## University of Miami Scholarly Repository

**Open Access Dissertations** 

**Electronic Theses and Dissertations** 

2015-04-30

## Dopaminergic Variants in Siblings at High Risk for Autism: Associations with Joint Attention and Behavior Problems

Devon N. Gangi University of Miami, devon.gangi@gmail.com

Follow this and additional works at: https://scholarlyrepository.miami.edu/oa dissertations

#### **Recommended** Citation

Gangi, Devon N., "Dopaminergic Variants in Siblings at High Risk for Autism: Associations with Joint Attention and Behavior Problems" (2015). *Open Access Dissertations*. 1391. https://scholarlyrepository.miami.edu/oa\_dissertations/1391

This Embargoed is brought to you for free and open access by the Electronic Theses and Dissertations at Scholarly Repository. It has been accepted for inclusion in Open Access Dissertations by an authorized administrator of Scholarly Repository. For more information, please contact repository.library@miami.edu.

#### UNIVERSITY OF MIAMI

# DOPAMINERGIC VARIANTS IN SIBLINGS AT HIGH RISK FOR AUTISM: ASSOCIATIONS WITH JOINT ATTENTION AND BEHAVIOR PROBLEMS

By

Devon Nicole Gangi

A DISSERTATION

Submitted to the Faculty of the University of Miami in partial fulfillment of the requirements for the degree of Doctor of Philosophy

Coral Gables, Florida

May 2015

©2015 Devon Nicole Gangi All Rights Reserved

#### UNIVERSITY OF MIAMI

#### A dissertation submitted in partial fulfillment of the requirements for the degree of Doctor of Philosophy

## DOPAMINERGIC VARIANTS IN SIBLINGS AT HIGH RISK FOR AUTISM: ASSOCIATIONS WITH JOINT ATTENTION AND BEHAVIOR PROBLEMS

Devon Nicole Gangi

Approved:

Daniel S. Messinger, Ph.D. Professor of Psychology, Pediatrics, Electrical and Computer Engineering, and Music Engineering Heather A. Henderson, Ph.D. Adjunct Associate Professor of Psychology

Jennifer C. Britton, Ph.D. Assistant Professor of Psychology Eden R. Martin, Ph.D. Professor of Human Genetics and Public Health Sciences

Michael L. Cuccaro, Ph.D. Associate Professor of Psychology and Human Genetics M. Brian Blake, Ph.D. Dean of the Graduate School Abstract of a dissertation at the University of Miami.

Dissertation supervised by Professor Daniel S. Messinger. No. of pages in text. (40)

Infant siblings at risk for Autism Spectrum Disorder (ASD; high-risk siblings) exhibit lower levels of joint attention and higher levels of behavior problems than lowrisk siblings (siblings with no family history of ASD), but also exhibit high levels of variability in these domains. The neurotransmitter dopamine is linked to brain areas associated with attention, reward, and motivation. Common genetic variants affecting dopamine neurotransmission, DRD4 and DRD2, have been associated with attention difficulties and behavior problems in typically developing children. We examined whether these variants explain variability in ASD-relevant behaviors in high-risk siblings. DRD4 and DRD2 genotypes for high-risk and low-risk siblings were coded according to dopaminergic functioning to create a gene score, with higher scores indicating more alleles associated with lower dopaminergic functioning. Initiating joint attention (IJA) was observed in the first year, and parents reported behavior problems at 3 years using the Child Behavior Checklist. Dopamine gene scores indicative of lower dopaminergic functioning were associated with less optimal behavior in the first year (lower levels of IJA) and at 3 years (higher levels of internalizing problems) for high-risk siblings, while the opposite pattern typically emerged in low-risk siblings. Lower dopaminergic function was associated with poorer referential communication and increased behavior problems only in the presence of familial risk for ASD. Findings suggest differential susceptibility—children's ASD-relevant behaviors were differentially affected by

dopaminergic functioning depending on their familial risk for ASD. Understanding genes linked to ASD-relevant behavioral difficulties in high-risk siblings will aid in the very early identification of children at greatest risk for such difficulties, opening the way for targeted prevention and intervention protocols.

## **TABLE OF CONTENTS**

LIST	OF FIGURESi	v
LIST	OF TABLES	7
Chapt	er	
1	INTRODUCTION       1         Genetics and ASD       1         Heterogeneity within ASD       2         Dopaminergic Variants and Behavior       3         ASD-Relevant Behavior       4         Current Study       6	l 2 3 4
2	METHOD	3
3	RESULTS       1         Dopaminergic Genotypes       1         Dependent Variables       1         Initiating Joint Attention       1         CBCL Externalizing Behavior Problems       1         CBCL Internalizing Behavior Problems       1         Mediation Model       1	2 3 3 4 5
4	DISCUSSION1	7
REFE	RENCES	23
FIGU	RES	31
TABL	JES	34

## LIST OF FIGURES

## Page

FIGURE 1	
FIGURE 2	
FIGURE 3	

## LIST OF TABLES

TABLE 1a	
TABLE 1b	
TABLE 2	
TABLE 3	
TABLE 4	
TABLE 5	
TABLE 6	40

#### **CHAPTER ONE**

#### **INTRODUCTION**

Autism Spectrum Disorder (ASD) is a complex neurodevelopmental condition characterized by a broad range of social and communication impairments and stereotyped patterns of behavior (American Psychiatric Association, 2013) with prevalence estimates of over 1 in 75 children (CDC, 2014). The younger siblings of children with ASD (highrisk siblings) have high rates of ASD diagnosis, with recurrence rates of 4.5-18.7%, and exhibit substantial heterogeneity in behaviors associated with ASD, with an additional one fifth of high-risk siblings exhibiting sub-clinical difficulties (Grønborg, Schendel, & Parner, 2013; Messinger et al., 2013; Ozonoff et al., 2011; Risch et al., 2014), including initiating joint attention and behavior problems. Common genetic variants such as dopaminergic genes *DRD4* and *DRD2* may aid in understanding the variability of phenotypic presentation in high-risk siblings. The current study examined these dopaminergic variants in high-risk siblings and low-risk siblings (siblings with no family history of ASD) to better understand heterogeneity in behavioral phenotypes particularly relevant to ASD, initiating joint attention and behavior problems.

#### Genetics and ASD

Although recent estimates suggest substantial heritability for ASD (Colvert et al., 2015; Hallmayer et al., 2011), specific genes responsible for this heritability are not clear (Geschwind, 2011). Both rare and common variants contribute to understanding genetic susceptibility in ASD. Several rare variants (mutations with a minor allele frequency of less than 1%) associated with ASD have been identified (Betancur, 2011); however, no specific gene accounts for a majority of ASD cases (Abrahams & Geschwind, 2008;

Geschwind, 2011; Muhle, Trentacoste, & Rapin, 2004). Even among siblings both diagnosed with ASD, most do not share the same ASD risk genes, underscoring the genetic heterogeneity of ASD (Yuen et al., 2015) and highlighting the potential difficulty of identifying replicable ASD susceptibility genes. Common variants (polymorphisms that occur in greater than 1-2% of the population) may comprise a substantial portion of the risk heritability of ASD (Gaugler et al., 2014; Klei et al., 2012). However, identified common variants that in combination or alone influence ASD susceptibility have not been well-replicated (Anney et al., 2010; Devlin, Melhem, & Roeder, 2011; Muhle et al., 2004). As genetic underpinnings of ASD are highly heterogeneous and a number of genes likely interact to influence susceptibility (Talkowski, Minikel, & Gusella, 2014), an approach focusing on the genetic basis of behaviors relevant to ASD may be productive in identifying genotypes associated with specific ASD-related traits (Muhle et al., 2004). *Heterogeneity within ASD* 

In addition to ASD's genetic variability, ASD is phenotypically heterogeneous, encompassing a broad spectrum of impairment. Those diagnosed can exhibit varied combinations of traits and symptoms (Rapin, 1991; Rutter & Schopler, 1987), resulting in a range of later outcomes (Howlin, Goode, Hutton, & Rutter, 2004). ASD-relevant behaviors, which are characteristic of the disorder and its symptomatology, show substantial variability in both children with ASD and in their younger siblings. Even without an ASD diagnosis, high-risk siblings exhibit elevated ASD symptoms, lower levels of developmental functioning, and behavioral difficulties (Gangi, Ibañez, & Messinger, 2014; Georgiades et al., 2013; Messinger et al., 2013). In low-risk children, common genetic variants have been linked to behavioral phenotypes (e.g., Bakermans-Kranenburg & van IJzendoorn, 2011; Lackner, Sabbagh, Hallinan, Liu, & Holden, 2012; Posner, Rothbart, & Sheese, 2007). Here, we aimed to examine common genetic variants implicated in behavior in the context of risk for ASD, to determine whether these variants may play a role in the heterogeneity seen in behaviors that are relevant to and have implications for ASD. Though individual common genetic variants are unlikely to distinguish children with ASD from case controls, these variants may be related to phenotypic variability in ASD-relevant behaviors (Geschwind, 2011) among high-risk siblings. We examined the role of two common genetic variants (*DRD4* and *DRD2*) in a sample including high-risk siblings to understand phenotypic, behavioral heterogeneity in the context of familial ASD risk. *Dopaminergic Variants and Behavior* 

While relationships between dopaminergic variants and behavior have been studied in typically-developing children, there has been little examination in children at risk for ASD. Dopamine is a catecholamine that functions as a neurotransmitter in the brain, and it plays a role in several key domains including attention, reward-motivated behavior, and motor control. In the brain, dopamine is produced in areas including the substantia nigra and ventral tegmental area and then is transmitted through several main pathways, some of which are associated with the control of motivation-linked systems relevant to the current study. The mesolimbic and mesocortical pathways begin in the ventral tegmental area and connect to the nucleus accumbens and cerebral cortex, respectively, and they are associated with response to reward and motivation. The nigrostriatal pathway begins in the substantia nigra and connects to the striatum, and it is associated with motor control.

