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To cite this article: Paul J. Fitzgerald , Savannah K. Kounelis-Wuillaume , Ali Gheidi , Jonathan D. Morrow , Joanna L. Spencer-Segal & Brendon O. Watson (2021): Sex- and stress-dependent effects of a single injection of ketamine on open field and forced swim behavior, *Stress*, DOI: [10.1080/10253890.2021.1871600](https://doi.org/10.1080/10253890.2021.1871600)

To link to this article: <https://doi.org/10.1080/10253890.2021.1871600>



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Published online: 01 Feb 2021.



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Sex- and stress-dependent effects of a single injection of ketamine on open field and forced swim behavior

Paul J. Fitzgerald^{a*}, Savannah K. Kounelis-Wuillaume^{b*}, Ali Gheidi^a, Jonathan D. Morrow^a, Joanna L. Spencer-Segal^{b,c} and Brendon O. Watson^a

^aDepartment of Psychiatry, Michigan Medicine, University of Michigan, Ann Arbor, MI, USA; ^bMichigan Neuroscience Institute, Michigan Medicine, University of Michigan, Ann Arbor, MI, USA; ^cDepartment of Internal Medicine, Michigan Medicine, University of Michigan, Ann Arbor, MI, USA

ABSTRACT

Ketamine has emerged as a novel treatment for common psychiatric conditions such as Major Depressive Disorder (MDD) and anxiety disorders, many of which can be initiated and exacerbated by psychological stress. Sex differences in the frequency of both anxiety and depressive disorders are well known and could be due to sex differences in neuroendocrine responses to stress. Ketamine is known to modulate the hormonal response to stress, specifically corticosterone. It is not clear if the acute effect of ketamine on corticosterone differs by sex, or what role this could play in subsequent behavior. Here we test whether a single injection of (*R,S*)-ketamine (30 mg/kg, i.p.), administered either with or without unpredictable chronic stress (UCS), has different sustained effects on open field test (OFT), elevated zero maze (EZM) or forced swim test (FST) behavior in female versus male C57BL/6J mice. In the OFT (24 h post-injection), ketamine increased center square exploration in males but not females. In contrast, in the FST (72 h post-injection), females showed a trend toward a decrease in immobility after ketamine whereas males were not strongly modulated. These behavioral effects of ketamine were stronger in the presence of UCS than in unstressed animals. UCS animals also showed lower corticosterone after injection than unstressed animals, and in the presence of UCS ketamine increased corticosterone; these effects were similar in both sexes. Corticosterone post-injection did not predict subsequent behavior. These findings complement a growing preclinical literature suggesting both stress-dependency and sex differences in OFT and FST behavioral responses to ketamine.

LAY SUMMARY

- In humans, it is known that major depression and anxiety disorders, which can be caused or made worse by exposure to psychological stress, occur roughly twice as frequently in women than in men, but the underpinnings of these effects are not well characterized. In the current study, we explored how sex interacts with stress and ketamine (a rapidly acting antidepressant) by assessing both open field and forced swim behavior in mice after chronic mild stress. We report the novel finding that male mice exhibit greater exploration of the aversive center square in the open field after ketamine, whereas females trended toward lower immobility (often interpreted as an antidepressant-like effect) in the forced swim test after this drug, and these effects were amplified by prior stress exposure.

ARTICLE HISTORY

Received 18 August 2020
Accepted 28 December 2020





KEYWORDS

Major depression; chronic mild stress; psychosocial stress; corticosterone; open field; forced swim

Introduction

There is now ample evidence that the anesthetic drug ketamine can rapidly improve symptoms of Major Depressive Disorder (MDD) in human patients when used at sub-anesthetic doses (Berman et al., 2000; Krystal et al., 2019; Wei et al., 2020; Zarate et al., 2006). However, potential sex differences in the effects of this drug are not well understood. One recent randomized study of treatment-resistant depression has suggested that there may *not* be significant sex differences in the therapeutic properties of ketamine in humans (Freeman et al., 2019), but a growing preclinical literature has

found a number of sex differences in FST behavior in mice and rats that were given this drug. Several of these studies have suggested that female mice and rats are more behaviorally responsive than males to ketamine, showing benefits at lower doses (Carrier & Kabbaj, 2013; Franceschelli et al., 2015; Sarkar & Kabbaj, 2016) and with more rapid onset but a shorter duration (Franceschelli et al., 2015). Potential sex differences in OFT behavior after a single injection of ketamine are somewhat less well studied than FST behavior [but see: (Franceschelli et al., 2015; Kara et al., 2017; Thelen et al., 2016)].

CONTACT Joanna L. Spencer-Segal  sjoanna@med.umich.edu  Michigan Neuroscience Institute, Michigan Medicine, University of Michigan, Ann Arbor, MI 48109-1382, USA; Brendon O. Watson  brendonw@umich.edu  Department of Psychiatry, Michigan Medicine, University of Michigan, Ann Arbor, MI 48109-1382, USA

*Equal contribution.

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Psychological stress is well known to be a causal or exacerbating factor in MDD and various anxiety disorders (Bonde et al., 2016; Costa e Silva & Steffen, 2019; Hosang et al., 2014; Lautarescu et al., 2020). It is also well established that women are roughly twice as likely as men to be diagnosed with MDD, but the hormonal and neurochemical basis (among other potential factors) for this discrepancy, including sex differences in response to various antidepressant drugs, is not well understood (Albert, 2015; Baxter et al., 2014; Whiteford et al., 2013). Women also suffer from anxiety disorders, such as specific phobias and generalized anxiety disorder, at higher rates than men (Baxter et al., 2014; McLean et al., 2011). As with MDD, the neurochemical and neurohormonal factors that may give rise to sex differences in anxiety disorders, including their stress-sensitivity, are not well characterized at this time.

