# Marquette University e-Publications@Marquette

Dissertations (2009 -)

Dissertations, Theses, and Professional Projects

# Synthetic and Mechanistic Studies of Coupling Reactions Involving C-H Bond Activation

Ki Hyeok Kwon Marquette University

**Recommended** Citation

Kwon, Ki Hyeok, "Synthetic and Mechanistic Studies of Coupling Reactions Involving C-H Bond Activation" (2011). *Dissertations* (2009 -). Paper 134. http://epublications.marquette.edu/dissertations\_mu/134

# SYNTHETIC AND MECHANISTIC STUDIES OF COUPLING REACTIONS INVOLVING C-H BOND ACTIVATION

by

Ki Hyeok Kwon, B.S., M.S.

A Dissertation submitted to the Faculty of the Graduate School, Marquette University, in Partial Fulfillment of the Requirements for the Degree of Doctor of Philosophy

Milwaukee, Wisconsin

August 2011

## ABSTRACT SYNTHETIC AND MECHANISTIC STUDIES OF COUPLING REACTION INVOLVING C-H BOND ACTIVATION

Ki Hyeok Kwon, B.S., M.S.

#### Marquette University, 2011

Transition metal-catalyzed C-C bond forming reactions involving C-H bond activation have been shown to be effective methods for functionalization of unreactive compounds. Since Murai's pioneering work on ruthenium-catalyzed regioselective arene-to-alkene coupling reactions, a number of well-defined, late transition metal catalysts have been shown to mediate regioseletive C-C bond forming reaction involving C-H bond activation. Recently, late transition metal complexes have also been found to catalyze the regioselective coupling reaction of nitrogen containing compounds with alkenes as well as sp<sup>3</sup> bond insertions and oxidative coupling reaction of aimdes. The catalytic C-C bond forming reaction involving C-H bond activation would provide simple and atom economic pathways for making functionalized molecules. Although there are many examples of stoichiometric reaction of aromatic C-H bond activation with transition metal compounds, catalytic C-H bond activation reactions via catalytic system have been rarely employed in organic synthesis.

An efficient catalytic conjugate addition reaction protocol has been developed for the synthesis of tetrasubstituted olefin products. The cationic ruthenium-hydride complex was found to be an effective catalyst for coupling reaction with  $\alpha$ , $\beta$ -unsaturated carbonyl compounds and simple alkenes. The kinetic and spectroscopic data are consistent with three different mechanistic pathways depending on substrate employed. The reaction of an  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with simple olefins predominantly gave the tetrasubstituted olefin products. The mechanistic studies suggest that the olefin insertion into an  $\alpha$ , $\beta$ -unsaturated carbonyl substrate is the rate-limiting step for conjugate addition reaction. On the other hand, the coupling reaction of  $\alpha$ -methyl  $\alpha$ , $\beta$ -unsaturated cinnamide with aromatic alkenes gave oxidative coupling products. The kinetic and spectroscopic studies support the different mechanistic pathway, which involves the rate-limiting vinyl C-H activation step, in comparison with the conjugate addition reaction. This synthetic methodology promises to provide a straightforward route to the valuable organic compounds such as tetrasubstituted olefins which are difficult to synthesize by using traditional synthetic methods.

# TABLE OF CONTENTS

LIST OF TABLES iv
LIST OF FIGURESvi
CHAPTER
I. INTRODUCTION1
1.1 C-H bond Activation of Alkanes2
1.2 Arene C-H Bond Activation 8
1.2.1 Chelate-Assisted Arene C-H bond Activation Reactions
1.2.2 Nonchelate-Asssisted Reactions
1.3 Chelate-Assisted C-H Bond Activation Reaction of Alkenes18
1.4 C-H Bond Activation Reactions of Nitrogen Containing Compounds
1.4.1 C-H Bond Activation of Pyrrolidine Derivatives
1.4.2 C-H Bond Activation of Indole and Quinoline derivatives25
1.4.3 C-H Bond Activation of Pyridine derivatives
1.4.4 C-H Bond Activation of Amide, Amine, and imine derivatives33
II. Synthetic and Mechanistic Studies of Chelate-Assisted Oxidative Coupling Reaction of Arylamides and Unactivated Alkenes
2.1 Optimization Studies for Coupling of Arylamide with cyclopentene 45
2.1.1 Catalyst Survey45
2.1.2 Solvent Effect46
2.2 Reaction Scope 48

2.3 Mechanistic Study51
2.3.1 Phosphine Inhibition51
2.3.2 Isotope Effect Study
2.3.3 Deuterium Labeling Study13
2.3.4 Isolation of Catalytically Relevant Species
2.3.5 VT NMR Study
2.4 Proposed Mechanism 63
III. Stereoselective Catalytic Conjugate Addition and Oxidative Coupling Reaction of Simple Alkenes to α,β-Unsaturated Carbonyl Compounds 67
3.1 Catalyst Survey71
3.2 Reaction Scope72
3.2.1 Coupling Reactions of α,β-Unsaturated Ester and Enones with Alkenes
3.2.2 Coupling Reactions of $\alpha$ , $\beta$ -Unsaturated Amides with Alkenes75
3.2.3 Coupling Reactions of α-Substituted α,β-Unsaturated Carbonyl Compounds with Alkenes
3.2.4 Oxidative Coupling Reactions of α-Substituted α,β-Unsaturated Carbonyl Compounds with Aromatic Alkenes81
3.4.5 Intramolecular Coupling Reaction of α,β-Unsaturated Esters with Simple Olefins
3.3 Mechanistic Studies
3.3.1 Reaction Profile
3.3.2 Hammett Study 89
3.3.3 Isotope Effect Studies
3.3.4 Deuterium Labeling Studies

