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RESEARCH LETTER

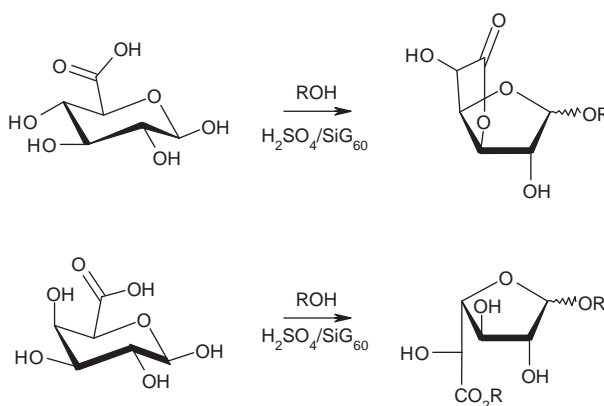
Efficient microwave-promoted synthesis of glucuronic and galacturonic acid derivatives using sulfuric acid impregnated on silica

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Monomode microwave-assisted syntheses of D-glucuronic and D-galacturonic acid derivatives are reported in the presence of a solid acid catalyst, consisting of sulfuric acid loaded onto silica. This approach affords a variety of surface-active monoglycosylated glucofuranosidurono-6,3-lactones and disubstituted galacturonic adducts in excellent yields in less than 10 min at 85°C. This study illustrates the application of microwave heating mode, in combination with a cost-effective solid catalyst, as an efficient, selective, and eco-friendly methodology in carbohydrate chemistry.



1 step, microwave - 60–85°C, 1–10 min

Keywords: microwave chemistry; carbohydrates; heterogeneous catalysis; silica; supported catalysts

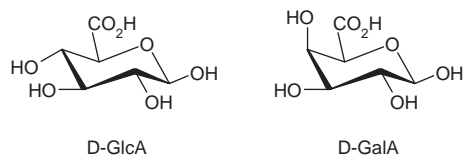
1. Introduction

In the context of the valorization of carbohydrates arising from agricultural resources (i.e. lignocellulosic feedstocks) (1), the quest of structurally homogeneous amphiphilic materials is one of the most developed research directions (2,3). Among these carbohydrate-based biodegradable amphiphilic molecules, derivatives of D-glucuronic (D-GlcA) and D-galacturonic (D-GalA) acids, with a single or two alkyl chains, are efficient, biocompatible and bioresorbable, nonionic surface-active reagents (4–11). A panel of other synthetical uronic acid-containing

entities find also special relevance in the design of bioactive molecules (12–16) as in supramolecular and materials chemistry (17–23).

D-GlcA and D-GalA are naturally occurring hexuronic acids that are derived from renewable raw materials such as hemicellulose (i.e. glucurono-arabinogalactans) and pectins (24,25). Containing two appropriate functional sites (anomeric hydroxyl and carboxylic acid) (Scheme 1), these monosaccharides are suited starting material for reactions with nucleophilic agents (alcohols, amines or thiols) (26). However, due to the same reactivity of both functional

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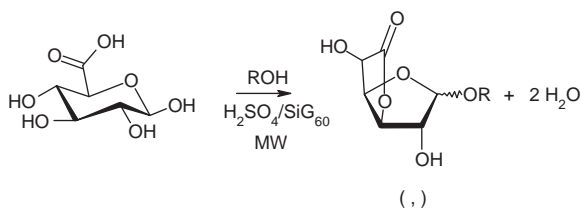


Scheme 1. Structures of β -D-glucuronic acid (D-GlcA) and β -D-galacturonic acid (D-GalA).

sites, the regioselective production of well-defined uronic acid derivatives, without generation of side-products, requires systematic multi-step pathways involving suitable protection, activation, and deprotection sequences (27,28). In this context, the development of new sustainable, one-step and efficient strategies for the synthesis of anomerically pure uronic acid-based products is a particularly challenging task (10,29,30).

Previously, we disclosed that the tandem use of microwaves (MW) and heterogeneous catalysts could offer a convenient opportunity for the direct formation of uronic acid derivatives (31). Particularly, we reported on the application of sulfuric acid impregnated onto silica gel ($\text{H}_2\text{SO}_4/\text{SiG}_{60}$) to convert quantitatively D-GlcA into monosubstituted β -D-glucofuranosidurono-6,3-lactones in a single step (Scheme 2). Reactions in closed vessels proceeded, in high yields and selectivities, after a few minutes (1–10 min) of microwave heating at 85°C with a maximum power of 80 Watt, which contrasted with previous works involving homogeneous Lewis acid conditions and extended reaction times (7,10,30). Moreover, it was notable that our solvent-free protocol presented attractive practical features, as easy set-up and convenient catalyst recovery (by filtration or centrifugation). The catalyst could also be reused for several consecutive batch reactions if carefully dried under static air at 160°C . Faced to environmental concerns, we anticipate herein that this approach may constitute a promising and low waste route to a range of new original tensioactive products.