Any sex differences in ketamine's effects on FST or OFT behaviors could be mediated by neuroendocrine factors. For example, it has been shown that ovariectomized female rats no longer exhibit favorable responses to ketamine, suggesting sex hormones such as estrogen and progesterone may be necessary for the therapeutic effects of this drug in females (Carrier & Kabbaj, 2013). Another candidate molecule for modulating the behavioral effects of ketamine in rodents is the adrenal corticosteroid, corticosterone. Ketamine has been shown to acutely boost corticosterone in the absence of chronic stress (Fahringer et al., 1974; Kennett Radford et al., 2018; Kudo et al., 1993; Nistico et al., 1978). Stress itself alters hypothalamic–pituitary–adrenal (HPA) axis dynamics, and so any difference in the effect of ketamine in stressed or non-stressed states could be mediated by its interaction with glucocorticoids.

In a previous study (Fitzgerald et al., 2019) we found that a single injection of low-dose ketamine (30 mg/kg, i.p.) decreased immobility in the FST 24 h after injection in unpredictable chronic stress (UCS)-exposed male mice, but not in unstressed animals. The goal of the current study was to determine whether ketamine has sex- and stress-specific effects on various behaviors using several classic measures of exploratory behavior (OFT, EZM) as well as the FST, and whether differential stimulation of corticosterone could explain these differences.

Methods

Subjects

Sixty-four ($n = 8$ per sex/stress/drug cohort) experimentally naïve adult (8 to 9-weeks-old upon arrival) C57BL/6J mice (32 females and 32 males) were obtained from The Jackson Laboratory (Bar Harbor, ME). Starting the day of arrival and throughout the experiments, mice were either group-housed (unstressed mice) or single-housed (unpredictable chronic stress (UCS)) in cages within a humidity- and temperature-controlled vivarium and kept on a 12:12 h light/dark cycle (lights on at 6 am) with ad libitum access to food and water. On the day of arrival at the facility, all mice were subjected to a 5-min locomotion test in a 30-cm square box under dim lighting conditions (30 lux) to assess basal locomotion. Mice

were then divided into groups of eight (as noted above) that were matched for total distance traveled, to control for baseline variability in locomotion. All experiments were carried out in the daytime and during the light phase. All procedures were conducted at the University of Michigan and were performed in accordance with the guidelines and regulations set forth by the National Institutes of Health and the University of Michigan, with approval from its Institutional Animal Care and Use Committee (Protocol number: PRO00007803).

Unpredictable chronic stress

Mice were chronically subjected to one of two behavioral procedures: unstressed/standard procedures or UCS. Unstressed procedures consisted of group housing of the mice (4 per cage) on the day of arrival in the vivarium and throughout experimentation, handling (i.e., allowing the mouse to explore the experimenter's gloved hand and covered forearm) for ~30 s a day for the first 5 days after arrival, and maintenance of standard levels of cage enrichment with a single package of nesting material (Enviropak, Lab Supply, Fort Worth, TX).

In contrast, mice subjected to UCS were singly housed on the day of arrival and throughout the experiments, were not handled by the experimenter in the first 5 days (and throughout the experiments, except when necessary), and their cages were not enriched with an Enviropak and were instead given two white nestlets. These UCS mice also received one stressor/hassle a day at a random time, beginning the day after arrival and lasting for 14 consecutive days (see S1 Table in Fitzgerald et al. (2019) for a description and the sequence of these stressors), completed the day of the ketamine injection and one day prior to the start of behavioral testing. These stressors consisted of a randomized sequence of cage tilting (20°, 2 h), homecage bedding change, cage change, turning ambient lights on and off repeatedly for 3 h, replacement of nest with two still-intact nestlets, placing the mouse in a novel empty cage for 1 h, and placing divider in homecage for 4 h. All mice received the same stressors at the same time, in the same sequence. Male and female mice were housed in separate rooms and stressed at different times in the same room. The experimental timeline is shown in Figure 1; all mice were weighed 2 days prior to injection with ketamine or vehicle.

Drug

(*R,S*)-ketamine hydrochloride (Ketalar, Par Pharmaceutical, Chestnut Ridge, NY) was stored in darkness at room temperature, and for administration was diluted in 0.9% physiological saline solution (vehicle) to a concentration of 3.0 mg/ml and injected intraperitoneally (i.p.), at a volume of 10 ml/kg to reach an injected dose of 30 mg/kg.

Corticosterone measurement

Thirty minutes after the ketamine or vehicle injection, mice were warmed for 1–2 min under an infrared lamp to dilate the tail veins. Each mouse was secured in a tail vein