3.3.5 Isolation of Catalytically	Relevant Species105
3.4 Empirical Rate Law Derivation	on 107
3.5 Thermodynamic Parameters .	
3.6 Proposed Mechanism	
<ul><li>IV. Aqueous Phase C-H Bond Oxidation Water-Soluble Cationic Ru(III) Pymo</li><li>4.1 Synthesis of Water-Soluble R</li></ul>	Reaction of Arylalkanes Catalyzed by a ox Complex118 uthenium Pymox Complexes
4.2 Optimization Studies for C-H	Oxidation Reaction of Arylalkanes135
4.2.1 Catalyst Survey	
4.2.2 Survey of Oxidizing Age	nts137
4.2.3 Reusability of the Cation Solution	c Ruthenium Complex in Aqueous
4.3 Reaction Scope	
4.4 Mechanistic Studies	
4.4.1 Isotope Effect Study	
4.4.2 Hammett Study	
4.4.3 Detection of Catalytically	Relevant Species144
4.5 Proposed Mechanism	
V. Experimental Section	151
References	
Appendix A (Molecular Structures of 90-	. <b>99</b> )
Appendix B (Crystal Data and Structure)	Refinement for <b>90-99</b> )218

## LIST OF TABLES

<b>Table 1.</b> Catalyst Suvery on the Reaction of $C_6H_5CONMe_2$ with Cyclopentene
<b>Table 2.</b> Solvent Effect on the Coupling Reaction of $C_6H_5CONMe_2$ with CyclopenteneCatalyzed by 26.47
<b>Table 3</b> . Oxidative C-H Coupling Reaction of Arylamides with Alkenes
<b>Table 4.</b> Average ${}^{13}$ C Integration of the Recovered and Virgin Sample of C <sub>6</sub> H <sub>5</sub> CONEt <sub>2</sub> .54
<b>Table 5.</b> Catalyst Survey for the Coupling Reaction of $(E)$ -C <sub>6</sub> H <sub>5</sub> CH=CHCO <sub>2</sub> Et and Propylene
<b>Table 6.</b> Conjugate Addition Reaction of Simple Olefins to α.β-Unsaturated Ester and Enones
<b>Table 7.</b> Conjugate Addition Reaction of Simple Olefins to α.β-Unsaturated         Amides       77
<b>Table 8.</b> Conjugate Addition Reaction of Simple Olefins to α-Substituted α.β-         Unsaturated Compounds       80
<b>Table 9.</b> Oxidative Coupling Reaction of Aromatic Olefins to α-Substituted α.β-         Unsaturated Amides
<b>Table 10.</b> Intramolecular Coupling Reaction of α,β-Unsaturated Esters with Simple         Olefins Catalyzed by 26
<b>Table 11.</b> Average ${}^{13}$ C Integration of the Recovered and Virgin Samples of (E)- $C_6H_5CH=CHCONEt_2$
<b>Table 12.</b> Average ${}^{13}$ C Integration of the Recovered and Virgin Samples of (E)- $C_6H_5CH=C(CH_3)CONHCH_3$
<b>Table 13.</b> Average ${}^{13}$ C Integration of the Recovered and Virgin Samples of (E)- $C_6H_5CH=C(CH_3)CONHCH_3$
<b>Table 14.</b> Kinetics and Isotope Effects for the Coupling Reaction of $\alpha,\beta$ -UnsaturatedCarbonyl Compounds with Alkene
<b>Table 15.</b> Kinetics, Intra- and Intermolecular Deuterium Isotope Effects for the Ruthenium Catalyzed Oxidation of Hydrocarbons with <i>t</i> -BuOOH

Table	<b>16.</b> Catalyst Survey for Oxidation Reaction of Ethylbenzene with <i>t</i> -BuOOH	.136
Table	<b>17.</b> Oxidizing Agent and Additive Effects on Oxidation of Ethylbenzene	137
Table	<b>18.</b> Aqueous Phase C-H Bond Oxidation of Arylalkanes	141
Table	<b>19</b> . Crystal Data and Structure Refinement for <b>30a</b>	197
Table	<b>20</b> . Crystal Data and Structure Refinement for <b>56</b>	198
Table	<b>21</b> . Crystal Data and Structure Refinement for <b>80</b>	199
Table	<b>22</b> . Crystal Data and Structure Refinement for <b>81</b>	200
Table	<b>23</b> . Crystal Data and Structure Refinement for <b>82</b>	201