As a part of our ongoing program dedicated to the elaboration of new chemical methodologies for the formation of high added-value materials from carbohydrates (31,32), we evaluate the applicability



Scheme 2. Microwave-promoted synthesis of monosubstituted lactones from glucuronic acid and alcohol (ROH).

of $\text{H}_2\text{SO}_4/\text{SiG}_{60}$ to promote the “one-pot” modification of unprotected acidic sugars under microwave conditions.

2. Results and discussion

2.1. Microwave-assisted reaction between D-GlcA and various nucleophilic agents

Microwave-induced transformations were typically performed between 60 and 85°C , using a CEM Discover reactor, with a maximum power of 80 Watt under closed vessel conditions. Real-time temperature within the reaction vial was monitored using an infrared detector. In a first set of trial, D-GlcA was coupled with fivefold excess of alcohol in the presence of 35 wt% $\text{H}_2\text{SO}_4/\text{SiG}_{60}$ (Table 1). We observed that, whatever the nature of the alcohol (ROH) investigated, D-GlcA conversions were nearly quantitative. After filtration of the solid catalyst through a glass frit and elimination of the excess of alcohol, the corresponding monosubstituted D-glucofuranosidurono-6,3-lactones **1–16** were recovered in good yields, mainly as the β anomer. No traces of disubstituted adducts (adducts where both the anomeric and the acid functions have been functionalized) and/or D-glucuronic acid γ -lactone were detected after microwave exposure, conversely to results obtained under classical thermally driven conditions (31). Structures of isolated products and anomeric configurations were unambiguously assigned by NMR (Nuclear Magnetic Resonance) spectroscopy. These monosubstituted D-glucofuranosidurono-6,3-lactones are suited candidates for applications as surface-active agents (33).

Microwave-promoted reactions between D-glucuronic acid and alkyl alcohols afforded in almost quantitative yields the corresponding alkyl D-glucofuranosidurono-6,3-lactones **1–6** with high anomeric selectivities ($\alpha:\beta = 10:90$) (31). Likewise, derivatisation of D-glucuronic acid with unsaturated moieties (such as allyl, butenyl, hexenyl, and undecenyl groups), in order to generate a new set of potent surface-active-agents, was conveniently achieved using our microwave strategy. Vinyl chains were selectively appended to the anomeric position in less than 10 min and without the recourse to solvent. Final lactonic products **7–10** were characterized by a typical ^1H NMR chemical shift around δ 5.04 ppm, associated to a lowfield resonance for the anomeric carbon at δ 109 ppm, clearly confirming the formation of the β -adduct. Incorporation of the vinyl fragment onto the sugar structure was also corroborated by ^{13}C NMR signals around 135 and 115 ppm. Monosubstituted α -D-glucofuranosidurono-6,3-lactone was recovered as a minor by-product.

Table 1. Microwave-assisted reaction between totally un-protected D-glucuronic acid and various nucleophilic agents in the presence of 35 wt% H₂SO₄/SiG₆₀.^a

| Nucleophile | Product | Overall yield (%) ^b | α : β ratio ^c |
|--|--------------|--------------------------------|---------------------------------------|
| <i>ROH</i> (alcohols) | | | |
| MeOH ^d | 1 | 83 | 10:90 |
| CH ₃ (CH ₂) ₃ OH ^d | 2 | 98 | 10:90 |
| CH ₃ (CH ₂) ₅ OH ^d | 3 | 96 | 10:90 |
| CH ₃ (CH ₂) ₇ OH ^d | 4 | 97 | 10:90 |
| CH ₃ (CH ₂) ₁₁ OH ^d | 5 | 62 (68 ^e) | 10:90 (20:80 ^e) |
| Isopropanol ^d | 6 | 87 | 5:95 |
| Allyl alcohol | 7 | 86 | 25:75 |
| 3-Buten-1-ol | 8 | 98 | 5:95 |
| 5-Hexen-1-ol | 9 | 99 | 15:85 |
| 10-Undecen-1-ol | 10 | 92 | 5:95 |
| Chloropropanol | 11 | 99 | 10:90 |
| Benzyl alcohol | 12 | 70 ^f | 25:75 |
| Phenol ^f | Not isolated | <5 | – |
| <i>p</i> -Cresol | Not isolated | <5 | – |
| Cyclohexanol | 13 | 95 | 15:85 |
| 1,6-Hexanediol | 14 | 92 | 30:70 |
| Propargyl alcohol | 15 | 83 | 25:75 |
| PEG200 | 16 | 99 (98 ^g) | 15:85 (15:85 ^g) |
| <i>RSH</i> (thiols) | | | |
| CH ₃ (CH ₂) ₇ SH | 17 | 97 | Pure β |
| CH ₃ (CH ₂) ₁₃ SH | 18 | 68 ^f | Pure β |

^aReaction conditions: GlcA (200 mg, 1.03 mmol), ROH or RSH (5 equiv.), and the catalyst (50 mg) were introduced in a 10 mL microwave reaction vial and irradiated, under agitation, at 85°C, during 10 min, and with a maximum power of 80 Watt. Commercial anhydrous alcohols were preferably selected for this study.

^bIsolated yields of pure anomeric mixtures.

^cAnomeric compositions of reaction mixtures were determined by ¹H NMR spectroscopy.