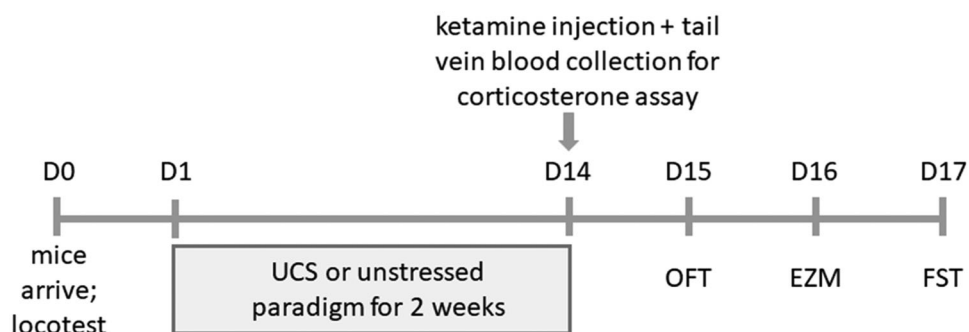


Figure 1. Experimental timeline. Mice arrived at the facility on Day 0. On Day 1, mice were subjected to either unstressed or unpredictable chronic stress (UCS) conditions composed of scheduled but varying daily hassles. Following this 2-week period of stress or no stress, all mice received a single injection (Day 14) of either vehicle (0.9% saline, i.p.) or 30 mg/kg ketamine. Thirty minutes after injection, tail vein blood was collected for corticosterone measurement. Twenty-four hours later (Day 15), mice were given an open field test (OFT), followed by an elevated zero maze (EZM) test on Day 16, and the forced swim test (FST) on Day 17.

restrainer (Braintree Scientific, Cambridge, MA), tail vein identified, and the area was cleaned with ethanol and incised using a razor blade. The tail vein was accessed using a small-gauge needle, and blood was collected using Microvette collection tubes (Braintree Scientific, Cambridge, MA) within 60 s of tail vein incision. Pressure was applied to achieve hemostasis before returning the mouse to its homecage. The blood samples were later centrifuged at 4000 rpm for 10 min to separate plasma. Corticosterone was measured from plasma using the DetectX Corticosterone Enzyme Immunoassay Kit (Arbor Assays, Ann Arbor, MI). One male mouse and two female mice had inadequate sample volume for analysis.

Behavioral testing

Beginning 24 h after ketamine injection, mice were subjected to daily behavioral tests in the following order: open field test (OFT), elevated zero maze (EZM), and forced swim test (FST); [Figure 1](#). Forced swim was performed last in the sequence to eliminate the possibility that this most stressful of the tests would affect the behavior of the mice in the OFT or EZM. The experimenter (SKK) was blind to the drug treatment groups, and all behavior was also scored in an automated fashion (see below). On each testing day, mice were allowed to acclimate to the testing room in their home cages for at least 30 min prior to testing. The open field consisted of a square box with sides 72 cm in length and walls 36 cm high. For analysis, the center region of the box floor was defined as 36 cm². The EZM (San Diego Instruments, Inc., San Diego, CA) was a circular track with an outside wall diameter 61 cm, an inside wall diameter 51 cm, and half of the track enclosed on both sides by 15-cm walls. Each mouse was allowed to explore the open field or elevated zero maze for 5 min under lighting conditions of 200 lux. At the end of testing, each mouse was immediately removed from the box or maze and returned to its homecage. The box or maze was cleaned with 50% ethanol solution between animals.

For the FST, up to four mice were tested simultaneously in a set of clear Plexiglas cylindrical tanks, 20 cm in diameter, filled halfway with water at 23–25 °C. Opaque plastic dividers were placed between the forced swim tanks to block the animals' view of one another. Ambient white lighting (200 lux) was present in the room during acclimation and throughout testing. Each trial lasted 6 min, but the behavior was only scored in the

last 5 min. At the end of the trial, the mouse was immediately removed from the tank, dried, and returned to its homecage. EthoVision version 11.5 was used for behavioral analysis (Noldus Information Technology, Leesburg, VA). For the FST, we defined “immobile” behavior in EthoVision as comprising bin-by-bin changes in mouse image pixelation of 0–10%.

Analysis and statistics

We analyzed the data with conventional parametric statistics (GraphPad Prism, GraphPad Software, La Jolla, CA). Two-way (stress × drug) or three-way (sex × stress × drug) analysis of variance (ANOVA) was used to assess general main effects and interactions ($\alpha = 0.05$). When there was a main effect of sex in the three-way ANOVA, we analyzed each sex separately with two-way ANOVA, followed by Tukey's post-hoc test when appropriate. Results are shown as mean ± standard error of the mean (SEM).

Results

To address how a single injection of ketamine influences behavior in stressed and unstressed animals of both sexes, 32 female and 32 male C57BL/6J mice underwent the experimental protocol shown in [Figure 1](#). Half of the mice were exposed to a 2-week protocol of unpredictable chronic stress (UCS) and half were not stressed. Following the completion of the stress protocol, mice were injected once with either 30 mg/kg (i.p.) ketamine or vehicle solution (0.9% saline). There were 8 mice per sex, drug, and stress-specific group. Outcome measures included a measure of corticosterone from tail vein blood 30 min after ketamine injection, to measure the effect of ketamine on the final product of HPA axis activation. Other outcome measures were OFT, EZM, and FST behavioral tests at 1, 2, and 3 days, respectively, post-injection.