## LIST OF FIGURES

<i>Figure 1</i> . Molecular Structure of Cationic Ruthenium-Hydride Complex <b>26</b>
<i>Figure 2.</i> PCy <sub>3</sub> Inhibition Study for the Coupling Reaction of C <sub>6</sub> H <sub>5</sub> CONEt <sub>2</sub> with Cyclopentene Catalyzed by <b>26</b>
<i>Figure 3</i> . First-Order Plots of $-ln([C_6H_5CONEt_2]_t/[C_6H_5CONEt_2]_0)$ vs Time for Coupling Reaction
<b>Figure 4.</b> (a) <sup>1</sup> H NMR Spectrum of Mixture of <b>23h</b> - $d_n$ and <b>24h</b> - $d_n$ , (b) <sup>2</sup> H NMR Spectrum of Mixture of <b>23h</b> - $d_n$ and <b>24h</b> - $d_n$ , (c) <sup>1</sup> H NMR Spectrum <b>24h</b> , (d) <sup>1</sup> H NMR Spectrum of <b>24h</b> - $d_n$ . 57
<b>Figure 5</b> . (a) <sup>1</sup> H NMR Spectrum of <i>N</i> , <i>N</i> -Diethyl Benzamide- $d_5$ , (b) <sup>1</sup> H NMR Spectrum of Recovered <i>N</i> , <i>N</i> -Diethyl Benzamide- $d_n$ , (c) <sup>2</sup> H NMR Spectrum of Recovered <i>N</i> , <i>N</i> -Diethyl Benzamide- $d_n$ , (d) <sup>2</sup> H NMR Spectrum of Recovered Cyclopentene- $d_n$ and Cyclopentane- $d_n$
<i>Figure 6.</i> Molecular Structure of <b>30</b>
<i>Figure 7.</i> van't Hoff Plot of <i>ln K</i> vs 1/T
<i>Figure 8.</i> Structure of Tamoxifen <b>33</b> and Rofecoxib <b>34</b>
<i>Figure 9</i> . Newman Projection of Compound <b>36t.</b>
<i>Figure 10</i> . Newman Projection of Compounds <b>371</b> and <b>37m</b>
<i>Figure 11.</i> Kinetic Profile of the Conversion of <b>44</b> to <b>35</b> g
<b>Figure 12.</b> Hammett Plot of the Coupling Reaction of <i>para</i> -Substituted ( <i>E</i> )- <i>p</i> -X-C <sub>6</sub> H <sub>4</sub> CH=CHCO <sub>2</sub> Et (X = OCH <sub>3</sub> , CH <sub>3</sub> , H, F, Cl, CO <sub>2</sub> Me, CF <sub>3</sub> ) with Propene
<b>Figure 13.</b> Hammett Plot of the Coupling Reaction of $(E)$ -C <sub>6</sub> H <sub>4</sub> CH=CHCO <sub>2</sub> Et with <i>para</i> -Substituted Styrene <i>p</i> -Y-C <sub>6</sub> H <sub>4</sub> CH=CH <sub>2</sub> (Y = OCH <sub>3</sub> , CH <sub>3</sub> , H, Cl, CF <sub>3</sub> )
<b>Figure 14.</b> Hammett Plot of the Coupling Reaction of $(E)$ -C <sub>6</sub> H <sub>4</sub> CH=C(CH <sub>3</sub> )CONHCH <sub>3</sub> with <i>para</i> -Substituted Styrene <i>p</i> -Y-C <sub>6</sub> H <sub>4</sub> CH=CH <sub>2</sub> (Y = OCH <sub>3</sub> , CH <sub>3</sub> , H, F, Cl, CF <sub>3</sub> ) 92
<b>Figure 15.</b> First-Order Plots of $-ln[cinnamate]_t/[cinnamate]_0$ vs Time for the Coupling Reaction of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CH=CHCO <sub>2</sub> Et with Ethylene or Ethylene- <i>d</i> <sub>4</sub>
<b>Figure 16.</b> First-Order Plots of $-ln([cinnamide]_t/[cinnamide]_0)$ vs Time for the Coupling Reaction of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CH=C(CH <sub>3</sub> )CONHCH <sub>3</sub> with Styrene or Styrene- <i>d</i> <sub>8</sub>

<i>Figure 32.</i> Cyclic Voltammogram of <b>80</b> in CH <sub>2</sub> Cl <sub>2</sub>
<b>Figure 31.</b> Hammett plot of the C-H Bond Oxidation Reaction of <i>para</i> -Substituted Ethylbenzene Substrates $p$ -X-C <sub>6</sub> H <sub>4</sub> CH <sub>2</sub> CH <sub>3</sub> (X = OMe, CH <sub>3</sub> , H, F, Cl) with <i>t</i> -BuOOH
<i>Figure 30.</i> First-Order Plots of $-ln([ethylbenzene]_t/[ethylbenzene]_0)$ vs Time for C-H Bond Oxidation Reaction of Ethylbenzene or Ethylbenzene- $d_{10}$ with <i>t</i> -BuOOH 143
<i>Figure 29.</i> Molecular Structure of <b>82.</b>
<i>Figure 28.</i> Molecular Structure of <b>81.</b>
<i>Figure 27.</i> Molecular Structure of <b>80.</b>
<i>Figure 26.</i> Selectivity for catalyzed C-H bond oxidation by [Fe( <i>S</i> , <i>S</i> -PDP)(CH <sub>3</sub> CN) <sub>2</sub> ] [SbF <sub>6</sub> ] <sub>2</sub> , <b>70</b>
<i>Figure 25.</i> Arrhenius Plot for the Coupling Reaction of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CH=CHCO <sub>2</sub> Et with Propene
<i>Figure 24</i> . Eyring Plot for the Coupling Reaction of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CH=CHCO <sub>2</sub> Et with Propene
<i>Figure 23.</i> Plot of Observed Reaction Rate vs ( $E$ )-C <sub>6</sub> H <sub>5</sub> CH=CHCO <sub>2</sub> Et Concentration for the Coupling Reaction
<i>Figure 22.</i> Plot of Observed Reaction Rate vs Propene Concentration for the Coupling Reaction
<i>Figure 21.</i> Plot of Observed Reaction Rate vs Catalyst Concentration for the Coupling Reaction
<i>Figure 20.</i> Molecular Structure of <b>56.</b>
<b>Figure 19.</b> (a) <sup>2</sup> H NMR Spectrum of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CD=C(CH <sub>3</sub> )CONHCH <sub>3</sub> , (b) <sup>2</sup> H NMR Spectrum of the Coupling Reaction Mixture of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CD=C(CH <sub>3</sub> )CONHCH <sub>3</sub> with Styrene
<b>Figure 18.</b> (a) <sup>1</sup> H NMR Spectrum of <b>36c</b> - $d_n$ , (b) <sup>2</sup> H NMR Spectrum of <b>36c</b> - $d_n$ , (c) <sup>2</sup> H NMR Spectrum of Recovered Propene
<b>Figure 17.</b> First-Order Plots of $-ln([cinnamide]_t/[cinnamide]_0)$ vs Time for the Coupling Reaction of ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CH=C(CH <sub>3</sub> )CONHMe or ( <i>E</i> )-C <sub>6</sub> H <sub>5</sub> CD=C(CH <sub>3</sub> )CONHMe with Styrene

