfish in lotic systems in the southeastern United States has been estimated at up to 15-18/ha for recently-introduced populations (Quinn 1988; Weller & Geihsler 1999), but is typically in the range of 1-8/ha for established populations (Kwak et al. 2006; Kaeser et al. 2011).

Flathead catfish in Lake Mitchell are a product of an unauthorized introduction and may still be experiencing population growth as newly-established populations tend to increase rapidly in abundance and biomass (Quinn 1988; Moser & Roberts 1999; Weller & Geihsler 1999). Accordingly, the Lake Mitchell population appears to be expanding because it is dominated by younger fish recruited from the large 2012 year class. Eventually, this growth phase will culminate at a peak density and biomass, which has been demonstrated to occur at approximately 10 to 15 years for flathead catfish introduced into rivers (Guier et al. 1981; Kaeser et al. 2011). This peak abundance is typically not sustained, but followed by a decline in abundance and growth rate (Kaeser et al. 2011).

Lake Mitchell flathead catfish exhibited consistent recruitment with only two missing year classes within the last 13 years. The limited information available suggests that consistent recruitment in flathead catfish is common in impoundments across the Great Plains (Layher & Boles 1979; Katt 2016a, Katt 2016b). Similar to Lake Mitchell, 13 of 16 year classes (1959-1974) were sampled at Milford Reservoir (Layher & Boles 1979) and 13 of 15 year classes (1997-2011) at Branched Oak Reservoir (Katt 2016b). Researchers attributed missing year classes to periods of low water in both reservoirs. Lake Mitchell has a stable reservoir hydrology (Tanner Stevens, unpublished data) which may help to explain the absence of missing year classes. Relatively consistent recruitment has also been reported for other populations in this area like in the lower unchannelized reach of the Missouri River (Gavin's Point Dam to the confluence with the Big Sioux River; Goble 2011) and the lower end of its South Dakota tributaries (James and Big Sioux Rivers; Arterburn 2001).

Annual mortality of flathead catfish in Lake Mitchell (39%) was at the upper end of the range reported from lentic and lotic systems. The only other estimate of annual mortality for a flathead catfish population in lentic waters came from at Lake Wilson, Alabama (17%; Marshall et al. 2009), and was lower than for Lake Mitchell. Estimates for lotic populations range from about 12%-36% (Daughtery & Sutton 2005; Kwak et al. 2006; Makinster & Paukert 2008; Jolley & Irwin 2011; Kaeser et al. 2011), however, values similar to Lake Mitchell were reported for the Missouri River (34%-59%, Gavin's Point Dam to the Kansas-Nebraska border; Goble 2011) and the Satilla River, Georgia (45%; Sakaris et al. 2006).

The large number of young flathead catfish from the 2012 year class may have inflated our estimate of annual mortality. The dominance of that year class was apparent by its much larger contribution to the total catch at age 1 and age 2 than that of subsequent year classes. Additionally, anecdotal evidence suggests that LFE is selective towards fish less than 600 mm (Brown 2009; Ford et al. 2011); another factor potentially biasing our estimated mortality. We would expect that angler exploitation of flathead catfish is low in Lake Mitchell based upon the observation that reservoir channel catfish fisheries in South Dakota are generally under-utilized and subject to low harvest (Galinat 2004; Stevens 2013; Lucchesi et al. 2015; Potter et al. 2016).

Flathead catfish in Lake Mitchell grew similarly to introduced riverine and native reservoir populations (Kwak et al. 2006) and faster than native riverine populations (Arterburn 2001; Kwak et al. 2006). Lake Mitchell fish also grew faster than fish in the James, Big Sioux and Missouri rivers (Arterburn 2001; Goble 2011). Growth to quality size (510 mm) was reached between age 4 and 5 in Lake Mitchell, but was not reached until nearly age 6 in the James and Big Sioux Rivers and between age 7 and age 8 in the Missouri River (Fort Randall Dam to the Kansas-Nebraska border). Similarly, Lake Mitchell fish reached preferred length (710 mm) between age 7 and 8, while it took between 9 and 10 years for fish in the Big Sioux and James rivers (Arterburn 2001). Lake Mitchell flathead catfish grew faster than fish in Nebraska reservoirs (Katt 2016a, 2016b), possibly because the Lake

Mitchell population was recently introduced while the Nebraska reservoirs contained well-established populations (Kwak et al. 2006; Kaeser et al. 2011).