Open field test

There were sex differences in open-field behavior independent of stress or drug group, with females showing greater general activity and exploratory behavior ([Figure 2](#)). Females traveled a greater distance in the arena than males [main effect of sex: $F(1, 56) = 13.80, p < 0.01$; [Figure 2\(A\)](#)] and in the center square [$F(1,$

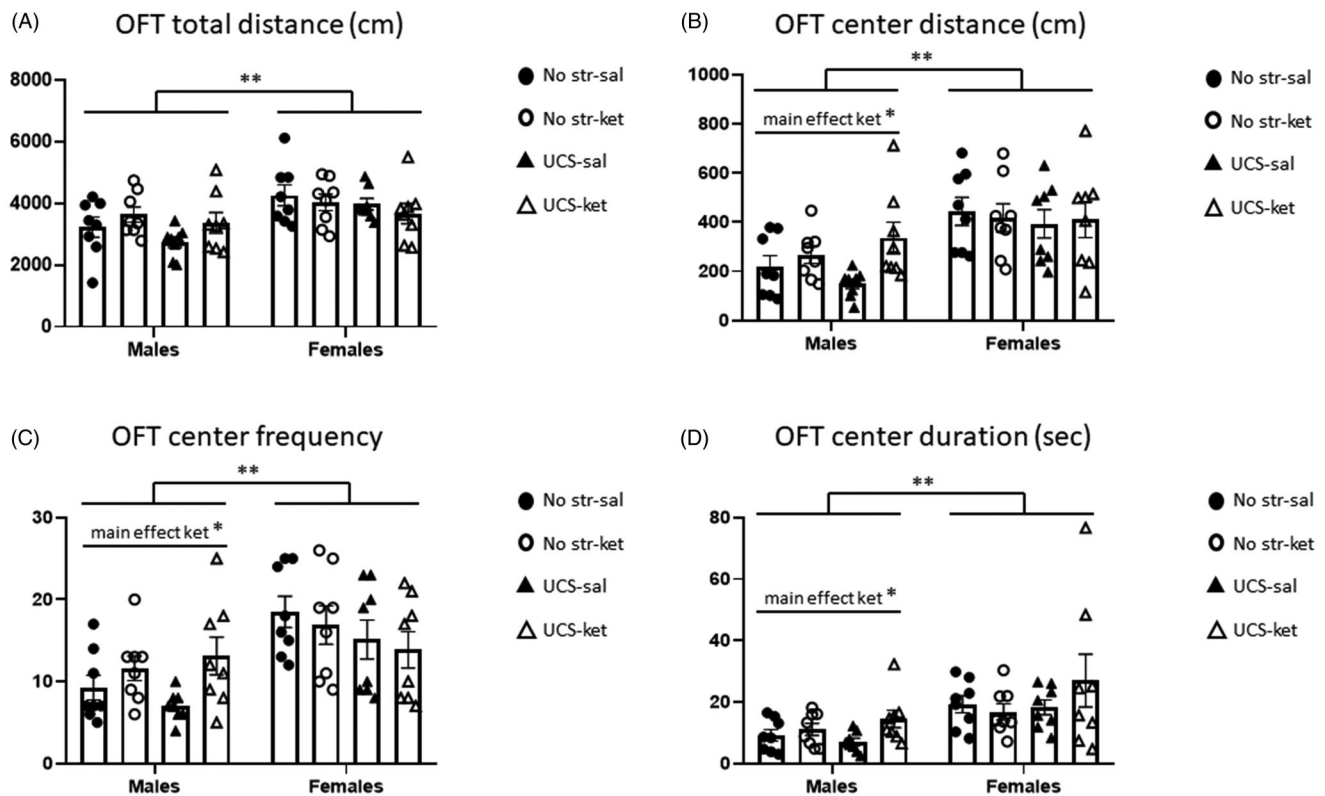


Figure 2. Male mice exhibited greater exploration of the center square when given ketamine after UCS in the open field test. Behavior (24 h post-injection) was automatically scored in EthoVision and parsed into: (A) total distance traveled; (B) center square distance; (C) center square frequency; (D) center square duration. Females were more active than males in all four measures. UCS-exposed males exhibited a significant, or trending, behavioral modulation after ketamine for measures (B–D) (see Table 1). Abbreviations: No str-sal: no stress saline; No str-ket: no stress ketamine; UCS-sal: unpredictable chronic stress saline; UCS-ket: unpredictable chronic stress ketamine. Error bars: \pm standard error of mean (SEM). Three-way ANOVA, main effect of sex: $**p < 0.01$, Two-way ANOVA, main effect of drug: $*p < 0.05$.

56) = 20.89, $p < 0.01$; Figure 2(B)], and females showed increased frequency in the center relative to males [$F(1, 56) = 18.29$, $p < 0.01$; Figure 2(C)] and duration [$F(1, 56) = 14.20$, $p < 0.01$; Figure 2(D)].

For OFT center distance, center frequency, and center duration, there were main effects of ketamine to increase exploration in males, irrespective of stress, but not in females [for males: center distance [$F(1, 28) = 7.27$, $p < 0.05$]; center frequency [$F(1, 28) = 7.05$, $p < 0.05$]; center duration [$F(1, 28) = 5.31$, $p < 0.05$]]. Post hoc tests in males (Table 1) showed that ketamine significantly increased center distance in the UCS but not the unstressed groups; the same trend was seen for center frequency and duration. The effect of ketamine on these three OFT measures in UCS males was substantial: center distance (saline mean = 149.2 cm, ketamine mean = 336.4 cm; 125.4% increase), center frequency (saline = 7.0, ketamine = 13.1; 87.5% increase), center duration (saline = 7.0 s, ketamine = 14.5 s; 106.5% increase).

In summary, in the OFT females were more active than males, and males but not females showed increased center square exploration after ketamine treatment, which was most pronounced in the UCS group.

Elevated zero maze

In the EZM (Figure 3), there were no sex differences and no significant modulation of any measure by ketamine. For EZM

total distance traveled (Figure 3(A)), there was no modulation of behavior by sex, stress, or drug (each $p > 0.05$). There was an overall effect of UCS to decrease open arm exploration compared to no-stress, seen as main effects of stress on open arm distance [$F(1, 56) = 4.60$, $p < 0.05$; Figure 3(B)], open arm frequency [$F(1, 56) = 7.45$, $p < 0.01$; Figure 3(C)], and open arm duration [$F(1, 56) = 6.92$, $p < 0.05$; Figure 3(D)].