Figure	<b>33.</b> U	V-vis spectra	of 82 (20 µN	<b>1</b> ), <b>82</b> (20	) μM)/t-Bu	OOH (30	equiv)/Na	$BF_4$ , and
80 (20)	μM) in	water	· · · · · · · · · · · · · · • • • • • •					146

# **Chapter I**

## Introduction

Over the past few decades, one of the main goals in catalytic chemistry has been the quest for the most economic ways to form C-C bonds.<sup>1</sup> Transition metal-catalyzed cross-coupling reactions have been one of the most useful synthetic tools for the formation of C-C bonds since Heck firstly reported his pioneering work in 1968.<sup>2</sup> Even though a broad range of donors and acceptors are amenable for Heck type coupling reactions, these coupling reactions absolutely require an aryl halide (or triflate) with an alkene and a stoichiometric amount of base and palladium catalyst in forming a substituted alkene products. To overcome these drawbacks for Heck type coupling reaction, the formation of C-C bond involving C-H bond activation of unreactive C-H bonds has attracted considerable interest and remains a long term challenge for the chemical and pharmaceutical industries. The catalytic C-H bond activation would provide simple and atom economic pathways for making functionalized molecules. Although there are many examples of stoichiometric aromatic C-H bond activation reactions with transition metal compounds, catalytic C-H bond activation reactions via catalytic system have been rarely employed in organic synthesis.

Generally, it is believed that the formation of C-C bond involving the cleavage of a C-H bond was a difficult process due to its bond strength. In 1993, Murai et al. reported a highly efficient, ruthenium-catalyzed addition of aromatic C-H bonds to olefins.<sup>3</sup> Trost and Murai independently reported ruthenium-catalyzed C-H activation and alkylation of  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with alkenes.<sup>4</sup> Since then, numerous examples of catalyzed C-H bond activation have been extended to a wide variety of aromatic and

olefinic substrates by a number of research groups.<sup>4-14</sup> However, examples of catalytic reactions involving the sp<sup>3</sup> C-H bond activation are still rare. Recently, Yu has reported a remarkably selective and direct olefination of sp<sup>3</sup> C-H bonds for amides by a Pd(II)-catalyzed reaction.<sup>15</sup> Ellman and Bergman have also reported a rhodium mediated stereoselective alkylation of  $\alpha$ , $\beta$ -unsaturated imines via the direct C-H bond activation.<sup>16</sup> Transition-metal-catalyzed regioselective and stereoselective C-H bond functionalizations have attracted much attention due to their synthetic utility in organic synthesis. This chapter provides a literature summary for the transition-metal-catalyzed formation of C-C bond via C-H activation and their applications.

## **1.1 C-H bond Activation of Alkanes**

Alkanes are among the least reactive compounds. Because of their low reactivity particularly at the terminal position, the direct conversion of alkane into industrially important and valuable chemicals, such as linear alcohols,  $\alpha$ -olefine, and amines, has been a long-term challenge in catalytic chemistry. Controlling selectivity for desired products containing a single functional group at the end of an alkyl chain is especially difficult because reactions of alkanes typically form a mixture of products. Certain transition metals were found to selectively react at the terminal carbon of the alkane to produce *n*-alkyl complexes,<sup>17</sup> but these transformations involve stoichiometric amounts of expensive metal reagents. Jensen and Goldman discovered efficient and selective dehydrogenation of linear alkanes to give  $\alpha$ -olefins in the presence of iridium "pincer" complexes (<sup>*i*-Pr</sup>PCP)IrH<sub>2</sub> (<sup>*i*-Pr</sup>PCP = 2,6-bis[di(isopropyl)phosphinomethyl]phenyl) (1) (eq 1).<sup>18</sup> Isomerization limits the yield of the  $\alpha$ -olefin products generated under these reaction

conditions. They suggested that preventing isomerization by modifying catalyst/acceptor combination or efficient separation of the  $\alpha$ -olefin would improve the yields of dehydrogenation products.



The mechanism of  $(PCP)IrH_2$  catalyzed transfer-dehydrogenation has been disclosed for the prototypical substrate/acceptor combination, COA/TBE (cyclooctane/*t*-butylethylene) at 55 °C (Scheme 1). The catalytic cycle may be viewed as the sum of two reaction: (1) hydrogenation of TBE by (PCP)IrH<sub>2</sub> and C-H addition of a second mole of TBE to give (PCP)IrH(*t*-butylvinyl) (**2**), and (2) dehydrogenation of COA by (PCP)IrH(*t*-butylvinyl) have been observed as resting states during catalysis and the ratio of these two complexes is found to be proportional to [TBE].<sup>19</sup> Based on the proportionality constant obtained and the catalytic rate as a function of [TBE] (which reachs a maximum at ca. 0.3 M), the respective rate constants for the hydrogenation and dehydrogenation segments can be obtained. A good agreement is found between reaction rates independently obtained from stoichiometric and catalytic runs. For the overall TBE-hydrogenation reaction, labeling experiments indicate that the rate-limiting step is reductive elimination of TBA (2,2-dimethylbutane) from (PCP)IrH(*t*-butylethyl) (**3**).

rate-limiting step for alkane dehydrogenations is the oxidative C-H addition to iridium center rather than the  $\beta$ -hydride elimination.