^dCompounds 1–6 are described in Ref. (31,32).

^eReaction performed in *tert*-butanol used as the solvent.

^fReaction was performed at 60°C during 10 min with a maximum microwave power of 10 Watt.

^gOverall yield and anomeric distribution after a reaction time of 2 min at 85°C.

The feasibility of our microwave-assisted procedure was also underlined for the functionalization of D-GlcA with other representative alcohols: chloropropanol, benzyl alcohol, phenol, 1,6-hexanediol, and propargyl alcohol. In most cases, syntheses were conveniently achieved, providing in good yields the corresponding β -adducts **11**–**15** as the dominant forms. Noteworthy, glycosylation between two sugar units and/or formation of diglycosylated adducts (for reaction with 1,6-hexanediol) were undetected.

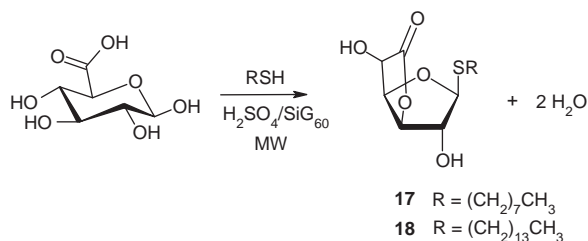
Derivatization of the starting D-GlcA with phenol was not successful, mainly because of the fast oxidation of phenol under reaction conditions. The same conclusion was formulated when using *p*-cresol.

Finally, glycosylation of polyethylene glycol afforded quantitatively, after 10 min in the microwave reactor, the corresponding poly(ethylene glycol)-D-glucofuranosidurono-6,3-lactone **16** (α : β = 15:85). This reaction proceeded very quickly in less than 2 min at 85°C and enough for the quantitative conversion of D-GlcA. On the other hand, we demonstrated that the initial amount of PEG200 could noticeably be reduced to two equivalents without affecting both overall yield and anomeric selectivity of the reaction. However, work with an equimolar sugar/PEG ratio was unrealizable as decomposition of D-GlcA, by browning reaction, was observed.

Yields for compounds **7**, **12**, and **15** were lower than for other products, logically because of the lower reactivity of allyl, benzyl, and propargyl alcohols. This impacts also over the anomeric selectivity as a 25/75 α / β ratio was recorded for these three final lactones. Independently from reactivity concerns, the functionalization of D-GlcA with benzyl alcohol or PEG200 is particularly noteworthy. Indeed, only for these two specific alcohols, we observed experimentally a decomposition of the sample by browning reaction if extended reaction times (up to >10 min) and/or higher temperatures were applied. As previously mentioned by Gabriel et al. (34), the kinematic viscosity and the loss tangent value ($\tan \delta$) are two relevant parameters for microwave applications.¹ For linear alkyl alcohols, as butanol, hexanol or octanol, the kinematic viscosity is ranging between 2.3 and 7.7 cP (at 25°C), while the loss tangent value decrease when increasing chain length ($\tan \delta = 0.571$, 0.344, and 0.181, respectively, for butanol, hexanol, and octanol). For benzyl alcohol and PEG200, these viscosity values attain, respectively, 8.0 cP and 50.0 cP and are associated to $\tan \delta$ values of, respectively, 0.667 and >1.0. These high $\tan \delta$ mean that both solvents are able to convert very efficiently microwave energy into heat energy and to transfer it quickly to reagents, leading probably to D-GlcA charring under viscous conditions.

Finally, our approach was also extended to the functionalization of D-GlcA by alkyl thiols. We notably demonstrated that less than 5 min at 60°C was enough to recover compounds **17** and **18** in, respectively, 97 and 68% yields, exclusively as β -anomer (Scheme 3).

These reactions are in agreement with the guidelines of green chemistry, as the application of microwave as an alternative heating source leads to improved yields and unprecedented selectivities. Moreover, our solvent-free protocol has a low environmental impact as water is the only side-product generated and as the catalyst can easily be recovered.



Scheme 3. Microwave-mediated functionalization of glucuronic acid by alkyl thiols (RSH).

2.2. Microwave-assisted functionalization of mixtures of D-GlcA and D-glucose

In order to substantially reduce both economical and environmental impacts related to the separation and the purification of sugars after hydrolysis of raw lignocellulosic materials, we proposed that our microwave strategy could also be exploited for the production of mixtures of surfactants. Thus, we considered the simultaneous benchmark derivatisation of D-glucose and D-glucuronic acid with *n*-octanol (Scheme 4). We anticipated that a synergy between both tensioactive molecules could be highlighted (35,36). An equimolar mixture of both sugars and catalytic amounts of 35 wt% H₂SO₄/SiG₆₀ in a fivefold excess of *n*-octanol was exposed to microwaves for 10 min at 85°C. NMR analysis of the crude reaction mixture, recovered after filtration of the solid acid promoter and elimination of the excess of alcohol, indicated the presence of octyl D-glucofuranosiduro-6,3-lactone **4** (98% yield based on D-glucose conversion, $\alpha:\beta = 5:95$) and octyl D-glucopyranoside **19** (89% yield, $\alpha:\beta = 30:70$). This result is quite interesting as it illustrates how silica sulfuric acid, in combination with microwaves, was able to catalyze efficiently and independently the alkylation of both sugars in high yields. As a fact, no evidence of a direct attachment between both sugars was underlined using two-dimensional NMR techniques.