In summary, UCS exposure tended to decrease open arm exploration in the EZM by suppressing open arm distance, frequency, and duration.

Forced swim test

In the FST, females had less immobility than males, when combining all stress and drug groups [main effect of sex: $F(1, 56) = 38.66$, $p < 0.01$] (Figure 4). Two-way ANOVA within each sex revealed that male mice did not exhibit modulation of immobility as a function of stress or drug. In contrast, stress trended toward a decrease in immobility in the females [main effect of stress: $F(1, 28) = 3.28$, $p = 0.081$], and females also showed a trend toward decreased immobility after ketamine independent of stress [main effect of drug: $F(1, 28) = 4.14$, $p = 0.052$]. UCS, ketamine-treated female mice showed the lowest immobility of all the groups.

Table 1. Post hoc comparison between ketamine and vehicle for various behavioral outcome measures.

Tukey's post-hoc comparison between ketamine and vehicle for various outcome measures		
	No stress <i>p</i> -value	UCS <i>p</i> -value
Figure 2(A): OFT total distance	0.73 (males)	0.39 (males)
Figure 2(B): OFT center distance	0.86 (males)	0.025 (males)
Figure 2(C): OFT center frequency	0.72 (males)	0.053 (males)
Figure 2(D): OFT center duration	0.91 (males)	0.067 (males)
Figure 4: FST immobility	0.84 (females)	0.19 (females)

In the open field test (OFT) and forced swim test (FST), a two-way (stress by drug) ANOVA was calculated for each sex, followed by Tukey's post-hoc test whose *p*-values are displayed here.

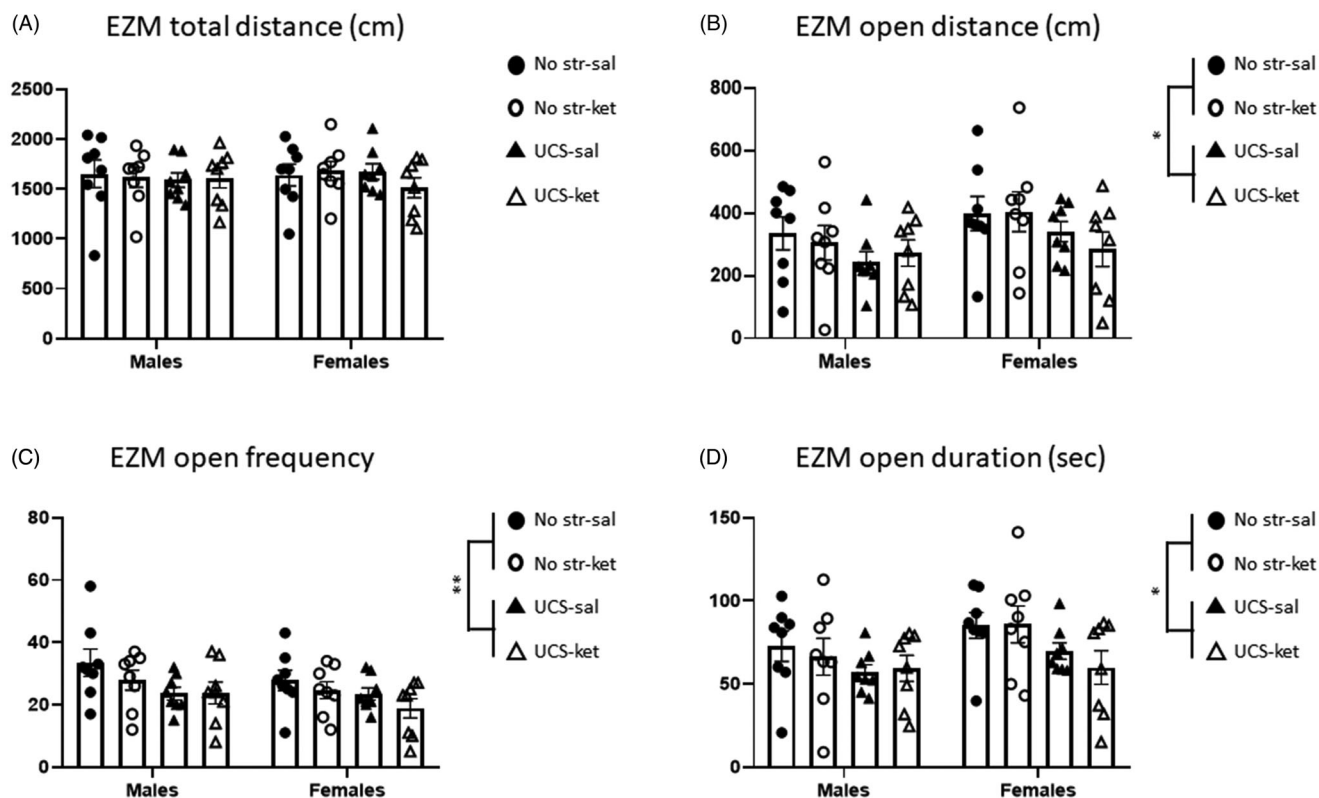


Figure 3. Ketamine did not modulate behavior in the elevated zero maze in either sex. Behavior (48 h post-injection) was automatically scored by computer and parsed into: (A) total distance traveled; (B) open arm distance; (C) open arm frequency; (D) open arm duration. UCS-exposed animals exhibited decreases in measures (B–D) irrespective of ketamine or sex. Error bars: \pm SEM. Three-way ANOVA, main effect of stress: * $p < 0.05$; ** $p < 0.01$.