## Scheme 1



Recently, our research group has reported that the tetranuclear ruthenium- $\mu$ -oxohydroxo-hydride complex {[(PCy<sub>3</sub>)(CO)RuH]<sub>4</sub>( $\mu$ <sub>4</sub>-O)( $\mu$ <sub>3</sub>-OH)( $\mu$ <sub>2</sub>-OH)} (**4**) is a highly effective catalyst for the transfer dehydrogenation of amines and carbonyl compounds.<sup>20</sup> For example, the initial turnover rate of the dehydrogenation of 2-methylindoline was measured to be 1.9 s<sup>-1</sup> with TON of 7950 after 1 h at 200 °C (eq 2). From a synthetic point of view, the ruthenium catalyst **4** has a number of salient features compared to other dehydrogenation catalysts, in that it is air- and water-stable in solid state and is compatible with a variety of heteroatom functional groups.



Hartwig reported regiospecific functionalization of linear alkanes catalyzed by the rhodium complex Cp\*Rh( $\eta^4$ -Cp\*) (5) under thermal conditions, specifically the coupling of linear alkanes with borane reagents of form primary alkylboranes (eq 3).<sup>14</sup> These borane products are valuable synthetic precusor in organic chemistry. They can be easily converted to amines, alcohols, alkenes, and other classes of functionalized molecules.



A detailed mechanism for the overall catalytic reaction has not been deduced, but an outline of possible mechanism is provided in Scheme 2. Oxidative addition of borane or diboron compound, depending on the boron reagent present, could form a hydridorhodium boryl complex or a rhodium bis(boryl) complex. After  $H_2$  or borane elimination, such complex could then react with alkane to produce an alkylrhodium boryl

complex (6). Rapid B-C bond formation would then lead to product. To probe for the presence of metal-boryl intermediates, they obtained <sup>11</sup>B NMR spectra of reactions of the diboron compound with *n*-octane at high catalyst loads. They observed a signal at  $\delta$  40 ppm during the reaction, and this chemical shift, which is located downfield of the organoboronate esters and pinacolborane, is characteristic of a metal-boryl species.





Direct oxidative functionalization of tertiary amines is of considerable importance in organic synthesis. Propargylic amines are of considerable pharmaceutical interest and are synthetic intermediates for the synthesis of various nitrogen containing compounds as well as carbohydrates. Li reported an oxidative cross-coupling of amines and alkynes via a direct sp<sup>3</sup> C-H bond alkynylation in presence of CuBr as a catalyst. The reaction of N,N-dimethylaniline with phenylacetylene catalyzed by CuBr with *t*-BOOH at 100 °C for 3 h was produced to *N*-methyl-*N*-(3-phenylprop-2-ynyl)benzenamine in 77% isolated yield (eq 4).<sup>21</sup>

$$\begin{array}{c} \swarrow -N + H = & \swarrow & \frac{CuBr}{t-BuOOH, 100 \ \ \ C, 3h} \\ \end{array}$$

$$\begin{array}{c} \searrow -N \\ \hline N \\ \hline 77\% \end{array}$$

$$(4)$$

Rhodium carbenoids derived from methyl aryldiazoacetates are efficient catalyst for asymmetric C-H activation of a range of alkanes and tetrahydrofuran through a C-H insertion mechanism. The decomposition of methyl aryldiazoacetates catalyzed by dirhodium tetrakis(*S*-(*N*-dodecylbenzenesulfonyl)prolinate) (Rh<sub>2</sub>(*S*-DOSP)<sub>4</sub>) (7) in the presence of alkenes gave an intermolecular C-H insertion having a good control of regioselectivity, diastereoselectivity, and enantioselectivity (eq 5).<sup>22</sup> The carbenoids derived from methyl aryldiazoacetates are greatly more chemoselective than those derived from diazoacetates. They favor C-H insertions into secondary and tertiary sites and the formation of side products which is caused by carbene dimerization is not a major problem with rhodium carbenoids derived from aryldiazoacetates.







# **1.2 Arene C-H Bond Activation**

#### **1.2.1 Chelate-Assisted Arene C-H bond Activation Reactions**

Murai pioneered chelate-assisted, regioselective, catalytic C-H bond activation of arene and olefin coupling reaction.<sup>3</sup> For example, acetophenone is added to vinyl silane to yield regioselectively alkylated product at the *ortho* position (eq 6). The ruthenium complex  $RuH_2(CO)(PPh_3)_3$  (8) was found to be an efficient catalyst for this coupling reaction. This reaction is a quite effective synthetic method, and, in many cases, gave nearly quantitative yields.

As shown Scheme 4, Murai proposed that the reaction involves the coordination of the carbonyl group of arylketones to the ruthenium center and is placed in a favorable position for the cleavage of the C-H bond. The ruthenium hydride complex **9** has been suggested as a key intermediate species. The rate-limiting step of the coupling reaction is the C-C bond forming step instead of C-H activation step.





The reaction was extended to various arene and olefin coupling reactions. One of the most important findings in elucidation of the mechanism of coupling reaction is that the C-H bond cleavage step is not the rate-limiting step. Instead a rapid equilibrium exists prior to the reductive elimination step that the leads to the C-C bond formation. Murai has established that the reductive elimination of alkylated products is the rate-determining step.