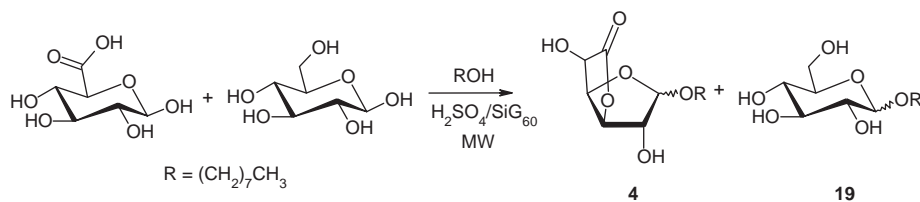
2.3. Extension of the microwave-assisted protocol to D-galacturonic acid

The microwave-assisted protocol developed previously was applied to D-galacturonic acid (Table 2). Reactions performed under microwave exposure

provided the corresponding functionalized galactosiduronates, mainly as the kinetically favored furanosidic isomers (Scheme 5). In this case, yields reached up to 85% and contrasted with previous work, involving extended reaction times and recourse to vacuum techniques (37). Unequivocal evidence of the predominant formation of α,β -furanosidic forms was supported by ¹³C NMR analyses (in combination with homonuclear and heteronuclear correlation experiments) with signals for C-4 around 83–85 ppm, more deshielded than in α,β -pyranuronate compounds ($\delta_{\text{C-4}}$ typically close to 69–71 ppm) (38,39).

3. Experimental

D-glucuronic acid ($\geq 98\%$), D-(+)-galacturonic acid monohydrate ($\geq 97\%$), anhydrous D-(+)-glucose, and linear poly(ethylene glycol) ($M_n = 200$ g/mol) were purchased from commercial suppliers and used as received. Silica gel 60 was calcinated at 200°C prior to use. Solvents were dried using well established procedures. Thin-layer chromatography was performed on silica gel 60 plates (with fluorescent indicator F254) with visualization by charring with 10% H₂SO₄/ethanol solution. Preparative column chromatography was achieved using 230–400 mesh ASTM silica gel. ¹H and ¹³C NMR, as two-dimensional spectra (gDQ-COSY, gHMQC-AD, gHMBC-AD, and NOESY), were recorded at 25°C on a Varian 600 MHz spectrometer. Chemical shifts are listed in parts per million downfield from TMS (tetramethylsilane) and are referenced by the residual solvent peaks. Infrared spectra were recorded at 4.0 cm⁻¹ resolution on a Bruker IFS-48 FT-IR spectrometer. Melting points were estimated with a Büchi[®] melting point apparatus and were uncorrected. Microwave-assisted reactions were performed in specific 10 mL microwave reaction vials and irradiated in a CEM Discover[®] microwave reactor, instrument as KBr (potassium bromide) pellets. Mass spectra were acquired on an Esquire HCT high capacity ion trap mass spectrometer (Bruker Daltonics, Bremen, Germany) equipped with an electrospray ionization source. Best results were obtained in positive mode with capillary, nebulizer pressure, drying gas flow,



Scheme 4. Simultaneous derivatisation of glucuronic acid and glucose.

Table 2. Microwave-assisted reaction between totally unprotected D-galacturonic acid and various alcohols in the presence of 35 wt% H₂SO₄/SiG₆₀.^a

| ROH | Product | Yield (%) ^b | |
|--|-----------|-----------------------------|-----------------------------|
| | | Pyranose (α:β) ^c | Furanose (α:β) ^c |
| MeOH | 20 | – | 89 (20:80) |
| CH ₃ (CH ₂) ₃ OH | 21 | m | 85 (15:85) |
| CH ₃ (CH ₂) ₅ OH | 22 | m | 86 (20:80) |
| CH ₃ (CH ₂) ₇ OH | 23 | m | 49 (15:85) |
| poly(ethylene glycol) | 24 | 30 (10:90) | 69 (40:60) |

^aReaction conditions: GalA.H₂O (220 mg, 1.03 mmol of GalA), ROH (5 equiv.), and the catalyst (50 mg) were introduced in a 10 mL microwave reaction vial and irradiated, under agitation, at 85°C, during 10 min, and with a maximum power of 80 Watt.

^bIsolated yields of pure isomeric mixtures. *m* stands for minute amounts.

^cAnomeric compositions were determined by ¹H NMR spectroscopy.

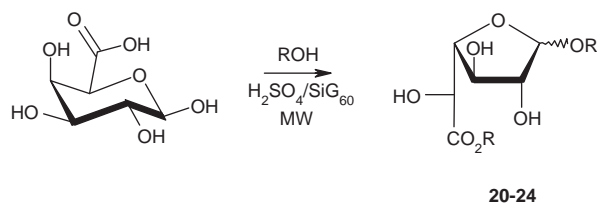
and drying gas temperature set to 10,000 nA, 40.0 psi (nitrogen), 9.0 L min⁻¹ (nitrogen), and 365°C, respectively. The scan range was adjusted to *m/z* 100–500 and the target mass set to 300. MS-data were recorded and processed using Bruker Daltonics data analysis software, version 3.0. All the mass spectra were acquired in ESI (electrospray ionization) in the positive mode.