Mean relative weight of Lake Mitchell flathead catfish was lower than for populations in South Dakota rivers (Arterburn 2001) and Nebraska reservoirs (Katt 2016a, 2016b) and formed a U-shaped pattern relative to length typical of most channel catfish populations (Brown et al. 1995; Doorenbos et al. 1999; Stevens 2013). The lower mean relative weight of stock to preferred length fish relative to other length categories may be related to fish size and a transition from a diet of invertebrates to fish and larger crustacea (Layher & Boles 1980, Jackson 1999, Vrtiska et al. 2003). Higher relative weight of stock to preferred length fish in other reservoir flathead catfish populations was most likely related to extremely abundant fish forage such as age 0 gizzard shad in Milford Reservoir (Layher & Boles 1980) or stunted white perch in Branched Oak Reservoir (Vrtiska et al. 2003). Fish have a higher caloric density than crayfish (Eggleton & Schramm 2002) and stock to preferred length flathead catfish in these reservoirs had a higher percent composition of fish to crayfish in their diets than in Lake Mitchell. High abundance of small forage fish may release fish from density dependence at a smaller size.

In Lake Mitchell, the recruitment of the large 2012 year classes coincided with reduced incremental growth and lower relative weight in 2015 and may suggest that the availability of quality foraging habitat is becoming scarce, and subsequently, the population is approaching carrying capacity. Layher and Boles (1980) postulated that population size in reservoirs may be limited by the availability of suitable habitat for foraging (e.g. interstitial spaces in the rip-rap) rather than the availability of forage. Most of the flathead catfish in Lake Mitchell were concentrated in and around rocky shorelines, rip-rap, and the dam face and had not expanded into woody or open habitats. Kwak et al. (2006) suggests that intraspecific competition and other density dependent factors may slow growth in introduced populations to where it approaches that of native populations. Kaeser et al. (2011) observed not only a decrease in growth rate, but also abundance over time in introduced riverine populations in Georgia.

Centrarchids comprised a larger proportion of the diet of flathead catfish in Lake Mitchell than in many other reservoir populations (Turner & Summerfelt 1970; Layher & Boles 1980; Haas et al. 2001; Jolley & Irwin 2003; Vrtiska et al. 2003). Extremely abundant forage species such as gizzard shad, white perch, freshwater drum or common carp often are the primary food item of reservoir flathead catfish. Although Lake Mitchell does not currently contain gizzard shad, it does support populations of freshwater drum (Aplodinotus grunniens), common carp, and black bullhead, important food items of flathead catfish in other lentic systems (Turner & Summerfelt 1970; Davis 1985). However, these fish species were not abundant during the study period (SDGFP, unpublished data) and not found in the diets. Centrarchids have been shown to be a primary food item of reservoir flathead catfish (Edmonson 1974; Herndon & Waters 2002) when extremely abundance forage species like gizzard shad are not available. Edmondson (1974) reported that sunfish were the dominant forage consumed by flathead catfish in Bluestone Reservoir, West Virginia. Centrarchids conservatively comprised 53% of diet items by number in flathead catfish in Lake Sutton, Georgia (Herndon & Waters 2002), and similar to Lake Mitchell, were present in fish of all sizes.