The rhodium bis-olefin complex  $[Cp*Rh(CH_2=CHSiMe_3)_2]$  (10) has also been shown to be an effective catalyst for selective addition of olefins to the *ortho* position of arylketones (eq 7).<sup>8</sup> A main difference between eq 1 and eq 3 is that in the rutheniumcatalyzed process, a carbonyl coordination is presumed to be the first step, which directs the C-H bond activation to the *ortho* position of the aromatic substrate.

$$\begin{array}{c} O \\ + \end{array} \\ Si(OEt)_3 \\ \hline C_6H_{12}, 120 \\ \hline C_6H_{12}, 120 \\ \hline C_6W_{12}, 120 \\ \hline C_{6}W_{12}, 120 \\ \hline C_{6$$

H/D exchange experiments establish that *para* and *meta* C-H bonds are not activated in the ruthenium system. The rhodium catalytic processes, on the other hand, are not discriminating in the C-H bond activation step and activation of all sites (*ortho, meta,* and *para*) of the substrate is observed. Murai has established that the C-H bond activation is fast and reversible in the ruthenium case and reductive elimination of the alkylated product is the rate-limiting step. It is believed that the similar features are applied to the rhodium system as well, but no conclusive evidence is available concerning this point. In the both systems, the energy barrier for reductive elimination which then forms the product is decreased by chelation of the carbonyl group to the metal center.

## Scheme 5



The *ortho* position of imine substituted aromatic compounds is directly arylated and alkenylated with organic halides in the presence of catalytic amount of a ruthenium-phosphine complex. In this reaction, the imine group acts as a directing group for *ortho*-metallation step (eq 8).<sup>23</sup>

$$\begin{array}{c} N^{Ph} & Br \\ + & K_2CO_3, NMP, 100 \ C \end{array} \begin{array}{c} Ph & N^{Ph} & Ph & N^{Ph} \\ \hline & & & & \\ 85 : 15 \ (78 \ \%) \end{array} \end{array}$$

Aromatic aldimines have been found to be a suitable substrate for coupling reaction with either vinylsilanes or styrenes in the presence of ruthenium(II) complex and phosphine ligand via C-H bond activation at the *ortho* position (eq 9).<sup>24</sup>

$$\begin{array}{c} \mathsf{N}t\text{-}\mathsf{Bu} \\ + & \mathsf{Si}(\mathsf{OMe})_3 \end{array} \xrightarrow[\mathsf{RuCl}_2(p\text{-}\mathsf{cymene})]_2 \\ \hline \mathsf{NaHCO}_2 \text{ cat., }\mathsf{PPh}_3 \\ \text{toluene, 140 } \mathfrak{C} \end{array} \xrightarrow[\mathsf{N}t\text{-}\mathsf{Bu} \\ \end{array}$$
(9)

Recently, Glorius has reported the selective rhodium catalyzed oxidative olefination of aromatic carbonyl compounds such as acetanilide, acetophenone and benzamide derivartives with olefins involving *ortho* C-H activation.<sup>25</sup> The C-H bond activation of acetanilide and subsequent coupling to styrene was found to proceed smoothly at 0.2 M in tert-amyl alcohol as solvent employing only 0.5 mol% of  $[RhCp*Cl_2]_2$  (**11**) catalyst precursor (eq 10).

Glorius has also published the oxidative olefination of acetophenones and electronically related benzamides with various olefins. Acetophenone and benzamide coupled with styrene selectively to give a single product each in good yield (eq 11).<sup>26</sup> He also proposed that the oxidative olefination reaction proceeds through chelate-assisted *ortho* C-H activation, followed by olefin insertion to rhodium center and subsequent  $\beta$ -hydride elimination to selectively yield the *E* selective coupling product. (Scheme 6).







Yu has developed a new dihydrobenzofuran ring-forming process via hydroxydirected C-H activation/C-O bond formation (eq 12).<sup>27</sup> Tertiary alcohols were cyclized to give the corresponding dihydrobenzofurans in good yield in the precence of Pd(OAc)<sub>2</sub> with PhI(OAc)<sub>2</sub> as an oxidant. This reaction provides a new method for constructing dihydrobenzofurans, including spirocyclic analogues, a process that is potentially applicable to natural product synthesis.



Sanford has developed a selective catalytic coupling method for the oxidative functionalization of C-H bonds.<sup>28</sup> The combination of 1 equiv of benzo[h]quinolone with PhI(OAc)<sub>2</sub> (2 equiv) and 2 mol% Pd(OAc)<sub>2</sub> in CH<sub>3</sub>CN at 75 °C for 12 h produces an 1:1 ratio of the monoacetoxylated product (X = OAc) and the analogus phenol (X = OH) in 86% isolated yield (eq 13). They has also found that simple modification of the reaction condition allows the selective installation of a variety of different functional groups at C<sub>10</sub>. For example, reaction of benzo[h]quinolone with PhI(OAc)<sub>2</sub>/catalytic Pd(II) in alcohol solvents produces a range of sterically and electronically diverse alkyl-aryl ethers (X = OMe, OEt, O*i*-Pr, OCH<sub>2</sub>CF<sub>3</sub>) in good yield. Alternatively, when oxidation is carried out in the presence of excess LiX (X = Cl, Br) in CH<sub>3</sub>CN, traces of mono-halogenated products are formed.



#### **1.2.2 Nonchelate-Assisted Reactions**

Fujiwara reported the effective metalation of aromatic C-H bonds at room temperature using highly-electrophilic, *in situ* generated, Pd(II) and Pt(II) cationic species

in trifluoroacetic acid (TFA), leading to the regioselective inter- and intramolecular addition of simple arenes to C-C multiple bonds (eqs 14 and 15).<sup>29</sup>

$$\begin{array}{c|c} & + & H & - = -R & - \frac{Pd(OAc)_2}{TFA, RT} & & & H \\ \hline & & & & & \\ \end{array}$$
(14)

In most cases, the addition of alkynes exclusively affords the thermodynamically less stable *cis*-aryl alkene. The intramolecular hydroarylation of C=C bonds is facile and regiospecific because of the electrophilic metalation of aromatic C-H bonds by the Pd(II) cationic species which is assisted by ethynyl coordination. In fact, this intramolecular reaction combines both chelation assistance and electrophilic metalation.