Several common polymorphisms affect dopamine neurotransmission. The DRD4 gene encodes for dopamine receptor D4, which is expressed in areas including the frontal cortex, hippocampus, amygdala, and hypothalamus (Beaulieu & Gainetdinov, 2011). Variants in a 48-base pair variable number tandem repeat of *DRD4* can influence gene expression, and a "long" version (the 7-repeat allele) has been associated with suppressed receptor expression (Schoots & Van Tol, 2003). The 7-repeat allele has been associated with varied attentional and behavioral difficulties in typically developing children and infants (Auerbach, Benjamin, Faroy, Geller, & Ebstein, 2001; Auerbach, Faroy, Ebstein, Kahana, & Levine, 2001; Gizer, Ficks, & Waldman, 2009; Schmidt, Fox, Perez-Edgar, Hu, & Hamer, 2001). Among children with ASD, those with the 7-repeat allele tend to have greater behavior problems than those without the 7-repeat allele (Gadow, DeVincent, Olvet, Pisarevskaya, & Hatchwell, 2010). The DRD2 gene encodes for the dopamine receptor D2, which is expressed in areas including the striatum and nucleus accumbens (Beaulieu & Gainetdinov, 2011), and is associated with the Taq1A polymorphism on ANKK1. The A allele of the polymorphism (hereafter DRD2) is linked to a reduction in D2 receptor expression (Thompson et al., 1997) and is associated with risk for ASD and social interaction and communication difficulties (Hettinger et al., 2012; Salem et al., 2013).

#### ASD-Relevant Behavior

We focused on two ASD-relevant behaviors, initiation of joint attention and behavior problems. Early deficits in initiating joint attention (IJA), a form of referential communication involving the use of gaze and gesture to coordinate attention between social partners and objects, are a core feature of ASD (Dawson et al., 2004; Mundy, Sigman, Ungerer, & Sherman, 1986). Among high-risk siblings, early IJA is predictive of later ASD symptomatology (Ibañez, Grantz, & Messinger, 2012). While some evidence suggests high-risk siblings tend to display fewer IJA behaviors than low-risk siblings (Cassel et al., 2007; Goldberg et al., 2005; Ibañez et al., 2012; Rozga et al., 2011), other investigations do not report differences (Toth, Dawson, Meltzoff, Greenson, & Fein, 2007; Yirmiya et al., 2006). These mixed findings highlight the necessity for empirical work to explain phenotypic variability among high-risk siblings. High levels of both internalizing and externalizing behavior problems are also reported in children with ASD (Mahan & Matson, 2011; Maskey, Warnell, Parr, Couteur, & McConachie, 2013) and their high-risk siblings (Fisman et al., 1996; Rodrigue, Geffken, & Morgan, 1993; Verté, Roeyers, & Buysse, 2003). Behavior problems are associated with both the severity of ASD symptomatology (Pearson et al., 2006) and parent stress and depression (Davis & Carter, 2008; Ekas & Whitman, 2010).

IJA and behavior problems are particularly relevant to high-risk siblings, as they may be predictive of later symptomatology and family emotional outcomes. Given the role of the dopaminergic system in reward sensitivity and motivation, it may influence whether an infant finds social interaction through joint attention rewarding and is motivated to perform IJA behaviors and whether a child is likely to exhibit externalizing or internalizing behavior problems. In addition, the dopaminergic system's role in motor control and attention may play a role in infants' ability to shift attentional focus and execute such behaviors. Despite the relationship between dopaminergic variants and related functioning in typical development, similar associations have not been examined within children at risk for ASD. Investigating relations between behavioral phenotypes and dopaminergic genotypes in the context of familial risk for ASD may aid in understanding the manifestation of early ASD-relevant behaviors, enabling early identification of behavioral targets for early intervention.

#### Current Study

The current study examined dopaminergic genotypes *DRD4* and *DRD2* in relation to ASD-relevant behavioral phenotypes (i.e., joint attention and behavior problems) in the context of familial autism risk.

Aim 1. Characterize dopaminergic genotype distributions in high-risk and low-risk siblings. We examined distributions of genotype frequencies (*DRD4*, *DRD2*, and a dopamine gene score comprised of both genes) in high- and low-risk siblings. We did not expect genotype frequencies to differ between groups (i.e., risk alleles would not be overrepresented in high-risk siblings).

Aim 2. Examine the relationship between dopaminergic variants and ASD-relevant behavioral phenotypes in high-risk and low-risk siblings. Regression models tested the effect of dopaminergic genotype, as well as its interaction with risk group status, on ASD-relevant behaviors (i.e., IJA in the first year and behavior problems at three years). We expected lower dopaminergic functioning to be associated with lower levels of IJA and higher levels of problem behaviors.

Aim 3. Determine whether ASD-relevant behavior in the first year (IJA) mediates a relationship between dopaminergic functioning and later ASD symptomatology. A mediation model tested whether dopaminergic functioning (indexed by dopaminergic

genotype composite) is associated with ASD symptomatology at 30 months through its relationship to IJA in the first year.

#### **CHAPTER TWO**

#### METHOD

#### *Participants*

Participants were the infant siblings of children diagnosed with Autism Spectrum Disorder (ASD; high-risk siblings, *n*=55) or the infant siblings of typically developing children with no history of ASD (low-risk siblings, *n*=38). High-risk siblings have at least one older sibling with a diagnosis of ASD, confirmed upon study enrollment by administration of the Autism Diagnostic Observation Schedule (ADOS; Lord et al., 2000) and clinical diagnosis by a licensed clinical psychologist. Low-risk siblings have older siblings with no evidence of ASD, confirmed by a score lower than 9 on the Social Communication Questionnaire (Berument, Rutter, Lord, Pickles, & Bailey, 1999), a conservative cutoff score, and no family history of ASD.

#### Measures

*Early Social Communication Scales (ESCS).* Joint attention was assessed within the ESCS (Mundy et al., 2003) at 8, 10, and 12 months. The ESCS is a semi-structured assessment of infants' nonverbal communication abilities, during which an examiner (seated across from the infant) presents and activates a series of toys, creating opportunities for the infant to initiate joint attention. After presenting and activating a toy, the examiner remains attentive and responds to the infants' joint attention bids briefly. The current study focused on initiating joint attention (IJA) bids occurring during the ESCS (e.g., when infant gazed between the examiner and activated toy or showed an object to the examiner). Videotaped assessments were reliably coded by trained coders. Rates per minute of joint attention were calculated for each assessment age; a mean was calculated from the standardized values of each assessment age to provide a measure of IJA in the first year for analyses.

*Child Behavior Checklist (CBCL)*. Parent-reported behavior problems were assessed within the CBCL (Achenbach, Edelbrock, & Howell, 1987; Achenbach & Rescorla, 2000) at 36 months. The CBCL is a well-validated parent-report measure of children's behavior problems and yields subscales of Internalizing and Externalizing problems, normed by age and sex.

ASD Symptomatology. ASD symptomatology was assessed within the Autism Diagnostic Observation Schedule (ADOS; Lord et al., 2000), a play-based observational measure during with an examiner administers behavioral presses designed to elicit ASD-relevant behaviors. This assessment was administered at 30 months, and children were administered either Module 1 (n=39) or Module 2 (n=41) based on language level. Risk groups did not differ in which ADOS Module was administered,  $\chi^2(1)$ =0.23, p=.63.