3.1. Preparation of H₂SO₄/SiG₆₀

The silica-supported sulfuric acid catalyst was prepared according to a well-established procedure (40) by incipient wetness impregnation of silica gel 60 (with BET specific surface area of 500 ± 50 m²/g and average pore diameter of 60 ± 5 Å) with an appropriated aqueous solution of H₂SO₄ yielding catalyst with an acid loading of 35 wt% (i.e. 35 g pure H₂SO₄/100 g final catalyst).

3.2. General procedure for the microwave-assisted synthesis and purification of compounds 1–24

Carbohydrates (D-GlcA, D-GalA, and D-glucose), alcohol/thiol (5 equiv.), and the acid catalyst were



Scheme 5. Microwave-synthesis of disubstituted galacturonic acid derivatives.

introduced together in a specific 10 mL pressure-resistant closed microwave vial. This vial was next placed within the microwave monomode cavity and irradiated under stirring. Control of the temperature during the experiment was assigned by an infrared detector. After heating, reactions were immediately quenched by forced compressed air cooling of the vial. Crude mixtures were filtrated to eliminate solid acid catalyst, washed with CHCl₃, and then classically chromatographed on silica gel. When possible, the alpha anomeric form was also isolated for at least NMR analysis.

Spectral and physico-chemical data for alkyl α,β-D-glucofuranosidurono-6,3-lactones **1–7** (31, (41–43), *n*-octyl glucoside **19** (44), methyl (methyl α,β-D-galactofuranosid)uronate **20** (43,45,46), butyl (butyl α,β-D-galactosid)uronate **21**, hexyl (hexyl α,β-D-galactosid)uronate **22**, and octyl (octyl α,β-D-galactosid)uronate **24**⁽³⁷⁾ were in agreement with those reported in the literature.

3.2.1. Allyl D-glucofuranosidurono-6,3-lactone 7

Mixture of α and β isomers: colorless oil. Only the β anomer was isolated. **7β**: off-white solid (recrystallised in light petroleum); mp 104–106°C. IR (KBr) ν 3440 cm⁻¹, 3017 cm⁻¹, 2976 cm⁻¹, 1758 cm⁻¹. ¹H NMR (600 MHz, D₂O) δ 5.72 (m, 1H, OCH₂CH=CH₂), 5.23 (m, 2H, OCH₂CH=CH₂), 5.08 (s, 1H, H-1), 4.92 (dd, 1H, *J*_{3,4} 4.8 Hz, *J*_{4,5} 6.0 Hz, H-4), 4.59 (d, 1H, *J*_{3,4} 4.8 Hz, H-3), 4.26 (s, 1H, H-2), 4.10 (m, 1H, OCH₂CH=CH₂), 3.96 (d, 1H, *J*_{4,5} 6.0 Hz, H-5), 3.87 (m, 1H, OCH₂CH=CH₂). ¹³C NMR (D₂O) δ 177.28 (C=O), 136.53 (CH=CH₂), 115.43 (CH=CH₂), 107.43 (C-1), 83.32 (C-3), 77.87 (C-4), 76.24 (C-2), 68.79 (C-5), 62.53 (OCH₂). ESI-HRMS+ calculated for C₉H₁₂LiO₆ [M + Li]⁺ *m/z* 223.1306, found 223.1204.

3.2.2. 3'-butenyl D-glucofuranosidurono-6,3-lactone 8

Mixture of α and β isomers: colorless oil. α and β isomers were isolated. IR (NaCl) ν 3429 cm⁻¹, 2933 cm⁻¹, 1783 cm⁻¹. **8β**: ¹H NMR (600 MHz, CDCl₃) δ 5.77 (m, 1H, CH=CH₂), 5.08 (m, 2H, CH=CH₂), 5.05 (s, 1H, H-1), 4.96 (dd, 1H, *J*_{3,4} 3.6 Hz, *J*_{4,5} 6.6 Hz, H-4), 4.81 (d, 1H, *J*_{3,4} 3.6 Hz, H-3), 4.35 (d, 1H, *J*_{4,5} 6.6 Hz, H-5), 4.33 (s, 1H, H-2), 3.72 (m, 2H, OCH₂), 2.30–2.25 (m, 2H, CH₂). ¹³C NMR (CDCl₃) δ 174.69 (C=O), 134.77 (CH=CH₂), 117.49 (CH=CH₂), 109.05 (C-1), 83.18 (C-3), 77.31 (C-2), 77.04 (C-4), 69.17 (C-5), 68.08 (OCH₂), 33.47 (C-H₂CH=CH₂). **8α**: ¹³C NMR (CDCl₃) δ 174.69 (C=O), 102.48 (C-1), 84.72 (C-3), 76.82 (C-2), 76.02 (C-4), 70.07 (C-5), 69.12 (OCH₂), 33.78 (C-H₂CH=CH₂).