Flathead catfish predation has been shown to reduce or nearly eliminate centrarchids in ponds (Hackney 1966; Swingle 1967) and rivers (Evans 1991; Thomas 1993), but has not been definitively tied to a decline in centrarchids in impoundments similar in size to Lake Mitchell. Herndon and Waters (2002) suggested that flathead catfish would have an impact on the largemouth bass and sunfish fishery in Lake Sutton, a 445-ha cooling reservoir, based on their high abundance and use of centrarchids as prey. Although assessing the impact of flathead catfish predation on bluegill and crappie populations in Lake Mitchell would be difficult as both are cyclical, the recent unprecedented decline in bluegill abundance gives some cause for concern. The four lowest trap-net catches of bluegill in the last 20 years were recorded from 2013 to 2016 (Figure 6; SDGFP, unpublished data). Troughs in

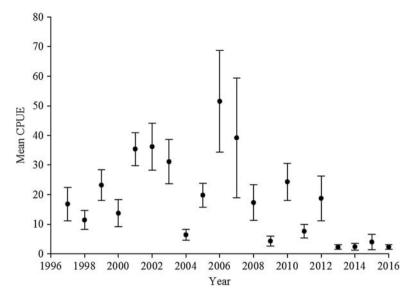


Figure 6. Mean catch per overnight set (CPUE) of bluegills in trap nets on Lake Mitchell, South Dakota, 1997–2016. Vertical bars represent \pm one SE.

bluegill abundance have traditionally lasted for only about a year before the numbers rebound in Lake Mitchell. Thus, the high percentage of centrarchids in flathead catfish diets along with an increase in flathead catfish abundance coinciding with a prolonged decrease in bluegill abundance may indicate that flathead catfish are negatively impacting the bluegill population in Lake Mitchell.

The population parameters estimated in this study most likely characterize the introduced Lake Mitchell flathead catfish population in its growth phase, possibly close to carrying capacity. The recently established population is now successfully reproducing resulting in a population currently skewed toward younger fish. Our assessment may have detected some early signs of intraspecific competition as the population approaches carrying capacity. Moreover, the increase in flathead catfish abundance has coincided with a decrease in bluegill abundance. A follow-up study of population dynamics would be beneficial to determine if Lake Mitchell flathead catfish experience a decline in growth and abundance similar to other introduced populations, and if flathead catfish diets vary over time.

Acknowledgments

We thank M. Brown and N. Kludt for help with statistical analyses and B. Blackwell for help with the processing of aging structures, statistical analyses, and preparation of the manuscript. We are grateful to all of the SDGFP personnel and South Dakota State University staff and students who helped with electrofishing and collecting the data. A review by S. Chipps improved an earlier version of this manuscript.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

South Dakota Department of Game, Fish and Parks.



Notes on contributors



David O. Lucchesi is a fisheries biologist with the South Dakota Department of Game, Fish and Parks (Sioux Falls, South Dakota). His research has primarily focused on coolwater fish population dynamics, fish stocking evaluation and determining angler use and harvest.



Matthew D. Wagner is a conservation resource biologist with Mississippi Department of Wildlife, Fisheries, and Parks (Jackson, Mississippi). His primary research focuses on the distribution and status of species of greatest conservation need in Mississippi. He is the curator of fishes at the Mississippi Museum of Natural Science, where he maintains a state-wide fish distribution database and manages the fish collection.



Tanner M. Stevens is a fisheries biologist with the Minnesota Department of Natural Resources (Little Falls, Minnesota). Tanner completed his BS degree from the University of Nebraska-Lincoln, and his MS degree from South Dakota State University. Primary research interests include black bass population dynamics, alternative fishing pressure estimation methods and evaluation of fisheries sampling methods.



Dr Brian Graeb is an assistant professor at the South Dakota State University School of Natural Resource Management (Brookings, South Dakota). His research focuses on reservoir fish population dynamics, community ecology and their relationship to habitat as well as the physiology and recovery of Pallid Sturgeon and other endangered fish species.

ORCID

David O. Lucchesi (D) http://orcid.org/0000-0001-8875-9481

References

Arterburn JE. 2001. Population characteristics and sampling methods of catfish for the James and Big Sioux rivers [M.S. Thesis]. Brookings (SD): South Dakota State University.