To capture a continuous measure of ASD symptomatology, overall calibrated severity scores (overall CSS) were calculated for each child based on Gotham, Pickles, and Lord's (2009) criteria. In addition, Hus, Gotham, and Lord's (2014) criteria were used to calculate calibrated severity scores for each child in two domains: social affect domain (SA-CSS) and restricted and repetitive behaviors domain (RRB-CSS). Scores for each domain range from 1 to 10 and account for the child's age and language level. High-risk siblings (M=3.07, SD=.2.17) had higher overall CCS than low-risk siblings (M=1.51, SD=1.27), t(78)=-3.76, p<.001; high-risk siblings (M=3.36, SD=2.27) had higher SA-CCS than low-risk siblings (M=1.91, SD=1.50), t(78)=-3.24, p=.002; and high-risk siblings (M=4.33, SD=2.59) had higher RRB-CCS than low-risk siblings

(M=2.20, SD=1.94), t(78)=-4.06, p<.001. Ten high-risk siblings had overall CSS at or above the cutoff for ASD (a score of 4 or above), and 7 had scores at or above the cutoff for autism (a score of 6 or above). One low-risk sibling had a score at or above the ASD or autism cutoffs.

Clinical diagnosis was determined at 36 months (*n*=85). The ADOS administered at 30 months, the Autism Diagnostic Interview-Revised (ADI-R; Lord, Rutter, & Couteur, 1994) administered at 36 months, and the Mullen Scales of Early learning administered at 36 months were used to inform the DSM-IV-based best-estimate diagnosis from a licensed psychologist. Twelve high-risk siblings received a diagnosis of ASD, and no low-risk siblings were diagnosed with ASD.

*Dopamine Genotypes*. Genetic data was collected from saliva samples from participants using Oragene DNA collection kits. Genetic samples were sent for extraction and analysis to the John P. Hussman Institute for Human Genomics (HIHG) at the University of Miami Miller School of Medicine. Genotyping was conducted for *DRD4* and *DRD2*. Genotypes for *DRD4* (rs1805186) were grouped according to the presence or absence of the 7-repeat allele ("0" = no 7-repeat, "1" = at least one 7-repeat). For *DRD2* (rs1800497), genotypes were grouped according the presence of the A allele ("0" = no A allele, "1" = at least one A allele).

A dopamine gene score was also created by coding *DRD4* and *DRD2* to reflect dopaminergic functioning. Higher scores indicate more "risk" alleles (indexing lower dopaminergic functioning) and lower scores indicated fewer "risk" alleles (indexing greater dopaminergic functioning). Gene scores serve as an index of cumulative dopaminergic functioning for analyses of outcomes (for similar approaches, see: Nikolova, Ferrell, Manuck, & Hariri, 2011; Pearson-Fuhrhop et al., 2014; Stice, Yokum, Burger, Epstein, & Smolen, 2012), with participants coded as having 0, 1, or 2 risk genotype sets.

#### Analytic Approach

For Aim 1, initial analyses examined distributions of genotype frequencies, testing whether allelic frequencies were consistent with Hardy-Weinberg equilibrium. Fisher's exact tests determined whether genotype frequencies differed between high- and low-risk siblings, between high-risk siblings diagnosed with and without an ASD, and between ethnicities. Relations between dependent variables were also examined. For Aim 2, I next determined whether individual genotypes interacted with participants' risk status to predict behavioral phenotypes. For each dependent variable (IJA, Internalizing, and Externalizing), a 2 (genotype) by 2 (risk group) ANOVA for each of the dopaminergic variants tested for main effects of genotype and risk status, as well as their interaction. This was followed by a regression in which dopamine score, status, and dopamine score\*status interaction were entered as predictors to determine the cumulative effect of both dopaminergic genes. Interaction effects were followed up with individual models in which dependent variables were regressed on gene score. For Aim 3, a mediation model was tested to determine whether IJA mediates a relationship between dopamine score and later ASD symptomatology in high-risk siblings.

#### **CHAPTER THREE**

#### RESULTS

#### Dopaminergic Genotypes

Allelic distributions for DRD4,  $\chi^2(1)=0.02$ , p=.85, and DRD2,  $\chi^2(1)=0.00$ , p=.82, were consistent with Hardy-Weinberg equilibrium (Rodriguez, Gaunt, & Day, 2009). Allele frequencies for DRD4, p=.82, and DRD2, p=.25, did not differ between high-risk and low-risk siblings (see Table 1a; all repeat alleles for DRD4 are presented in Table 1b). Within high-risk siblings, allele frequencies for DRD4, p=1.00, and DRD2, p=.47, did not differ between children diagnosed with and without ASD at 36 months (see Table 2). Allele frequencies for DRD4, p=.15, and DRD2, p=.14, did not differ by ethnicity (Non-Hispanic White/Caucasian, Hispanic/Latino, and Other; see Table 3).

For analyses, genotypes for *DRD4* and *DRD2* were grouped according to the presence or absence of any alleles indicating lower dopaminergic functioning (7-repeat or A allele, respectively). Genotype frequencies for *DRD4*, p=.48, and *DRD2*, p=.37, did not differ between high-risk and low-risk siblings (see Table 4). Among high-risk siblings, genotype frequencies for *DRD4*, p=1.00, and *DRD2*, p=.32, did not differ between children diagnosed with and without ASD at 36 months (see Table 5). Genotype frequencies for *DRD4*, p=.14, did not differ by ethnicity (see Table 6). Dopamine composite scores also did not differ between high-risk and low-risk siblings, p=.69, between high-risk siblings diagnosed with and without ASD at 36 months, p=.51, or by ethnicity, p=.23 (see Tables 4-6).

#### Dependent Variables

In high-risk siblings, Internalizing Problems was correlated with Externalizing Problems, r=.62, p<.001. IJA was not correlated with Internalizing Problems, r=.16, p=.36, but was correlated with Externalizing Problems, r=.39, p=.02. In low-risk siblings, Internalizing Problems was correlated with Externalizing Problems, r=.49, p=.007. IJA was not correlated with Internalizing Problems, r=.32, p=.09, but was correlated with Externalizing Problems, r=.45, p=.02.

#### Initiating Joint Attention

*DRD4.* There was no main effect of genotype, F(1, 86)=0.05, *p*=.82, partial  $\eta^2$ =.001, power=.06, on IJA. There was a main effect of group status, F(1, 86)=12.08, *p*=.001, partial  $\eta^2$ =.12, power=.93, with high-risk siblings exhibiting lower levels of IJA than low-risk siblings. This main effect was modified by a genotype\*status interaction effect, F(1, 86)=8.80, *p*=.004, partial  $\eta^2$ =.09, power=.84. Among children without the 7-repeat allele, levels of IJA did not differ between high-risk (*M*=0.01, *SD*=0.84) and low-risk (*M*=0.11, *SD*=0.73) siblings, *t*(62)=0.48, *p*=.64. Among children with the 7-repeat allele, however, high-risk siblings (*M*=-0.52, *SD*=0.75) exhibited lower levels of IJA than low-risk siblings (*M*=0.73, *SD*=1.01), *t*(24)=3.62, *p*=.001.

*DRD2.* There was no main effect of genotype, F(1, 89)=0.01, p=.91, partial  $\eta^2=.00$ , power=.05, on IJA. There was a main effect of group status, F(1, 89)=6.58, p=.01, partial  $\eta^2=.07$ , power=.72, with high-risk siblings (*M*=-0.13, *SD*=0.84) exhibiting lower levels of IJA than low-risk siblings (*M*=0.29, *SD*=0.86). There was no genotype\*status interaction effect, F(1, 89)=1.56, p=.22, partial  $\eta^2=.02$ , power=.24.

*Dopamine Score.* A regression model assessed effects of the dopamine score, risk group status, and their interaction on IJA, adjusted  $R^2$ =0.13, F(3, 86)=5.31, p=.002, power=.87. There was no main effect of status, b=0.03, t=0.13, p=.90. There was a main effect of dopamine score, b=0.50, t=2.34, p=.02, such that children with higher dopamine scores tended to have higher IJA levels. There was also a dopamine score\*status interaction effect, b=-0.81, t=-3.09, p=.003. Regression analyses by risk group indicated that in high-risk siblings, IJA levels decreased as dopamine scores increased, b=-0.31, t=-2.03, p=.047, while in low-risk siblings, IJA levels increased as dopamine scores explained a significant proportion of variance in IJA in high-risk siblings, adjusted  $R^2$ =0.06, F(1, 52)=4.13, p=.047, power=.45, and in low-risk siblings, adjusted  $R^2$ =0.12, F(1, 34)=5.53, p=.03, power=.57.

#### CBCL Externalizing Behavior Problems

*DRD4.* There was no main effect of genotype, F(1, 57)=0.04, p=.84, partial  $\eta^2=.001$ , power=.05, on Externalizing Problems. There was a main effect of group status, F(1, 57)=5.14, p=.03, partial  $\eta^2=.08$ , power=.61, with high-risk siblings exhibiting higher levels of Externalizing Problems than low-risk siblings. This main effect was modified by a genotype\*status interaction effect, F(1, 57)=4.14, p=.047, partial  $\eta^2=.07$ , power=.52. Among children without the 7-repeat allele, levels of Externalizing Problems did not differ between high-risk (M=42.92, SD=9.13) and low-risk (M=42.28, SD=9.13) siblings, t(42)=-0.23, p=.82. Among children with the 7-repeat allele, however, low-risk siblings (M=36.11, SD=9.35) exhibited lower levels of Externalizing Problems than high-risk siblings (M=48.00, SD=12.38), t(15)=-2.25, p=.04.