3.2.3. 5'-hexenyl D-glucofuranosidurono-6,3-lactone 9. Mixture of α and β isomers: colorless oil. Only the β anomer was isolated. **9 β** : IR (NaCl) ν 3418 cm^{-1} , 2937 cm^{-1} , 2864 cm^{-1} , 1785 cm^{-1} . ^1H NMR (600 MHz, CDCl_3) δ 5.74 (m, 1H, $\text{CH}=\text{CH}_2$), 5.02 (s, 1H, H-1), 4.94 (m, 3H, H-4, $\text{CH}=\text{CH}_2$), 4.81 (d, 1H, $J_{3,4}$ 4.2 Hz, H-3), 4.38 (d, 1H, $J_{4,5}$ 6.6 Hz, H-5), 4.29 (s, 1H, H-2), 3.66 (m, 1H, OCH_2), 3.33 (m, 1H, OCH_2), 2.04-1.98 (q, 2H, J 6.6 Hz, $\text{CH}_2\text{CH}=\text{CH}_2$), 1.54-1.48 (m, 2H, OCH_2CH_2), 1.38 (m, 2H, CH_2). ^{13}C NMR (CDCl_3) δ 175.13 (C = O), 138.56 ($\text{CH}=\text{CH}_2$), 114.55 ($\text{CH}=\text{CH}_2$), 109.03 (C-1), 83.25 (C-3), 77.28 (C-2), 77.07 (C-4), 69.18 (C-5), 68.53 (OCH_2), 33.32 ($\text{CH}_2\text{CH}=\text{CH}_2$), 31.90, 28.49, 24.94. ESI-HRMS+ calculated for $\text{C}_{12}\text{H}_{19}\text{LiO}_6$ [$\text{M} + \text{Li}$] $^+$ m/z 265.2199, found 265.2204.

3.2.4. 10'-undecenyl D-glucofuranosidurono-6,3-lactone 10

Mixture of α and β isomers: off-white solid. Only the β anomer was isolated. **10 β** : white solid (recrystallised in Et_2O). mp 77–79°C. IR (KBr) ν 3440 cm^{-1} , 2924 cm^{-1} , 2854 cm^{-1} , 1758 cm^{-1} . ^1H NMR (600 MHz, CDCl_3) δ 5.78 (m, 1H, $\text{CH}=\text{CH}_2$), 5.04 (s, 1H, H-1), 4.96 (m, 2H, H-4, $\text{CH}=\text{CH}_2$), 4.89 (d, 1H, $\text{CH}=\text{CH}_2$), 4.83 (d, 1H, $J_{3,4}$ 4.2 Hz, H-3), 4.36 (d, 1H, $J_{4,5}$ 7.2 Hz, H-5), 4.33 (s, 1H, H-2), 3.59 (m, 1H, OCH_2), 3.37 (m, 1H, OCH_2), 2.02-1.99 (q, 2H, J 6.6 Hz, $\text{CH}_2\text{CH}=\text{CH}_2$), 1.54-1.48 (m, 2H, OCH_2CH_2), 1.25 (m, 12H, CH_2). ^{13}C NMR (CDCl_3) δ 174.69 (C = O), 139.17 ($\text{CH}=\text{CH}_2$), 114.08 ($\text{CH}=\text{CH}_2$), 109.08 (C-1), 83.18 (C-3), 77.32 (C-2), 77.03 (C-4), 69.14 (C-5, OCH_2), 33.76 ($\text{CH}_2\text{CH}=\text{CH}_2$), 29.51, 29.45, 29.16, 28.89, 25.68.

3.2.5. 3'-Chloropropyl D-glucofuranosidurono-6,3-lactone 11

Mixture of α and β isomers: yellow oil. Only the β anomer was isolated. **11 β** : colorless oil. TLC (EtOAc/*n*-hexane 1/1 v/v): R_f 0.17. IR (NaCl) ν 3345 cm^{-1} , 2964 cm^{-1} , 2888 cm^{-1} , 1783 cm^{-1} , 1052 cm^{-1} . ^1H NMR (600 MHz, CDCl_3) δ 5.06 (s, 1H, H-1), 4.98 (dd, 1H, $J_{3,4}$ 3.6 Hz, $J_{4,5}$ 6.6 Hz, H-4), 4.84 (d, 1H, $J_{3,4}$ 3.6 Hz), 4.41 (d, 1H, $J_{4,5}$ 6.6 Hz, H-5), 4.34 (s, 1H, H-2), 3.85 (m, 2H, OCH_2), 3.47 (t, 2H, J 7.2 Hz, CH_2Cl), 1.94 (m, 2H, OCH_2CH_2). ^{13}C NMR (CDCl_3) δ 175.07 (C = O), 109.19 (C-1), 83.17 (C-3), 77.47 (C-2), 76.85 (C-4), 69.32 (C-5), 65.03 (OCH_2), 41.76 (CH_2Cl), 31.94 (OCH_2CH_2).