Baumann JR, Kwak TJ. 2011. Trophic relations of introduced flathead catfish in an Atlantic river. T Am Fish Soc. 140:1120–1134.

Bodine KA, Shoup DE, Olive J, Ford ZL, Krogman R, Stubbs TJ. 2013. Catfish sampling techniques: where we are now and where we should go. Fisheries 38:529–546.

Bowen SH. 1996. Quantitative description of the diet. In: Murphy BR, Willis DW, editors. Fisheries techniques. 2nd ed. Bethesda (MD): American Fisheries Society; p. 513–532.

Brown Z. 2009. Current trends in catfish sampling techniques and information needs. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 61:6–9.

Brown ML, Jaramillo Jr. F, Gatlin III DM, Murphy, BR. 1995. A revised standard weight (Ws) equation for channel catfish. J Freshwater Ecol. 10:295–302.

Carlander KD. 1982. Standard intercepts for calculating lengths from scale measurements for some centrarchid and percid fishes. T Am Fish Soc. 111:332–336.



Cunningham KK. 1995. Comparison of stationary and mobile electrofishing for sampling flathead catfish. North Am J Fish Mana. 15:515–517.

Cunningham KK. 2000. Influence of environmental variables on flathead catfish electrofishing catch. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 52:125–135.

Cunningham KK. 2004. Efficacy of a chase boat for electrofishing flathead catfish in three Oklahoma reservoirs. North Am J Fish Mana. 24:1427–1430.

Daughtery DJ, Sutton TM. 2005. Population abundance and stock characteristics of flathead catfish in the lower St. Joseph River, Michigan. North Am J Fish Mana. 25:1191–1201.

Davis RA. 1985. Evaluation of flathead catfish as a predator in a Minnesota lake. St. Paul: Minnesota Department of Natural Resources, Section of Fisheries. (Investigational Report 384).

Doorenbos R, Dieterman D, Berry Jr. C. 1996. Recreational use of the Big Sioux River, Iowa and South Dakota. Pierre (SD): South Dakota Department of Game, Fish and Parks. Special Report Number 9614.

Doorenbos R, Berry CR, Wickstrom G. 1999. Ictalurids in South Dakota. In: Irwin ER, Hubert WA, Rabeni CF, Schramm Jr. HL, Coon T, editors. Catfish 2000. Proceedings of the international ictalurid symposium. Bethesda (MD): American Fisheries Society, Symposium 24; p. 377–389.

Edmundson Jr. JP. 1974. Food habits, age and growth of flathead catfish, *Pylodictis olivaris* (Rafinesque), in Bluestone Reservoir, West Virginia [M.S. Thesis]. Morgantown (WV): West Virginia University.

Eggleton MA, Schramm Jr. HL. 2002. Caloric densities of selected fish prey organisms in the lower Mississippi River. J Freshwater Ecol. 17:409–414.

Evans JW. 1991. A fisheries and recreational use survey of the upper Ocmulgee River (Final Report, Federal Aid Project F–33). Social Circle (GA): Georgia Department of Natural Resources, Game and Fish Division.

Ford ZL, Sullivan KP, Vining IW, Kulowiec TG, Pitchford GD, Dames HR, Dent RJ, Colvin E. 2011. Sampling statistics and size distributions for flathead catfish populations in four Missouri rivers. In: Michaletz PH, Travnichek VH, editors. Conservation, ecology, and management of catfish: the second international symposium. Bethesda (MD): American Fisheries Society, Symposium 77; p. 95–104.

Foster JR. 1977. Pulsed gastric lavage: an efficient method of removing the stomach contents of live fish. Progressive Fish–Culturist. 39:166–169.

Galinat G. 2004. Angler use and fish harvest surveys on Angostura Reservoir, South Dakota, 2001 (Progress Report 04–17). Pierre (SD): South Dakota Department of Game, Fish and Parks.

Gilliland E. 1988. Telephone, micro-electronic, and generator-powered electrofishing gear for collecting flathead catfish. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 41:221–229.