*DRD2*. There was no main effect of genotype, F(1, 60)=0.87, p=.35, partial  $\eta^2=.01$ , power=.15, or group status, F(1, 60)=2.77, p=.10, partial  $\eta^2=.04$ , power=.37, on levels of Externalizing Problems, and there was no genotype\*status interaction effect, F(1, 60)=0.27, p=.61, partial  $\eta^2=.004$ , power=.08.

*Dopamine Score.* A regression model assessed effects of the dopamine score, risk group status, and their interaction on Externalizing Problems, adjusted  $R^2$ =0.04, F(3, 57)=1.89, p=.14, power=.65. There was no main effect of status, b=0.03, t=0.01, p=.99, or dopamine score, b=-2.53, t=-0.85, p=.40. There was no dopamine score\*status interaction effect, b=6.36, t=1.65, p=.10 (see Figure 2).

#### CBCL Internalizing Behavior Problems

*DRD4.* There was no main effect of genotype, F(1, 57)=0.01, p=.94, partial  $\eta^2=.00$ , power=.05, on Internalizing Problems. There was a main effect of group status, F(1, 57)=4.30, p=.04, partial  $\eta^2=.07$ , power=.53, with high-risk siblings exhibiting higher levels of Internalizing Problems than low-risk siblings. There was no genotype\*status interaction effect, F(1, 57)=3.39, p=.07, partial  $\eta^2=.06$ , power=.44.

*DRD2.* There was a main effect of genotype, F(1, 60)=8.03, p=.01, partial  $\eta^2=.12$ , power=.80, on levels of Internalizing Problems, with children with an A allele (*M*=46.27, *SD*=9.37) exhibiting higher levels of Internalizing Problems than children without an A allele (*M*=38.69, *SD*=9.16). There was no main effect of group status, F(1, 60)=1.99, p=.16, partial  $\eta^2=.03$ , power=.28, and there was no genotype\*status interaction effect, F(1, 60)=3.29, p=.08, partial  $\eta^2=.05$ , power=.43.

*Dopamine Score.* A regression model assessed effects of the dopamine score, risk group status, and their interaction on Internalizing Problems, adjusted  $R^2$ =0.13, *F*(3,

86)=5.31, p=.002, power=.69. There was no main effect of status, b=-2.84, t=-0.90, p=.38, or dopamine score, b=-2.89, t=-1.04, p=.30. There was a dopamine score\*status interaction effect, b=10.42, t=2.91, p=.005. Regression analyses by risk group indicated a significant effect of dopamine score for high-risk siblings, b=7.54, t=3.27, p=.003, but not for low-risk siblings, b=-2.32, t=-1.07, p=.30 (see Figure 3). Dopamine scores did not explain a significant proportion of variance in IJA in high-risk siblings, adjusted  $R^2$ =0.04, F(1, 32)=2.45, p=.13.

#### Mediation Model

In high-risk siblings who had completed an ADOS assessment at 30 months (n=44), dopamine score did not have a direct effect on overall CSS, b=-0.02, SE=0.44, p=.96. Dopamine score did not predict IJA, b=-0.19, SE=0.18, p=.29, and IJA did predict overall CSS, b=-0.81, SE=0.38, p=.04. The indirect effect was tested using a bootstrap estimation approach with 1000 samples (Preacher & Hayes, 2008); the indirect effect was not significant, b=0.15, SE=0.16, 95% CI=-0.12, 0.53.

Dopamine score did not have a direct effect on SA-CSS, b=0.31, SE=0.47, p=.52. Dopamine score did not predict IJA, b=-0.19, SE=0.18, p=.29, and IJA did predict SA-CSS, b=-0.92, SE=0.39, p=.02. The indirect effect, tested using a bootstrap estimation approach with 1000 samples, was not significant, b=0.18, SE=0.18, 95% CI=-0.15, 0.56.

Dopamine score did not have a direct effect on RRB-CSS, b=0.26, SE=0.56, p=.65. Dopamine score did not predict IJA, b=-0.19, SE=0.18, p=.29, and IJA did not predict RRB-CSS, b=-0.42, SE=0.48, p=.39. The indirect effect, tested using a bootstrap estimation approach with 1000 samples, was not significant, b=0.08, SE=0.15, 95% CI=-0.09, 0.56.

#### **CHAPTER FOUR**

#### DISCUSSION

Children at elevated risk for ASD exhibit heterogeneity in symptomatology, other behaviors relevant to ASD, and outcomes. Among high-risk siblings, early behavior often predicts diagnosis, but these patterns of prediction are not clear. We aimed to refine our understanding of heterogeneity in early behavior relevant to ASD by examining the role of common genetic variants. We examined the association between common variants related to dopaminergic functioning and initiating joint attention (IJA) and behavior problems. High-risk siblings with *DRD4* and *DRD2* genotypes linked to lower dopaminergic functioning exhibited lower levels of IJA and higher levels of internalizing behavior problems than high-risk siblings with variants linked to greater dopaminergic functioning. To our knowledge, this is the first investigation of these genetic variants in relation to attention and behavior problems in siblings at risk for ASD.

For high-risk siblings, lower dopaminergic functioning (indexed by higher dopamine scores) was associated with less optimal behavior both in the first year, with lower levels of IJA, and at three years, with elevated levels of internalizing behavior problems. IJA and behavior problems are both important for the development of highrisk siblings. Referential communication such as IJA is central to later language and social functioning in children at risk for ASD (Gangi et al., 2014; Ibañez et al., 2012; Malesa et al., 2012), and behavior problems have been associated with symptomatology in children with ASD and augment parent stress, with likely effects on the family system (Davis & Carter, 2008; Ekas & Whitman, 2010). Early referential communication difficulties and later behavior problems likely impact social functioning in children at risk

17

for ASD, and these behaviors appear to be influenced by dopaminergic genotypes. This link may allow for early identification of high-risk siblings at greatest risk for behavioral difficulties in these areas.

Although higher dopamine scores were associated with less optimal behavior among high-risk siblings, the opposite pattern emerged in low-risk siblings. Low-risk siblings with higher dopamine scores exhibited *higher* levels of IJA. This pattern suggests differential susceptibility, the hypothesis that children vary in their susceptibility to both adverse and beneficial effects of their environments (Belsky, 2005; Belsky, Bakermans-Kranenburg, & van IJzendoorn, 2007; Belsky & Pluess, 2009). Common genetic variants have been identified as potential susceptibility factors that modify individuals' susceptibility to influences affecting outcomes. The variants in the current study, *DRD4* and *DRD2*, act as susceptibility genes in multiple contexts (e.g., Bakermans-Kranenburg & van IJzendoorn, 2006, 2011; Sheese, Voelker, Rothbart, & Posner, 2007; Van IJzendoorn & Bakermans-Kranenburg, 2006), but have not been examined in the context of familial risk for ASD.

Although the differential susceptibility hypothesis is often conceptualized as susceptibility to rearing, it may also encompass sensitivity to a broader range of influences (Belsky & Pluess, 2009). For example, stronger associations between difficult child temperament and externalizing problems are found in children who have older siblings (Mesman et al., 2009). Endogenous factors have also been conceptualized as internal environments that affect the relationship between genes and outcomes (Schmidt, Fox, Perez-Edgar, & Hamer, 2009). That is, factors *within* an individual may play a role in moderating the association between genotype and developmental outcomes. Familial risk for ASD confers increased risk for ASD and related sub-clinical deficits to younger siblings of children diagnosed with ASD. Within a differential susceptibility framework, we conceptualize familial ASD risk as a functional context. Familial ASD risk likely encompasses combinations of genetic and environmental factors to which children may be more or less susceptible. Here, high-risk siblings with alleles linked to lower dopaminergic functioning exhibited lower levels of IJA and higher levels of behavior problems. In siblings with no alleles linked to lower dopaminergic functioning exhibited similar levels of IJA. Additional research will be necessary to determine specific genetic and environmental factors responsible for differential susceptibility among high-risk siblings (Hallmayer et al., 2011; Newschaffer et al., 2012; Yuen et al., 2015).

Within high-risk siblings, IJA did not mediate a relationship between dopamine score and later ASD symptomatology. However, this model was also unable to detect the initial association between dopamine score and IJA, likely due to the reduced number of high-risk siblings able to be included in the analysis (44 high-risk siblings who had completed an ADOS at 30 months). Sample size also limited analysis of high-risk siblings with ASD. Twelve high-risk siblings in the study sample were later diagnosed with ASD, a number insufficient for separate analyses. Findings from the current study should also be interpreted with caution until replicated with larger sample sizes. Particularly for analyses of behavior problems, which included fewer participants than IJA analyses, models may have been underpowered to detect interaction effects (power estimates for ANOVAs ranged from .08-.37 and for regressions ranged from .49-.91 for interaction effects). If employing corrections for multiple significance tests, with division by three for the number of dependent variables tested, the adjusted significance level would be p=.02. All but one significant interaction effect (*DRD4*\*status interaction for Externalizing Problems, p=.047) would survive this correction. Future research aimed at replicating our findings with larger sample sizes would strengthen our findings of a relationship between dopaminergic genotypes and ASD-relevant behaviors and could profitably investigate this relationship among high-risk children with ASD outcomes.