Milstein's Heck-type oxidative coupling reaction, which was found to be promoted by molecular oxygen, is among the most remarkable examples of ruthenium-catalyzed arene C-H bond insertion reactions (eqs 16 and 17).<sup>30</sup> The catalytic oxidative coupling of arenes with alkenes to generate aryl alkenes, in which the double bond is preserved, is a highly desirable goal. Since the reaction does not require the utilization of reactive substituent and does not form any byproducts, it has a unique advantage over other methods for the preparation of aromatic alkenes, especially when compare to the well-known Heck reaction for the vinylation of aryl halides.



In Milstein's Heck type coupling, dioxygen can be directly used as the oxidant, while still retaining good catalytic activity. The reaction is catalyzed by ruthenium complexes under a CO atmosphere condition. Readily available ruthenium complexes such as RuCl<sub>3</sub>'3H<sub>2</sub>O, [Ru(CO)<sub>3</sub>Cl<sub>2</sub>]<sub>2</sub>, [( $\eta^6$ -C<sub>6</sub>H<sub>6</sub>)RuCl<sub>2</sub>]<sub>2</sub> Ru(NO)Cl<sub>3</sub>'5H<sub>2</sub>O, and Ru(F<sub>3</sub>CCOCHCOCF<sub>3</sub>)<sub>3</sub> show essentially the same catalytic activity.

Smith has described that iridium catalysts (MesH)Ir(BPin)<sub>3</sub> (MesH =  $\eta^6$ mesitylene, BPin = oxaborolane) (12) are efficient for direct synthesis of arylboron coumpounds from aromatic hydrocarbons and boranes under "solventless" conditions (eq 18).<sup>31</sup> The Ir catalysts are highly selective for C-H bond activation and do not interfere with subsequent transformations, including Pd-mediated cross-coupling with aryl halides. By virtue of their favorable activities and exceptional selectivities, these Ir catalysts impart the synthetic versatility of arylboron reagents to C-H bonds both aromatic and heteroaromatic hydrocarbons.

$$H \to H \to B(OR)_2$$
   
 $150 \, \text{°C}, \, 15 \, \text{h}$   $H \to B(OR)_2$  (18)





Smith proposed a reaction mechanism involving Ir(III) and Ir(V) intermediates as shown in Scheme 7. The following evidence supports the mechanism proposed in Scheme 7: (a) Borylation products of iodobenzene are not obtained when Ir(I) sources are used under stoichiometric or catalytic conditions, whereas Ir(III) complexes effect both stoichiometric and catalytic borylations. (b) Improved catalytic activity is observed with chelating phosphines and inhibition is observed when [phosphine]:[iridium] ratios equal or exceed 3:1, strongly supporting the viability of bisphosphine intermediates (Scheme 4, n = 2), which could be generated via E-E reductive elimination (E = H, BPin) from an 18electron bisphosphine Ir(V) resting state. (c) The 18 electron bisphosphine compound, IrH<sub>5</sub>(PMe<sub>3</sub>)<sub>2</sub>, is an efficient precatalyst for the borylation.

+ HSiEt<sub>3</sub> 
$$\frac{\text{Tp}^{\text{Me2}}\text{Pt}(\text{Me})_2(\text{H})}{200 \, \text{C}, 24 \, \text{h}}$$
  $\text{SiEt}_3 + \text{H}_2$  (19)

Hartwig discovered a highly selective, acceptorless dehydrogenative coupling of silanes with arene C-H bonds in good yields in the presence of  $Tp^{Me2}PtMe_2H$  ( $Tp^{Me2} =$  hydridotris(3,5-dimethylpyrazolyl)borate) and related platinum(IV) complexes (eq 19).<sup>32</sup> The reaction of the arenes occurred selectively at the least sterically hindered C-H bonds and preferentially with more electron-poor arenes. If the dehydrogenative coupling of simple silanes with aromatic and aliphatic C-H bonds could be developed, then useful reactions could result. Because of the higher stability of silanes toward disproportionation than boranes, they could undergo tandem or sequential reactions that would be impossible using borane or even disilane reagents.

#### 1.3 Chelate-Assisted C-H Bond Activation Reaction of Alkenes

Trost and Murai independently reported the ruthenium catalyzed C-H activation and alkylation of 1-acetylcyclohexene and  $\alpha$ , $\beta$ -unsaturated esters with alkenes such as silylethylenes and styrene.<sup>4(b),(e)</sup> The reaction of 1-acetylcyclohexene and *N*methylcinnamide with triethoxyvinylsilane in presence of Ru(H)<sub>2</sub>(CO)(PPh<sub>3</sub>)<sub>3</sub> (**8**) gave the coupling product involving vinyl C-H activation (eqs 20 and 21). They proposed the plausible reaction mechanism for olefin-olefin coupling reaction as shown in Scheme 8. The chelation-assisted insertion of ruthenium into the C-H bond by vinyl C-H activation gives **13**. Hydrometallation of alkene would result in the formation of **14**. Reductive elimination from **14** provides coupling product, regenerating the catalytically active species.



Scheme 8



Jun has developed a rhodium(I) catalyzed  $\beta$ -alkylation of the  $\alpha$ , $\beta$ -unsaturated ketone with 1-alkenes employing diethylamine as a efficient chelation-assistant tool.<sup>33</sup> The  $\beta$ -alkylation of 4-phenyl-3-butene-2-one with 1-hexene in presence of RhCl(PPh<sub>3</sub>)<sub>3</sub>

(15), benzoic acid and diethylamine was found to proceed smoothly at 130 °C for 24 h in good yield (eq 22).