Goble CW. 2011. Ecology and management of channel catfish *Ictalurus punctatus* and flathead catfish *Pylodictis oliva*ris populations in the Missouri River, NE [M.S.Thesis]. Lincoln (NE): University of Nebraska.

Guier CR, Nichols LE, Rachels RT. 1981. Biological investigations of flathead catfish in the Cape Fear River. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 35:607–621.

Guy CS, Willis DW. 1995. Population characteristics of black crappies in South Dakota waters: a case for ecosystem-specific management. North Am J Fish Mana. 15:887–897.

Haas JJ, Jahn LA, Hilsaback R. 2001. Diet of introduced flathead catfish (*Pylodictis olivaris*) in an Illinois reservoir. J Freshwater Ecol. 16:551–555.

Hackney PA. 1966. Predator–prey relationships of the flathead catfish in ponds under selected forage fish conditions. Proc Southeast Assoc Game Fish Commissioners. 19:217–222.

Herndon Jr. TM, Waters CT. 2002. Flathead catfish diet analysis, stock assessment, and effects of removal on Sutton Lake, North Carolina. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 54:70–79.

Jackson DC. 1999. Flathead catfish: biology, fisheries, and management. In: Irwin ER, Hubert WA, Rabeni CF, Schramm Jr. HL, Coon T, editors. Catfish 2000. Proceedings of the international ictalurid symposium. Bethesda (MD): American Fisheries Society, Symposium 24; p. 23–35.

Jolley JC, Irwin ER. 2003. Food habits of catfishes in tailwater and reservoir habitats in a section of the Coosa River, Alabama. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 57:124–140.

Jolley JC, Irwin ER. 2011. Catfish population characteristics in tailwater reservoir habitats of the Coosa River, Alabama. In: Michaletz PH, Travnichek VH, editors. Conservation, ecology, and management of catfish: the second international symposium. Bethesda (MD): American Fisheries Society, Symposium 77; p. 155–166.

Kaeser, AJ, Bonvechio TF, Harrison D, Weller RR. 2011. Population dynamics of introduced flathead catfish in rivers of southern Georgia. In: Michaletz PH and Travnichek VH, editors. Conservation, ecology, and management of catfish: the second international symposium. Bethesda (MD): American Fisheries Society, Symposium 77; p. 409–422.

Katt J. 2016a. Effects of a low-dose rotenone treatment on flathead catfish in Pawnee Reservoir, project update 2013–2014. Lincoln (NE): Nebraska Game and Parks Commission.

Katt J. 2016b. Population dynamics, habitat use, and electrofishing catchability of flathead catfish in Branched Oak Reservoir, project update 2010–2014. Lincoln (NE): Nebraska Game and Parks Commission.

Koch JD, Quist MC. 2007. A technique for preparing fin rays and spines for age and growth analysis. North Am J Fish Mana. 27:782–784.



Kwak TJ, Waters DS, Pine III WE. 2006. Age, growth and mortality of introduced flathead catfish in Atlantic rivers and a review of other populations. North Am J Fish Mana. 26:73-87.

Layher WG, Boles RJ. 1979. Growth of Pylodictis olivaris (Rafinesque) in a Kansas reservoir. T Kansas Acad Sci. 82:36-48. Layher, W. G., R. J. Boles. 1980. Food habits of the flathead catfish, Pylodictis olivaris (Rafinesque), in relation to length and season in a large Kansas reservoir. T Kansas Acad Sci. 83:200-214.

Lucchesi DO, St. Sauver TR, Chamblin CL, Kaiser C, Gust B. 2015. Angler use and harvest surveys on Lake Alvin, Wall Lake and East Vermillion Lake, 1995-2002 (Completion Report 15-01). Pierre (SD): South Dakota Department of Game, Fish and Parks.

Makinster AS, Paukert CP. 2008. Effects and utility of minimum length limits and mortality caps for flathead catfish in discrete reaches of a large prairie river. North Am J Fish Mana. 28:97-108.