In addition to the *DRD4* and *DRD2* variants examined in the current study, other dopaminergic variants might be examined in future investigations of behavioral characteristics of high-risk siblings and children with autism. For example, a VNTR in the *DAT1* gene is associated with expression of the dopamine transporter (Fuke et al., 2001). Together, these variants might provide a more comprehensive index of dopaminergic functioning.

Genotypes outside the dopaminergic system may also impact the outcome of dopaminergic functioning and might further our understanding of dopamine's role in behavioral outcomes. For example, catechol-O-methyl transferase (encoded by the *COMT* gene) is an enzyme that degrades catecholamines including dopamine, and a polymorphism in the *COMT* gene is associated with dopaminergic function (Chen et al., 2004). Brain-derived neurotrophic factor (BDNF; coded for by the *BDNF* gene) may influence dopamine activity as well (Goggi, Pullar, Carney, & Bradford, 2003; Narita, Aoki, Takagi, Yajima, & Suzuki, 2003; Savitz, Solms, & Ramesar, 2006). Serotonergic function may also interact with dopaminergic function to influence behavioral outcomes, particularly internalizing and externalizing behavior problems. Levels of dopaminergic functioning might influence sensitivity to reward, leading to either high or low motivation toward rewards. Dopaminergic functioning might then interact with levels of serotonergic functioning influencing effortful control, which could aid in regulation of approach tendencies related to reward sensitivity (Carver, Johnson, & Joormann, 2009). Thus, a combination of high dopaminergic functioning (high reward sensitivity/approach) and low serotonergic functioning (low effortful control) might result in high levels of externalizing behavior.

In the current study, we found that dopaminergic risk alleles were associated with lower levels of IJA and higher levels of behavior problems in high-risk siblings. Given the systems in which dopamine plays a role, dopaminergic functioning could potentially affect children's social motivation, reward sensitivity, attention coordination, and even motor control. High-risk siblings with lower dopaminergic functioning appear to exhibit less optimal behavior, both in early social interaction and in later levels of behavior problems.

As the search for replicable genes associated with ASD risk is ongoing, an approach investigating genes that may be relevant to specific behaviors important for the development of children at risk for ASD may be a productive avenue of research. Genes that may not be associated with ASD itself may still be linked to particular behaviors. In addition to aiding in identifying high-risk siblings at greatest risk for difficulties, findings may also aid in identifying resilient children. High-risk siblings with fewer genes associated with lower dopaminergic functioning were exhibiting fewer difficulties in ASD-relevant behaviors than high-risk siblings carrying more genotypes associated with lower dopaminergic functioning. Referential communication and behavior problems are associated with ASD symptoms and outcome. Links between dopaminergic variants and behavioral phenotypes relevant to ASD, such as joint attention and behavior problems, can aid in understanding the developmental heterogeneity of high-risk siblings. Identification of common genetic variants—assessable at birth—that confer increased risk for ASDrelevant behaviors has the potential to aid in assessing risk and informing preventive interventions. If replicated, the current results suggest that genotype screening could aid in identifying siblings at the greatest risk for difficulties in areas relevant to later outcomes, even before the emergence of delays or difficulties. Developmental psychopathology could benefit from utilizing genetic markers with documented roles in healthy and problematic behaviors to assess risk and inform preventive interventions.

#### REFERENCES

- Abrahams, B. S., & Geschwind, D. H. (2008). Advances in autism genetics: on the threshold of a new neurobiology. *Nature Review Genetics*, 9(5), 341-355. doi: 10.1038/nrg2346
- Achenbach, T. M., Edelbrock, C., & Howell, C. T. (1987). Empirically based assessment of the behavioral/emotional problems of 2- and 3- year-old children. *Journal of Abnormal Child Psychology*, 15(4), 629-650. doi: 10.1007/bf00917246
- Achenbach, T. M., & Rescorla, L. (2000). *Child Behavior Checklist for Ages 1 1/2 5*. Burlington, VT: ASEBA, University of Vermont.
- Anney, R., Klei, L., Pinto, D., Regan, R., Conroy, J., Magalhaes, T. R., . . . Hallmayer, J. (2010). A genome-wide scan for common alleles affecting risk for autism. *Human Molecular Genetics*. doi: 10.1093/hmg/ddq307
- Association, A. P. (2013). *Diagnostic and Statistical Manual of Mental Disorders: DSM-V*. Washington, DC: American Psychiatric Association.
- Auerbach, J. G., Benjamin, J., Faroy, M., Geller, V., & Ebstein, R. (2001). DRD4 related to infant attention and information processing: A developmental link to ADHD? *Psychiatric Genetics*, 11(1), 31-35.
- Auerbach, J. G., Faroy, M., Ebstein, R., Kahana, M., & Levine, J. (2001). The association of the dopamine D4 receptor gene (DRD4) and the serotonin transporter promoter gene (5-HTTLPR) with temperament in 12-month-old infants. *Journal of Child Psychology and Psychiatry*, 42(6), 777-783. doi: 10.1111/1469-7610.00774
- Bakermans-Kranenburg, M. J., & van IJzendoorn, M. H. (2006). Gene-environment interaction of the dopamine D4 receptor (DRD4) and observed maternal insensitivity predicting externalizing behavior in preschoolers. *Developmental Psychobiology*, 48(5), 406-409. doi: 10.1002/dev.20152
- Bakermans-Kranenburg, M. J., & van IJzendoorn, M. H. (2011). Differential susceptibility to rearing environment depending on dopamine-related genes: New evidence and a meta-analysis. *Development and Psychopathology*, 23(01), 39-52. doi: 10.1017/S0954579410000635
- Beaulieu, J.-M., & Gainetdinov, R. R. (2011). The physiology, signaling, and pharmacology of dopamine receptors. *Pharmacological Reviews*, 63(1), 182-217. doi: 10.1124/pr.110.002642
- Belsky, J. (2005). Differential susceptibility to rearing influence: An evolutionary hypothesis and some evidence. In B. Ellis & D. Bjorklund (Eds.), Origins of the social mind: Evolutionary psychology and child development (pp. 139-163). New York: Guilford.

- Belsky, J., Bakermans-Kranenburg, M. J., & van IJzendoorn, M. H. (2007). For better and for worse: Differential susceptibility to environmental influences. *Current Directions in Psychological Science*, 16(6), 300-304. doi: 10.1111/j.1467-8721.2007.00525.x
- Belsky, J., & Pluess, M. (2009). Beyond diathesis stress: Differential susceptibility to environmental influences. *Psychological Bulletin*, 135(6), 885-908. doi: 10.1037/a0017376
- Berument, S. K., Rutter, M., Lord, C., Pickles, A., & Bailey, A. (1999). Autism screening questionnaire: Diagnostic validity. *The British Journal of Psychiatry*, 175, 444-451. doi: 10.1192/bjp.175.5.444
- Betancur, C. (2011). Etiological heterogeneity in autism spectrum disorders: More than 100 genetic and genomic disorders and still counting. *Brain Research*, *1380*(0), 42-77. doi: http://dx.doi.org/10.1016/j.brainres.2010.11.078
- Carver, C. S., Johnson, S. L., & Joormann, J. (2009). Two-mode models of selfregulation as a tool for conceptualizing effects of the serotonin system in normal behavior and diverse disorders. *Current Directions in Psychological Science*, 18(4), 195-199. doi: 10.1111/j.1467-8721.2009.01635.x
- Cassel, T. D., Messinger, D. S., Ibanez, L. V., Haltigan, J. D., Acosta, S. I., & Buchman, A. C. (2007). Early social and emotional communication in the infant siblings of children with autism spectrum disorders: An examination of the broad phenotype. *Journal of Autism and Developmental Disorders*, 37(1), 122-132. doi: 10.1007/s10803-006-0337-1
- CDC. (2014). Prevalence of Autism Spectrum Disorder Among Children Aged 8 Years -Autism and Developmental Disabilities Monitoring Network, 11 Sites, United States, 2010. Centers for Disease Control and Prevention, Morbidity and Mortality Weekly Report, Surveillance Summaries, 63, 2.
- Chen, J., Lipska, B. K., Halim, N., Ma, Q. D., Matsumoto, M., Melhem, S., . . . Weinberger, D. R. (2004). Functional analysis of genetic variation in catechol-Omethyltransferase (COMT): Effects on mRNA, protein, and enzyme activity in postmortem human brain. *Am J Hum Genet*, 75(5), 807-821. doi: 10.1086/425589
- Colvert, E., Tick, B., McEwen, F., Stewart, C., Curran, S. R., Woodhouse, E., . . . Bolton, P. (2015). Heritability of autism spectrum disorder in a UK population-based twin sample. *JAMA Psychiatry*. doi: 10.1001/jamapsychiatry.2014.3028
- Davis, N. O., & Carter, A. (2008). Parenting stress in mothers and fathers of toddlers with Autism Spectrum Disorders: Associations with child characteristics. *Journal of Autism and Developmental Disorders*, 38(7), 1278-1291. doi: 10.1007/s10803-007-0512-z