In summary, in an FST 3 days post-injection, there was a tendency for a decrease in immobility for stressed females given ketamine, but not in males given ketamine.

Plasma corticosterone

To understand how stress and ketamine differentially affect the corticosterone response to acute injection, tail vein blood was collected 30 min after ketamine or vehicle injection (Figure 5), in the same mice for which behavior is described above. There were no sex differences in corticosterone. A three-way ANOVA (sex \times stress \times drug) revealed a main effect of stress, indicating that the UCS-exposed mice showed lower levels of corticosterone than unstressed mice after injection [$F(1, 53) = 7.33$, $p < 0.01$]. There was also a stress \times drug interaction [$F(1, 53) = 5.30$, $p < 0.05$]. Separate two-way ANOVAs for the no-stress and UCS groups separately showed no significant main effects or interaction for unstressed mice (each $p > 0.05$), whereas in UCS animals there was a trend toward ketamine increasing

corticosterone [main effect of the drug: $F(1, 26) = 3.64$, $p = 0.068$]. Corticosterone was much lower in stressed mice given saline injection than in the unstressed mice given saline, suggesting habituation of the stress response in the group that had been repeatedly stressed. Among stressed mice, the ketamine-injected mice had higher mean corticosterone than saline-injected mice, while the opposite was true for the unstressed groups. Upon Pearson correlation testing, there was no consistent animal-by-animal relationship found between corticosterone and any of the subsequent behavioral measures, either in the full sample or when analyzing the two sexes separately. Thus, plasma corticosterone measured 30 min post-injection was lower in stressed mice and did not predict subsequent behavior.

Discussion

In this study, we investigated the effects of sex and chronic stress history on corticosterone and subsequent behavior in response to a single subanesthetic dose of ketamine in

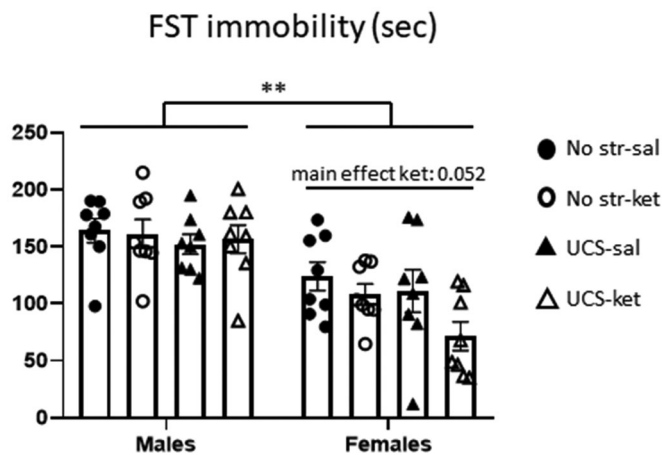


Figure 4. Female mice trended toward a decrease in immobility after ketamine in the forced swim test. Immobility behavior (72 h post-injection) was automatically scored. Females showed lower immobility than males. Females, especially those exposed to UCS, showed a trend toward a therapeutic-like response to ketamine. Error bars: \pm SEM. Three-way ANOVA, main effect of sex: $**p < 0.01$, Two-way ANOVA, main effect of drug: $*p = 0.052$.

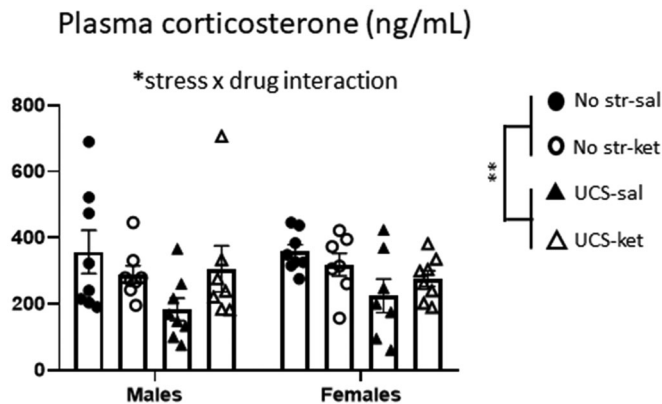


Figure 5. Chronic stress and ketamine interacted to modulate the corticosterone response to acute injection stress. Plasma corticosterone was measured 30 min after ketamine or vehicle injection, by tail vein puncture. Prior UCS exposure suppressed corticosterone irrespective of sex. There was also a significant stress by drug interaction. Error bars: \pm SEM. Three-way ANOVA, main effect of stress: $**p < 0.01$, stress \times drug interaction: $*p < 0.05$.

C57BL/6J mice. We found contrasting effects of sex, stress, and ketamine on behavior in the three different tests. Specifically, ketamine caused increased center exploration in the OFT in males only, regardless of stress history. In both sexes, stress decreased open arm exploration in the EZM, which was not reversed by ketamine. Ketamine tended to decrease immobility in females in the FST, a finding that is often interpreted as an antidepressant-like response. Finally, stress history modulated the effect of ketamine on corticosterone in a sex-independent manner, with ketamine tending to increase corticosterone in chronically stressed mice.

The effects of ketamine to increase center exploration in the OFT in males and to reduce immobility in the FST in females were strongest in the UCS group. This finding is in line with our previous studies (Fitzgerald et al., 2019; Polis et al., 2019) which showed that increased activity after ketamine in the FST tended to only be observed in C57BL/6J mice in the presence of UCS but not in unstressed animals.