Marshall MD, Holley MP, Maceina MJ. 2009. Assessment of the flathead catfish population in a lightly exploited fishery in Lake Wilson, Alabama. North Am J Fish Mana. 29:869-875.

Moser ML, Roberts SB. 1999. Effects of nonindigenous ictalurids and recreational electrofishing on the ictalurid community of the Cape Fear River drainage, North Carolina. In: Irwin ER, Hubert WA, Rabeni CF, Schramm Jr. HL, Coon T, editors. Catfish 2000. Proceedings of the international ictalurid symposium. Bethesda (MD): American Fisheries Society, Symposium 24; p. 479-485.

Potter K, Meyer H, Hanten R, Greiner M, Fincel M, Smith M. 2016. Annual fish population and angler use, harvest and preference surveys on Lake Oahe, South Dakota, 2015 (Annual Report 16-03). Pierre (SD): South Dakota Department of Game, Fish and Parks.

Quinn SP. 1986. Effectiveness of an electrofishing system for collecting flathead catfish. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 40:85-91.

Quinn SP. 1988. Flathead catfish abundance and growth in the Flint River, Georgia. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 42:141–148.

Quinn SP. 1989. Evaluation of a length-categorization system for flathead catfish. Proc Annu Conf Southeastern Assoc Fish Wildlife Agencies. 43:146–152.

Ricker WE. 1975. Computation and interpretation of biological statistics of fish populations. Fish Res Board Can Bull 191. Ottawa.

Sakaris PC, Irwin ER, Jolley JC, Harrison D. 2006. Comparison of native and introduced flathead catfish populations in Alabama and Georgia: growth, mortality, and management. North Am J Fish Mana. 26:867-874.

Slipke JW, Maceina MJ. 2014. Fishery analysis and modeling simulator (FAMS). Version 1.64. Bethesda (MD): American Fisheries Society.

Stevens TM. 2013. Feeding ecology and factors influencing growth and recruitment of channel catfish in South Dakota reservoirs [M.S. Thesis]. Brookings (SD): South Dakota State University.

Swingle HS. 1967. Experiments with the flathead catfish (*Pylodictis olivaris*) in ponds. Proc Southeast Assoc Game Fish Commissioners. 18:303-308.

Thomas ME. 1993. Monitoring the effects of introduced flathead catfish on sportfish populations in the Altamaha River, Georgia. Proc Annu Conf Southeastern Assoc of Fish Wildlife Agencies. 47:531–538.

Travnichek VH. 2011. Monthly variation and influences of habitat and river stage on electrofishing catch rates and population indices of flathead catfish from the lower Missouri River. In: Michaletz PH, Travnichek VH, editors. Conservation, ecology, and management of catfish: the second international symposium. Bethesda (MD): American Fisheries Society, Symposium 77; p. 581-588.

Turner RP, Summerfelt RC. 1970. Food habits of adult flathead catfish Pylodictis olivaris (Rafinesque) in Oklahoma reservoirs. Proc Southeast Assoc Game Fish Commissioners. 24:387-401.

Vrtiska Jr. LA, Peters EJ, Porath MT. 2003. Flathead catfish habitat use and predation on a stunted white perch population in Branched Oak Reservoir, Nebraska. J Freshwater Ecol. 18:605-613.

Waters DS, Kwak TJ, Arnott JB, Pine III WE. 2004. Evaluation of stomach tubes and gastric lavage for sampling diets from blue catfish and flathead catfish. North Am J Fish Mana. 24:258-261.

Weller RR, Geihsler MR. 1999. Angler attitudes concerning the management of the introduced flathead catfish in the Altamaha River system, Georgia. In: Irwin ER, Hubert WA, Rabeni CF, Schramm Jr. HL, Coon T, editors. Catfish 2000. Proceedings of the international ictalurid symposium. Bethesda (MD): American Fisheries Society, Symposium 24; p. 435-442.