- Dawson, G., Toth, K., Abbott, R., Osterling, J., Munson, J., Estes, A., & Liaw, J. (2004). Early social attention impairments in autism: Social orienting, joint attention, and attention to distress. *Developmental Psychology*, 40(2), 271-283. doi: 10.1037/0012-1649.40.2.271
- Devlin, B., Melhem, N., & Roeder, K. (2011). Do common variants play a role in risk for autism? Evidence and theoretical musings. *Brain Research*, 1380(0), 78-84. doi: http://dx.doi.org/10.1016/j.brainres.2010.11.026
- Ekas, N., & Whitman, T. L. (2010). Autism symptom topography and maternal socioemotional functioning. *American Journal on Intellectual and Developmental Disabilities*, 115(3), 234-249. doi: 10.1352/1944-7558-115.3.234
- Fisman, S., Wolf, L., Ellison, D., Gillis, B., Freeman, T. O. M., & Szatmari, P. (1996). Risk and protective factors affecting the adjustment of siblings of children with chronic disabilities. *Journal of the American Academy of Child & Adolescent Psychiatry*, 35(11), 1532-1541. doi: http://dx.doi.org/10.1097/00004583-199611000-00023
- Fuke, S., Suo, S., Takahashi, N., Koike, H., Sasagawa, N., & Ishiura, S. (2001). The VNTR polymorphism of the human dopamine transporter (DAT1) gene affects gene expression. *Pharmacogenomics J*, 1(2), 152-156.
- Gadow, K. D., DeVincent, C. J., Olvet, D. M., Pisarevskaya, V., & Hatchwell, E. (2010). Association of DRD4 polymorphism with severity of oppositional defiant disorder, separation anxiety disorder and repetitive behaviors in children with autism spectrum disorder. *European Journal of Neuroscience*, 32(6), 1058-1065. doi: 10.1111/j.1460-9568.2010.07382.x
- Gangi, D. N., Ibañez, L. V., & Messinger, D. S. (2014). Joint attention initiation with and without positive affect: Risk group differences and associations with ASD symptoms. *Journal of Autism and Developmental Disorders*, 44(6), 1414-1424. doi: 10.1007/s10803-013-2002-9
- Gaugler, T., Klei, L., Sanders, S. J., Bodea, C. A., Goldberg, A. P., Lee, A. B., ... Buxbaum, J. D. (2014). Most genetic risk for autism resides with common variation. *Nature Genetics, advance online publication*. doi: 10.1038/ng.3039
- Georgiades, S., Szatmari, P., Zwaigenbaum, L., Bryson, S., Brian, J., Roberts, W., ... Garon, N. (2013). A prospective study of autistic-like traits in unaffected siblings of probands with autism spectrum disorder. *JAMA Psychiatry*, 70(1), 42-48. doi: 10.1001/2013.jamapsychiatry.1
- Geschwind, D. H. (2011). Genetics of autism spectrum disorders. *Trends in Cognitive Sciences*, *15*(9), 409-416. doi: http://dx.doi.org/10.1016/j.tics.2011.07.003

- Gizer, I. R., Ficks, C., & Waldman, I. D. (2009). Candidate gene studies of ADHD: A meta-analytic review. *Human Genetics*, *126*(1), 51-90. doi: 10.1007/s00439-009-0694-x
- Goggi, J., Pullar, I. A., Carney, S. L., & Bradford, H. F. (2003). Signalling pathways involved in the short-term potentiation of dopamine release by BDNF. *Brain Research*, 968(1), 156-161. doi: http://dx.doi.org/10.1016/S0006-8993(03)02234-0
- Goldberg, W. A., Jarvis, K. L., Osann, K., Laulhere, T. M., Straub, C., Thomas, E., . . . Spence, M. A. (2005). Brief Report: Early social communication behaviors in the younger siblings of children with autism. *Journal of Autism and Developmental Disorders*, 35(5), 657-664. doi: 10.1007/s10803-005-0009-6
- Gotham, K., Pickles, A., & Lord, C. (2009). Standardizing ADOS scores for a measure of severity in autism spectrum disorders. *Journal of Autism and Developmental Disorders*, 39(5), 693-705. doi: 10.1007/s10803-008-0674-3
- Grønborg, T. K., Schendel, D. E., & Parner, E. T. (2013). Recurrence of autism spectrum disorders in full- and half-siblings and trends over time: A population-based cohort study. *JAMA Pediatrics*, -. doi: 10.1001/jamapediatrics.2013.2259
- Hallmayer, J., Cleveland, S., Torres, A., Phillips, J., Cohen, B., Torigoe, T., ... Risch, N. (2011). Genetic heritability and shared environmental factors among twin pairs with autism. *Archives of General Psychiatry*, 68(11), 1095-1102. doi: 10.1001/archgenpsychiatry.2011.76
- Hettinger, J. A., Liu, X., Hudson, M. L., Lee, A., Cohen, I. L., Michaelis, R. C., . . . Holden, J. J. (2012). DRD2 and PPP1R1B (DARPP-32) polymorphisms independently confer increased risk for autism spectrum disorders and additively predict affected status in male-only affected sib-pair families. *Behavioral and Brain Functions*, 8(1), 19.
- Howlin, P., Goode, S., Hutton, J., & Rutter, M. (2004). Adult outcome for children with autism. *Journal of Child Psychology and Psychiatry*, 45(2), 212-229. doi: 10.1111/j.1469-7610.2004.00215.x
- Hus, V., Gotham, K., & Lord, C. (2014). Standardizing ADOS domain scores: Separating severity of social affect and restricted and repetitive behaviors. *Journal of Autism* and Developmental Disorders, 44(10), 2400-2412. doi: 10.1007/s10803-012-1719-1
- Ibañez, L. V., Grantz, C. J., & Messinger, D. S. (2012). The Development of Referential Communication and Autism Symptomatology in High-Risk Infants. *Infancy*, n/an/a. doi: 10.1111/j.1532-7078.2012.00142.x

- Klei, L., Sanders, S., Murtha, M., Hus, V., Lowe, J., Willsey, A., . . . Devlin, B. (2012). Common genetic variants, acting additively, are a major source of risk for autism. *Molecular Autism*, 3(1), 9.
- Lackner, C., Sabbagh, M. A., Hallinan, E., Liu, X., & Holden, J. J. A. (2012). Dopamine receptor D4 gene variation predicts preschoolers' developing theory of mind. *Developmental Science*, 15(2), 272-280. doi: 10.1111/j.1467-7687.2011.01124.x
- Lord, C., Risi, S., Lambrecht, L., Cook, E. H., Jr., Leventhal, B. L., DiLavore, P. C., ... Rutter, M. (2000). The Autism Diagnostic Observation Schedule—Generic: A standard measure of social and communication deficits associated with the spectrum of autism. *Journal of Autism and Developmental Disorders*, 30(3), 205-223. doi: 10.1023/a:1005592401947
- Lord, C., Rutter, M., & Couteur, A. (1994). Autism Diagnostic Interview-Revised: A revised version of a diagnostic interview for caregivers of individuals with possible pervasive developmental disorders. *Journal of Autism and Developmental Disorders, 24*(5), 659-685. doi: 10.1007/bf02172145
- Mahan, S., & Matson, J. L. (2011). Children and adolescents with autism spectrum disorders compared to typically developing controls on the Behavioral Assessment System for Children, Second Edition (BASC-2). *Research in Autism Spectrum Disorders*, 5(1), 119-125. doi: http://dx.doi.org/10.1016/j.rasd.2010.02.007
- Malesa, E., Foss-Feig, J., Yoder, P., Warren, Z., Walden, T., & Stone, W. (2012). Predicting language and social outcomes at age 5 for later-born siblings of children with autism spectrum disorders. *Autism.* doi: 10.1177/1362361312444628
- Maskey, M., Warnell, F., Parr, J. R., Couteur, A., & McConachie, H. (2013). Emotional and behavioural problems in children with Autism Spectrum Disorder. *Journal of Autism and Developmental Disorders*, 43(4), 851-859. doi: 10.1007/s10803-012-1622-9
- Mesman, J., Stoel, R., Bakermans-Kranenburg, M. J., IJzendoorn, M. H., Juffer, F., Koot, H. M., & Alink, L. R. A. (2009). Predicting growth curves of early childhood externalizing problems: Differential susceptibility of children with difficult temperament. *Journal of Abnormal Child Psychology*, 37(5), 625-636. doi: 10.1007/s10802-009-9298-0
- Messinger, D., Young, G. S., Ozonoff, S., Dobkins, K., Carter, A., Zwaigenbaum, L., ... Sigman, M. (2013). Beyond autism: A Baby Siblings Research Consortium study of high-risk children at three years of age. *Journal of the American Academy of Child and Adolescent Psychiatry*, 52(3), 300-308.e301.