While in those previous studies we actually found that unstressed mice showed an *opposing* response to ketamine relative to stress-exposed animals, the data we report here at least show that stress can amplify the therapeutic effect of this drug as assessed by the FST (and the OFT) relative to unstressed animals. It should be noted that the sex of the experimenter, as well as the brand of ketamine, are potential sources of variability in behavioral studies of this drug (Chapman et al., 2018; Fitzgerald et al., 2019; Georgiou et al., 2018). In this study, the FST and OFT experiments were carried out by a female experimenter while stress was carried out by a male experimenter – whereas, in our previous study, both were carried out by a male. We further suggest that the relatively modest effects that UCS produced in our three behavioral tests are consistent with the relative stress resilience of the C57BL/6J strain (Mehta & Schmauss, 2011; Millstein & Holmes, 2007; Shanks et al., 1990). We also point out that since the daily hassles in UCS mice did not extend into the 3 days of behavioral testing, although these tests themselves are stressors, the behavioral modulation (or lack thereof) that we observed after chronic stress may be attributed to stress cessation.

We are not the first research group to suggest that there are sex differences in the FST mobility behavior of rodents in response to low dose ketamine. For example, a pair of studies in Sprague Dawley rats found that, in the dosage range of 2.5–10 mg/kg, females respond to lower doses than males in the FST, with or without chronic stress (Carrier & Kabbaj, 2013; Sarkar & Kabbaj, 2016). Likewise, data in C57BL/6J mice from another group that used a similar dosage range has shown that females respond at a lower dose than males do 24 h post-injection (Franceschelli et al., 2015). Perhaps surprisingly, in an experiment using chronic mild stress, this group also found that males that had been given 10 mg/kg still responded at the 7-day time point, whereas females did not (Franceschelli et al., 2015). Thus, the existing literature suggests that females show decreased immobility after ketamine at lower doses, but that the duration of action may be longer in males. This latter finding may stand in contrast to our current result that showed females trended toward a decrease in immobility 72 h (i.e. 3 days) post-injection but males did not, although we used a higher dose (30 mg/kg) and did not test at the 7-day time point. We used 30 mg/kg as our standard dose here because we have previously shown that in male C57BL/6J mice, this dose (but not 10 mg/kg) has mobility enhancing properties in the 24-h post-injection FST in UCS-exposed (but not unstressed) mice (Fitzgerald et al., 2019; Polis et al., 2019).

Here we present the novel finding that stressed males (but not females) given 30 mg/kg ketamine showed approximately twice as much investigation of the center square as vehicle animals in the OFT. While these results could be confounded by generalized increases in locomotion in males given ketamine, there was no significant effect of ketamine on the total distance traveled in the OFT. Previous studies have examined sex differences in response to ketamine in the OFT at different time points. In C57BL/6J mice, 10 mg/kg ketamine was found to induce an acute (30 min after injection) reduction in center square exploration in female mice

irrespective of stress, whereas males were not affected by this drug or stress (Franceschelli et al., 2015). In a subsequent study, this group found that *chronic* ketamine can reduce center square exploration in the OFT in female mice without doing so in males (Thelen et al., 2016). A different group of researchers found that chronic administration of this drug to mice from the ICR strain did not modulate center square exploration in either males or females (Kara et al., 2017). Thus, the effect of ketamine on open field exploration may depend on the dose, timing, and chronicity of exposure in addition to the mouse strain. From our study, we conclude not that one sex is more or less sensitive to ketamine, but that their specific behavioral sensitivities to ketamine differ. The time point at which the specific tests are run, and the ketamine dose, are important variables. The sex differences in behavioral sensitivity to ketamine shown in this study could be due to fundamentally different effects of this compound on neurotransmitter signaling or neuroplasticity (Thelen et al., 2019).

In this study, there was a dissociation between the effects of UCS and ketamine on two different tests of novelty exploration, the OFT and EZM, which are often thought of as similar tests of anxiety-like behavior. We observed that UCS exposure produced decreased exploration of the open arms at the 48-h post-injection time point in the EZM, irrespective of ketamine and sex. We saw no similar overall effect of UCS on OFT behavior, and there were no ketamine effects in the EZM, unlike in the OFT. While the tests were done at different time points, we think this finding likely reflects important differences in the neural substrates underlying performance in these two tests. It should also be noted that the mice exhibited relatively greater avoidance of the OFT center square than the EZM open arms. While this could be related to the inherent geometry of the two mazes, another possibility is that the timing of the two tests relative to injection and tail vein puncture, with OFT carried out first, influenced these behavioral outcomes.

In our measurement of plasma corticosterone 30 min post-injection, we found that stressed mice had lower corticosterone after a saline injection relative to unstressed mice. This, most likely, represents a blunted HPA axis response to the injection itself in the stressed mice, although we did not carry out baseline corticosterone measurement prior to injection. This could reflect neuroendocrine habituation of these mice to a novel stressor after UCS, or another mechanism; for example, changes in noradrenergic tone as a result of chronic stress. Ketamine-injected mice showed similar corticosterone levels in both stressed and unstressed groups. The mean corticosterone level was higher in stressed mice after ketamine injection as compared to saline, implying stimulation of the HPA axis by ketamine in this group. It could be that this effect was not seen in the unstressed mice due to a “ceiling” effect; that is, maximal stimulation of the HPA axis by the saline injection alone. It may also be that ketamine acutely affects the HPA axis differently in stressed and non-stressed animals. These findings do not specifically support the idea that acute ketamine-induced corticosterone changes mediate the behavioral effects of ketamine. This lack of mediation is particularly supported by the lack of correlation between

post-injection corticosterone and the subsequent behavioral measures. However, these findings do support differences in HPA axis regulation in the UCS animals compared to the unstressed condition. While we also find that ketamine affects behavior differently in UCS and unstressed animals, the role of central HPA axis activity or glucocorticoids in this phenomenon remains to be explored.