3.2.6. Benzyl D-glucofuranosidurono-6,3-lactone 12

Mixture of α and β isomers: yellow oil. IR (NaCl) ν 3395 cm^{-1} , 3034 cm^{-1} , 2879 cm^{-1} , 1793 cm^{-1} .

Only the β anomer was isolated. **12 β** : TLC (EtOAc/*n*-hexane 1/1 v/v): R_f 0.12. ^1H NMR (600 MHz, CDCl_3) δ 7.41–7.33 (m, 5H, H_{arom}), 5.07 (s, 1H, H-1), 4.79 (dd, 1H, $J_{3,4}$ 3.6 Hz, $J_{4,5}$ 6.6 Hz, H-4), 4.62 (m, 3H, CH_2Ph , H-3), 4.29 (s, 1H, H-2), 4.16 (d, 1H, $J_{4,5}$ 6.6 Hz, H-5). ^{13}C NMR (CDCl_3) δ 174.97 (C = O), 140.84 (C_{ispo}), 128.45 (C_{meta}), 128.30 (C_{para}), 127.56 (C_{ortho}), 108.51 (C-1), 83.14 (C-3), 77.61 (C-2), 76.92 (C-4), 70.41 (C-5), 66.81 (OCH_2). ESI-HRMS+ calculated for $\text{C}_{13}\text{H}_{14}\text{LiO}_6$ [$\text{M} + \text{Li}$] $^+$ m/z 273.1929, found 273.1932.

3.2.7. Cyclohexyl D-glucofuranosidurono-6,3-lactone 13

Mixture of α and β anomers: colorless oil. Only the β anomer was isolated. **13 β** : IR (NaCl) ν 3416 cm^{-1} , 2946 cm^{-1} , 2862 cm^{-1} , 1786 cm^{-1} . ^1H NMR (600 MHz, CDCl_3) δ 5.21 (s, 1H, H-1), 4.95 (dd, 1H, $J_{3,4}$ 4.8 Hz, $J_{4,5}$ 6.6 Hz, H-4), 4.82 (d, 1H, $J_{3,4}$ 4.8 Hz, H-3), 4.33-4.31 (m, 2H, H-5, H-2), 3.74-3.68 (m, 1H, OCH), 1.85 (m, 2H, CH_2), 1.69 (m, 2H, CH_2), 1.51 (d, 1H, J 6.0 Hz, CH_2), 1.28-1.12 (m, 5H, CH_2). ^{13}C NMR (CDCl_3) δ 174.39 (C = O), 106.79 (C-1), 83.23 (C-3), 77.84 (C-2), 76.81 (C-4), 70.35 (OCH), 68.99 (C-5), 32.71 (CH_2), 25.40 (CH_2), 24.09 (CH_2). ESI-HRMS+ calculated for $\text{C}_{12}\text{H}_{18}\text{LiO}_6$ [$\text{M} + \text{Li}$] $^+$ m/z 265.2169, found 265.2164.

3.2.8. 6'-hydroxy-*n*-hexyl D-glucofuranosidurono-6,3-lactone 14

Mixture of α and β isomers: yellow oil. Only the β anomer was isolated. **14 β** : IR (NaCl) ν 3345 cm^{-1} , 2932 cm^{-1} , 2859 cm^{-1} , 1782 cm^{-1} . ^1H NMR (600 MHz, CDCl_3) δ 5.06 (s, 1H, H-1), 4.98 (dd, 1H, $J_{3,4}$ 4.8 Hz, $J_{4,5}$ 7.2 Hz, H-4), 4.82 (d, 1H, $J_{3,4}$ 4.8 Hz, H-3), 4.35 (m, 2H, H-5, H-2), 3.71 (m, 2H, OCH_2), 3.61 (m, 2H, HOCH_2), 1.52 (OCH_2CH_2), 1.33 (CH_2). ^{13}C NMR (CDCl_3) δ 175.92 (C = O), 109.45 (C-1), 83.33 (C-3), 77.46 (C-2), 77.12 (C-4), 69.48 (C-5), 62.80 (OCH_2), 62.18 (OCH_2), 32.48 (OCH_2CH_2), 25.71 (CH_2).

3.2.9. Propargyl D-glucofuranosidurono-6,3-lactone 15

Mixture of α and β isomers: colorless oil. IR (NaCl) ν 3440 cm^{-1} , 2938 cm^{-1} , 2875 cm^{-1} , 2121 cm^{-1} , 1783 cm^{-1} . α and β isomers were isolated. **15 β** : ^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 5.09 (s, 1H, H-1), 4.77 (dd, 1H, $J_{3,4}$ 4.8 Hz, $J_{4,5}$ 6.6 Hz, H-4), 4.70 (d, 1H, $J_{3,4}$ 4.8 Hz, H-3), 4.41 (d, 1H, $J_{4,5}$ 6.6 Hz, H-5), 4.21 (s, 2H, OCH_2), 4.06 (s, 1H, H-2), 3.35 (s, $\text{C}\equiv\text{CH}$). ^{13}C NMR ($\text{DMSO}-d_6$) δ 175.27 (C = O), 106.55 (C-1), 84.42 ($\text{C}\equiv\text{CH}$), 83.01 (C-3), 78.82 (C-4), 77.49 (C-2), 75.15 ($\text{C}\equiv\text{CH}$), 69.02 (C-5), 55.83 (OCH_2). **15 α** : ^{13}C NMR ($\text{DMSO}-d_6$) δ 175.27 (C = O), 101.62 (C-1), 85.00

(C-3), 84.41 (C≡CH), 79.75 (C-2), 76.82 (C-4), 76.17 (C≡CH), 69.09 (C-5), 55.04 (OCH₂).