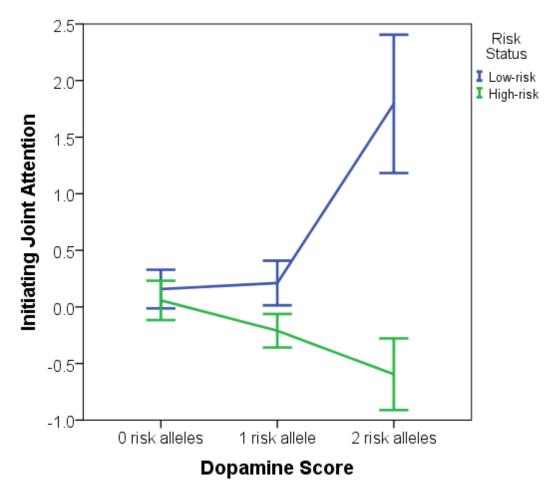
- Muhle, R., Trentacoste, S. V., & Rapin, I. (2004). The genetics of autism. *Pediatrics*, 113(5), e472-e486.
- Mundy, P., Delgado, C., Block, J., Venezia, M., Hogan, A., & Siebert, J. (2003). *A* manual for the abridged Early Social Communication Scales (ESCS). Coral Gables, FL: University of Miami, Department of Psychology.
- Mundy, P., Sigman, M., Ungerer, J., & Sherman, T. (1986). Defining the social deficits of autism: The contribution of non-verbal communication measures. *Journal of Child Psychology and Psychiatry*, 27(5), 657-669. doi: 10.1111/j.1469-7610.1986.tb00190.x
- Narita, M., Aoki, K., Takagi, M., Yajima, Y., & Suzuki, T. (2003). Implication of brainderived neurotrophic factor in the release of dopamine and dopamine-related behaviors induced by methamphetamine. *Neuroscience*, 119(3), 767-775. doi: http://dx.doi.org/10.1016/S0306-4522(03)00099-X
- Newschaffer, C. J., Croen, L. A., Fallin, M. D., Hertz-Picciotto, I., Nguyen, D. V., Lee, N. L., . . . Shedd-Wise, K. M. (2012). Infant siblings and the investigation of autism risk factors. *Journal of Neurodevelopmental Disorders*, 4, 7.
- Nikolova, Y. S., Ferrell, R. E., Manuck, S. B., & Hariri, A. R. (2011). Multilocus genetic profile for dopamine signaling predicts ventral striatum reactivity. *Neuropsychopharmacology*, *36*(9), 1940-1947. doi: 10.1038/npp.2011.82
- Ozonoff, S., Young, G. S., Carter, A., Messinger, D., Yirmiya, N., Zwaigenbaum, L., . . . Stone, W. L. (2011). Recurrence risk for autism spectrum disorders: A Baby Siblings Research Consortium study. *Pediatrics*, *128*(3), e488-e495.
- Pearson-Fuhrhop, K. M., Dunn, E. C., Mortero, S., Devan, W. J., Falcone, G. J., Lee, P., . . Cramer, S. C. (2014). Dopamine genetic risk score predicts depressive symptoms in healthy adults and adults with depression. *PLoS ONE*, 9(5), e93772. doi: 10.1371/journal.pone.0093772
- Pearson, D. A., Loveland, K. A., Lachar, D., Lane, D. M., Reddoch, S. L., Mansour, R., & Cleveland, L. A. (2006). A comparison of behavioral and emotional functioning in children and adolescents with Autistic Disorder and PDD-NOS. *Child Neuropsychology*, 12(4-5), 321-333. doi: 10.1080/09297040600646847
- Posner, M. I., Rothbart, M. K., & Sheese, B. E. (2007). Attention genes. *Developmental Science*, *10*(1), 24-29. doi: 10.1111/j.1467-7687.2007.00559.x
- Preacher, K., & Hayes, A. (2008). Asymptotic and resampling strategies for assessing and comparing indirect effects in multiple mediator models. *Behavior Research Methods*, 40(3), 879-891. doi: 10.3758/BRM.40.3.879

- Rapin, I. (1991). Autistic children: Diagnosis and clinical features. *Pediatrics*, 87(5), 751-760.
- Risch, N., Hoffmann, T. J., Anderson, M., Croen, L. A., Grether, J. K., & Windham, G. C. (2014). Familial recurrence of autism spectrum disorder: Evaluating genetic and environmental contributions. *American Journal of Psychiatry*.
- Rodrigue, J. R., Geffken, G. R., & Morgan, S. B. (1993). Perceived competence and behavioral adjustment of siblings of children with autism. *Journal of Autism and Developmental Disorders*, 23(4), 665-674. doi: 10.1007/bf01046108
- Rodriguez, S., Gaunt, T. R., & Day, I. N. M. (2009). Hardy-Weinberg Equilibrium Testing of Biological Ascertainment for Mendelian Randomization Studies. *American Journal of Epidemiology*, 169(4), 505-514. doi: 10.1093/aje/kwn359
- Rozga, A., Hutman, T., Young, G. S., Rogers, S. J., Ozonoff, S., Dapretto, M., & Sigman, M. (2011). Behavioral profiles of affected and unaffected siblings of children with autism: Contribution of measures of mother–infant interaction and nonverbal communication. *Journal of Autism and Developmental Disorders*, 41(3), 287-301. doi: 10.1007/s10803-010-1051-6
- Rutter, M., & Schopler, E. (1987). Autism and pervasive developmental disorders: Concepts and diagnostic issues. *Journal of Autism and Developmental Disorders*, *17*(2), 159-186. doi: 10.1007/bf01495054
- Salem, A. M., Ismail, S., Zarouk, W. A., Abdul Baky, O., Sayed, A. A., Abd El-Hamid, S., & Salem, S. (2013). Genetic variants of neurotransmitter-related genes and miRNAs in Egyptian autistic patients. *The Scientific World Journal*, 2013, 7. doi: 10.1155/2013/670621
- Savitz, J., Solms, M., & Ramesar, R. (2006). The molecular genetics of cognition: dopamine, COMT and BDNF. *Genes, Brain and Behavior, 5*(4), 311-328. doi: 10.1111/j.1601-183X.2005.00163.x
- Schmidt, L. A., Fox, N. A., Perez-Edgar, K., & Hamer, D. H. (2009). Linking gene, brain, and behavior: DRD4, frontal asymmetry, and temperament. *Psychological Science*, 20(7), 831-837. doi: 10.1111/j.1467-9280.2009.02374.x
- Schmidt, L. A., Fox, N. A., Perez-Edgar, K., Hu, S., & Hamer, D. H. (2001). Association of DRD4 with attention problems in normal childhood development. *Psychiatric Genetics*, 11(1), 25-29.
- Schoots, O., & Van Tol, H. H. M. (2003). The human dopamine D4 receptor repeat sequences modulate expression. *Pharmacogenomics Journal*, *3*(6), 343-348.

- Sheese, B. E., Voelker, P. M., Rothbart, M. K., & Posner, M. I. (2007). Parenting quality interacts with genetic variation in dopamine receptor D4 to influence temperament in early childhood. *Development and Psychopathology*, 19(04), 1039-1046. doi: doi:10.1017/S0954579407000521
- Stice, E., Yokum, S., Burger, K., Epstein, L., & Smolen, A. (2012). Multilocus genetic composite reflecting dopamine signaling capacity predicts reward circuitry responsivity. *The Journal of Neuroscience*, 32(29), 10093-10100. doi: 10.1523/jneurosci.1506-12.2012
- Talkowski, M. E., Minikel, E. V., & Gusella, J. F. (2014). Autism spectrum disorder genetics: Diverse genes with diverse clinical outcomes. *Harvard Review of Psychiatry*, 22(2), 65-75. doi: 10.1097/HRP.0000000000000002
- Thompson, J., Thomas, N., Singleton, A., Piggot, M., Lloyd, S., Perry, E. K., . . . Court, J. A. (1997). D2 dopamine receptor gene (DRD2) Taql A polymorphism: reduced dopamine D2 receptor binding in the human striatum associated with the A1 allele. *Pharmacogenetics and Genomics*, 7(6), 479-484.
- Toth, K., Dawson, G., Meltzoff, A., Greenson, J., & Fein, D. (2007). Early social, imitation, play, and language abilities of young non-autistic siblings of children with autism. *Journal of Autism and Developmental Disorders*, 37(1), 145-157. doi: 10.1007/s10803-006-0336-2
- Van IJzendoorn, M. H., & Bakermans-Kranenburg, M. J. (2006). DRD4 7-repeat polymorphism moderates the association between maternal unresolved loss or trauma and infant disorganization. *Attachment & Human Development*, 8(4), 291-307. doi: 10.1080/14616730601048159
- Verté, S., Roeyers, H., & Buysse, A. (2003). Behavioural problems, social competence and self-concept in siblings of children with autism. *Child: Care, Health and Development, 29*(3), 193-205. doi: 10.1046/j.1365-2214.2003.00331.x
- Yirmiya, N., Gamliel, I., Pilowsky, T., Feldman, R., Baron-Cohen, S., & Sigman, M. (2006). The development of siblings of children with autism at 4 and 14 months: Social engagement, communication, and cognition. *Journal of Child Psychology and Psychiatry*, 47(5), 511-523. doi: 10.1111/j.1469-7610.2005.01528.x
- Yuen, R. K. C., Thiruvahindrapuram, B., Merico, D., Walker, S., Tammimies, K., Hoang, N., . . . Scherer, S. W. (2015). Whole-genome sequencing of quartet families with autism spectrum disorder. *Nature Medicine, advance online publication*. doi: 10.1038/nm.3792