The sex differences we observed in the effect of ketamine on behavior could suggest that ketamine modulates different neural circuitry in males and females. Alternatively, it could modulate similar neural circuits but this circuitry is sexually dimorphic in its behavioral output. In either case, these findings potentially have translational relevance to our understanding of human affective disorders. These data may also suggest that any putative sex differences in the therapeutic properties of ketamine are not mediated by upstream, acute effects on HPA axis responsivity, as we found this to be similar in our male and female animals. The possibility of sex differences in the modulation of the neural circuitry underlying anxiety disorders versus MDD should be addressed in future clinical studies as it may be relevant to the clinical use of this drug (Chen et al., 2014; Coyle & Laws, 2015; Niciu et al., 2014). Our data also lend support to a growing body of evidence suggesting that the time delay between administration of ketamine and the onset and continuation of its therapeutic-like effects is of critical importance and may differ across sexes (Fitzgerald et al., 2019; Franceschelli et al., 2015).

In conclusion, here we have shown novel, stress-dependent sex differences in behavior in response to a single injection of subanesthetic dose ketamine in C57BL/6J mice. We found that the male mice exhibited an increase in center square exploration after ketamine in the OFT (24 h post-injection) which was more pronounced in the stressed group, whereas females showed a trend toward decreased immobility in the FST (72 h post-injection) which was also more pronounced in the stressed group. Ketamine appeared to stimulate corticosterone in the UCS group, while this was not seen in unstressed animals perhaps due to maximal stimulation by the injection itself. The ketamine-stimulated corticosterone did not predict subsequent behavior. While future studies may further elucidate the neural mechanisms underlying these effects, such as sex-specific modulation of glutamatergic signaling in the medial prefrontal cortex (Thelen et al., 2019), it should be noted that pharmacokinetic differences may play a role (Saland & Kabbaj, 2018). Our findings here describing stress-sensitivity reinforce that the effects of antidepressant medications may differ depending on stress history, dovetailing with our previous work (Fitzgerald et al., 2019; Polis et al., 2019). More generally, the discrepancies reported in the literature on the behavioral effects of ketamine in rodents – as a function of sex, strain, chronic stress, dose, behavioral test, testing lag – suggest that the field should seek clearer, more consistent biological measures of ketamine efficacy.

Our data on a single injection of this drug also complement an emerging literature on rodent ketamine sex differences (or similarities, including investigations of females instead of just males) with respect to: chronic dosing of ketamine (Thelen et al., 2016), ketamine enantiomers (Chang et al.,

2018), drug self-administration or addictive behaviors (Schoepfer et al., 2019; Wright et al., 2019; Wright & Kabbaj, 2018), hyperactivity (McDougall et al., 2019; Wilson et al., 2005, 2007), neurohormonal regulation (Dossat et al., 2018; Picard et al., 2019; Saland et al., 2016; Wright et al., 2017), fear conditioning (Mastrodonato et al., 2018), and synaptic mechanisms (Sarkar & Kabbaj, 2016; Strong et al., 2017; 2019). These previous studies, as well as our data here and the literature described earlier in the Discussion, highlight a variety of sex differences in response to ketamine in mice and rats, and collectively underscore the importance of gaining a greater understanding of the neural mechanisms that mediate these sex differences and how they may translate to human psychopathology.

Geolocation information

All authors are located in Ann Arbor, Michigan, USA. This is where all research was carried out.

Acknowledgments

We thank Anuska Martinez Argueda, Jessica Babel, Andrew Drumheller, Tiffany Lapworth, Benjamin Stefadu, Brookelyn Wheeler, and Yuhuan Ye for their assistance with this study.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

BW: [NIH K08 MH107662], Collaboration Grant from The Kavli Neuroscience Innovators at University of Michigan, Neuroscience Fellows at the University of Michigan, Frances and Kenneth Eisenberg Scholar Award, The Taubman Emerging Scholars Award at the University of Michigan, The Pritzker Neuropsychiatric Disorders Research Consortium; JSS: [NIH K08 MH116267], NARSAD Young Investigator Grant from the Brain and Behavior Research Foundation, Collaboration Grant from The Kavli Neuroscience Innovators at University of Michigan; PF: University of Michigan Depression Center STAR Award; AG: National Institute on Drug Abuse (NIDA) [T32 DA07268]; JM: National Institute on Drug Abuse (NIDA) [R01 DA044961], Frances and Kenneth Eisenberg Scholar Award, The Taubman Emerging Scholars Award at the University of Michigan.

Data availability statement

We will make all data associated with this study freely available upon publication acceptance.

Notes on contributors

Paul J. Fitzgerald is a neuroscience postdoc working in Brendon Watson's lab at the University of Michigan.

Savannah K. Kounelis-Wuillaume is now an MD/PhD student at the Uniformed Services University of the Health Sciences.

Ali Gheidi is now a Research Fellow at Wayne State University.

Jonathan D. Morrow is an Associate Professor in the Department of Psychiatry at the University of Michigan.

Joanna L. Spencer-Segal is an Assistant Professor in the Department of Internal Medicine at the University of Michigan.

Brendon O. Watson is an Assistant Professor in the Department of Psychiatry at the University of Michigan.

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