3.2.10. Poly(ethylene glycol) D-glucofuranosidurono-6,3-lactone 16

Mixture of α and β isomers: light yellow oil. Only the β anomer was isolated. **16 β** : IR (NaCl) ν 3382 cm⁻¹, 2877 cm⁻¹, 1789 cm⁻¹. ¹H NMR (600 MHz, D₂O) δ 5.07 (s, 1H, H-1), 4.93 (m, 2H, H-3, H-4), 4.59 (d, 1H, *J*_{5,4} 6.0 Hz, H-5), 4.28 (s, 1H, H-2), 3.72 (m, 1H, OCH₂), 3.58 (m, 1H, OCH₂), 3.45 (br, CH₂-PEG). ¹³C NMR (D₂O) δ 177.28 (C=O), 108.37 (C-1), 83.40 (C-3), 77.74 (C-4), 76.18 (C-2), 68.95 (C-5), 66.99 (OCH₂).

3.2.11. n-Octanethio-D-glucofuranosidurono-6,3-lactone 17

Isolated as the β anomer. Light yellow oil. ¹H NMR (600 MHz, CDCl₃) δ 5.17 (s, 1H, H-1), 4.44 (dd, 1H, H-5), 4.32 (d, 1H, H-4), 3.74 (t apparent, 1H, H-3), 3.42 (d, 1H, H-2), 2.62 (m, 2H, SCH₂), 1.54 (m, 2H, SCH₂CH₂), 1.26 (m, 10H, CH₂), 0.81 (t, 3H, CH₃). ¹³C NMR (CDCl₃) δ 173.8 (C=O), 90.5 (C-1), 86.16 (C-4), 70.68 (C-5), 54.9 (C-2), 48.1 (C-3), 31.7 (SCH₂), 29.8, 29.4, 29.3, 28.9, 28.8, 24.5 (CH₂), 13.9 (CH₃).

3.2.12. n-Tetradecanethio-D-glucofuranosidurono-6,3-lactone 18

Isolated as the β anomer. Light brown oil. ¹H NMR (600 MHz, CD₃OD) δ 4.99 (s, 1H, H-1), 4.86 (dd, 1H, *J*_{4,5} 6.5 Hz, H-4), 4.79 (d, 1H, *J*_{3,4} 4.1 Hz, H-3), 4.51 (d, 1H, *J*_{5,4} 6.5 Hz, H-5), 4.16 (s, 1H, H-2), 3.78 (dt, 1H, -OCH₂), 3.26 (dt, 1H, -OCH₂), 1.48 (m, 2H, -OCH₂C-H₂), 1.33-1.22 (m, 22H, (CH₂)₁₁), 0.88 (t, 3H, CH₃). RMN ¹³C (CD₃OD): δ (ppm) = 175.6 (C-6), 109.06 (C-1), 84.3 (C-3), 78.1 (C-2), 77.8 (C-4), 70.8 (C-5), 67.83 (-OCH₂), 31.64, 29.36, 29.32, 29.28, 29.23, 29.16, 29.03, 28.83, 25.70, 22.29, (-CH₂), 13.1 (CH₃).

3.2.13. PEG (PEG D-galactosid)uronate 24

Mixture of furanosid and pyranosid forms. PEG (PEG β -D-galactofuranosid)uronate **21 β** : yellow oil. TLC (EtOAc): *R*_f 0.12. IR (NaCl) ν 3410 cm⁻¹, 1750 cm⁻¹. ¹³C NMR (D₂O) δ 172.88 (C=O), 107.47 (C-1), 83.33 (C-4), 80.66 (C-2), 75.71 (C-3), 69.47 (PEG), 68.27 (C-5), 66.86 (OCH₂), 64.72 (CO₂CH₂), 60.39 (PEG).

4. Conclusion

In summary, we have illustrated that combination of microwave and a supported acid catalyst can be used to carry out the one-step functionalization of two

representative acidic carbohydrates in short times and in solventless conditions. This straightforward approach can prove beneficial as the solid catalyst was removed by filtration at the end of the process, thus minimizing wastes. The advantage of microwaves is evident as high yields were associated to high chemo- and anomeric-selectivities.

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Note

1. The loss factor (tan δ) is defined as the ratio between dielectric loss (ϵ'') and the dielectric constant (ϵ') and is used to quantify the ability of a material to convert microwaves into heat energy at a given temperature and frequency. For practical applications, at 2.45 GHz and 20°C, solvents could be categorized into “excellent” (with tan $\delta > 0.6$ –0.5), “medium” (0.5 > tan $\delta > 0.1$) or “poor” (tan $\delta < 0.1$) candidates for microwave applications.

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