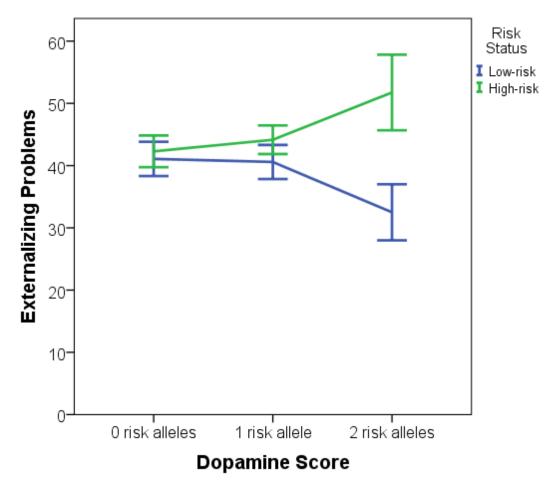
## Figure 1

Mean levels of Initiating Joint Attention (IJA) by group.



*Note.* Error bars reflect +/- 1 SE. Initiating joint attention reflects a mean of standardized values. In high-risk siblings, 28 had a dopamine score of 0, 18 had a score of 1, and 8 had a score of 2. In low-risk siblings, 19 had a dopamine score of 0, 14 had a score of 1, and 3 had a score of 2.

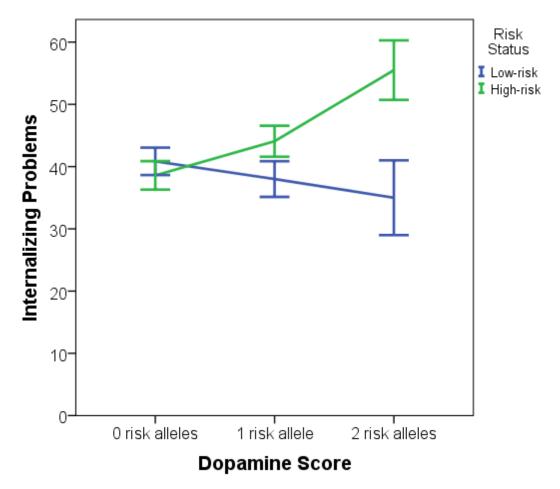
## Figure 2



Mean levels of CBCL Externalizing by group.

*Note.* Error bars reflect +/- 1 SE. The dopamine score\*status interaction effect was not significant for Externalizing Problems. In high-risk siblings, 17 had a dopamine score of 0, 13 had a score of 1, and 4 had a score of 2. In low-risk siblings, 13 had a dopamine score of 0, 12 had a score of 1, and 2 had a score of 2.

## Figure 3



Mean levels of CBCL Internalizing by group.

*Note.* Error bars reflect  $\pm -1$  SE. In high-risk siblings, 17 had a dopamine score of 0, 13 had a score of 1, and 4 had a score of 2. In low-risk siblings, 13 had a dopamine score of 0, 12 had a score of 1, and 2 had a score of 2.

	High-risk Siblings		Low-risk Siblings		
	Frequency	Percentage	Frequency	Percentage	
DRD4					
_/_	41	74.5%	24	66.7%	
7/-	13	23.6%	11	30.6%	
7/7	1	1.8%	1	2.8%	
DRD2					
G/G	35	63.6%	28	73.7%	
A/G	19	34.5%	8	21.1%	
A/A	1	1.8%	2	5.3%	
7/7 <i>DRD2</i> G/G A/G	1 35 19	1.8% 63.6% 34.5%	1 28 8	2.8% 73.7% 21.1%	

Table 1a. DRD4 and DRD2 allele frequencies by risk group.

	High-ris	k Siblings	Low-risk Siblings		
	Frequency	Percentage	Frequency	Percentage	
DRD4					
2/2	1	1.8%	1	2.8%	
2/3	1	1.8%	1	2.8%	
2/4	8	14.5%	3	8.3%	
2/7	1	1.8%	2	5.6%	
3/4	3	5.5%	2	5.6%	
3/7	0	0.0%	1	2.8%	
4/4	27	49.1%	17	47.2%	
4/7	10	18.2%	8	22.2%	
4/8	1	1.8%	0	0.0%	
5/7	1	1.8%	0	0.0%	
7/7	1	1.8%	1	2.8%	
7/8	1	1.8%	0	0.0%	

Table 1b. DRD4 repeat allele frequencies by risk group.

	$A_{*}$	SD	No ASD		
	Frequency	Percentage	Frequency	Percentage	
DRD4					
_/_	8	72.7%	27	73.0%	
7/-	3	27.3%	9	24.3%	
7/7	0	0.0%	1	2.7%	
DRD2					
G/G	6	50.0%	25	67.6%	
A/G	6	50.0%	11	29.7%	
A/A	0	0.0%	1	2.7%	

 Table 2. DRD4 and DRD2 allele frequencies of high-risk siblings by ASD diagnosis.

	Hispanic/Latino			lispanic aucasian	Other		
	Frequency Percentage				Frequency	Percentage	
DRD4							
_/_	34	70.8%	27	79.4%	4	44.4%	
7/-	13	27.1%	7	20.6%	4	44.4%	
7/7	1	2.1%	0	0.0%	1	11.1%	
DRD2							
G/G	30	58.8%	26	78.8%	7	77.8%	
A/G	20	39.2%	5	15.2%	2	22.2%	
A/A	1	2.0%	2	6.1%	0	0.0%	

 Table 3. DRD4 and DRD2 allele frequencies by ethnicity.

	High-risk Siblings		Low-rish	k Siblings
	Frequency	Percentage	Frequency	Percentage
DRD4				
7-repeat allele	14	25.5%	12	33.3%
No 7-repeat allele	41	74.5%	24	66.7%
DRD2				
A allele	20	36.4%	10	26.3%
No A allele	35	63.6%	28	73.7%
Dopamine Score				
0 risk alleles	28	51.9%	19	52.8%
1 risk allele	18	33.3%	14	38.9%
2 risk alleles	8	14.8%	3	8.3%

 Table 4. Genotype frequencies by risk group.

	$A_{*}$	SD	No ASD		
	Frequency	Percentage	Frequency	Percentage	
DRD4					
7-repeat allele	3	27.3%	10	27.0%	
No 7-repeat allele	8	72.7%	27	73.0%	
DRD2					
A allele	6	50.0%	12	32.4%	
No A allele	6	50.0%	25	67.6%	
Dopamine Score					
0 risk alleles	5	45.5%	19	51.4%	
1 risk allele	3	27.3%	14	37.8%	
2 risk alleles	3	27.3%	4	10.8%	

Table 5. Genotype frequencies of high-risk siblings by ASD diagnosis.

Table 6.	Genotype	frequencies	bv	ethnicity.
14010 0.	Senorype.	<i>j</i> : <i>equeneres</i>	0,	connocry.

	Non-Hispanic						
	Hispanic/Latino		White/Caucasian		Other		
	Frequency	Percentage	Frequency	Percentage	Frequency	Percentage	
DRD4							
7-repeat allele	14	29.2%	7	20.6%	5	55.6%	
No 7-repeat allele	34	70.8%	27	79.4%	4	44.4%	
DRD2							
A allele	21	41.2%	7	21.2%	2	22.2%	
No A allele	30	58.8%	26	78.8%	7	77.8%	
Dopamine Score							
0 risk alleles	21	43.8%	22	66.7%	4	44.4%	
1 risk allele	21	43.8%	8	24.2%	3	33.3%	
2 risk alleles	6	12.5%	3	9.1%	2	22.2%	