He also proposed the plausible mechanism of this reaction which is illustrated in Scheme 9. Precoordination of a rhodium species to the nitrogen functionality of the dienamide, derived from condensation of the enone with dialkylamine, can facilitate the cleavage of the vinyl C-H bond to generate the stable 5-membered metallacyclic rhodium hydride **16**. Coordination of an olefin followed by migratory insertion of an olefin into the Rh-H bond gives alkylrhodium species **17**. Reductive elimination in **17** furnishes the  $\beta$ -alkylated dienamine compound that equilibrates with regioisomer and regenerates a rhodium catalyst to complete the catalytic cycle. Following acidic hydrolysis of both dienamine regioisomers leads to the  $\beta$ -alkylated products.



## Scheme 9



Ellman and Bergman have reported the stereoselective alkylation of  $\alpha$ , $\beta$ unsaturated imines via the directed activation of a C-H bond by rhodium catalyst.<sup>34</sup> The rhodium complex which is [RhCl(COE)<sub>2</sub>]<sub>2</sub> has been found to be effective catalyst with phosphine ligands for coupling reaction with  $\alpha$ , $\beta$ -unsaturated imines and a variety of alkenes (eq 23).



Imine groups are synthetically useful because they can be easily converted to many other functional groups such as ketones, alcohols, carboxylic acids, and amines and the imine is sometimes utilized as a directing group in transition metal catalyzed direct funtionalization of aryl or aliphatic compounds using alkenes and alkynes.

## **1.4 C-H Bond Activation Reactions of Nitrogen Containing Compounds**

The catalytic reactions and functionalizations which involve cleavage of sp<sup>2</sup> or sp<sup>3</sup> C-H bonds within nitrogen containing compounds, such as amines and amides have attracted considerable attention, in part due to their prominent role in the synthetic utilities of natural products and pharmaceutical agents. Highly regioselective catalytic reactions via C-H activation of nitrogen-containing compounds, especially heterocyclic compounds have been reported by several research groups.<sup>35-47</sup> However, coupling reactions of amide substrates by regioselective C-H activation with alkenes are still very rare.<sup>16, 48-51, 53</sup>

## **1.4.1 C-H Bond Activation of Pyrrolidine Derivatives**

Sames and co-workers have reported the oxidative intramolecular cyclization reaction of alkene-amide substrates under neutral and catalytic conditions. This overall transformation requires tandem sp<sup>3</sup> C-H bond activation at the position adjacent to the amide nitrogen and C-C bond formation. For example, pyrrolidine substrate was converted to pyrrolizidinone and indolizidinone products in 66% and 17% yield, respectively, in presence of  $[Ir(COE)_2Cl]_2$  with carbene ligand IPr (*N*,*N*'-bis(2,6-diisopropylphenyl)imidazolyl carbene), and the hydrogen acceptor NBE or TBE (NBE = norbornene) (eq 24).<sup>35</sup>



They proposed a reaction mechanism involving sp<sup>3</sup> C-H activation and C-C bond formation as shown in Scheme 10. The results presented in this study suggest the complex [IPrIr(Cl)(substrate)] (**18**) is key intermediate which is generated by replacing COE ligands at the iridium center by a carbene ligand and the substrate. The second step involves rearrangement via C-H activation adjacent amide nitrogen, providing an alkyl iridium hydride, and then undergoes new C-C bond formation by alkene insertion. In the fourth step, dihydride-alkene complex was produced by  $\beta$ -hydride elimination (reductive elimination would give the saturated product). To regenerate [IPrIr(Cl)(substrate)], two hydrides were added to a hydrogen acceptor, followed by release of the product.





They also describe the direct arylation of  $sp^3$  C-H bonds through the intermediacy of a metal-alkoxide, formed via ketone insertion, which could undergo transmetalation

with an arylboronate ester.<sup>36</sup> Importantly,  $Ru_3(CO)_{12}$  as the catalyst gave the desired product in good yield. The amidine substrates and phenylboronate ester in the presence of ruthenium catalyst and ketone produced a 3:1 ratio of *trans* and *cis* diastereomers (eq 25). The trinuclear ruthenium cluster is required for  $\alpha$ -arylation of amidine derivatives because mononuclear and dinuclear ruthenium complexes were found to be much less effective (<10%).



Murai reported catalytic reactions that involve the cleavage of the sp<sup>3</sup> C-H bond adjacent to the nitrogen atom in *N*-2-pyridinyl alkylamines. The use of  $Ru_3(CO)_{12}$  as the catalyst resulted in the insertion of alkenes into sp<sup>3</sup> C-H bonds of the pyrrolidine to give the coupling products (eq 26).<sup>37</sup>



A variety of terminal, internal, and cyclic alkenes were found to be suitable substrates for the coupling reaction. The presence of a pyridine ring as a directing group on the nitrogen in cyclic amines is necessary for the reaction to proceed, suggesting the importance of the coordination of the pyridine nitrogen to ruthenium.

Yi has reported that the complex  $(PCy_3)_2(CO)RuHCl$  (**19**) is an effective catalyst for dehydrogenative coupling reactions of cyclic amines and alkenes. The coupling reaction of secondary cyclic amine with unactivated alkenes, especially terminal alkenes such as ethylene, propylene, and TBE, preferentially afforded C-H bond insertion products which were selectively activated by sp<sup>3</sup> C-H bond adjacent nitrogen in cyclic amines (eq 27a). In contrast, the reaction of amines with vinylsilane gave the *N*-silylation products (eq 27b).<sup>38</sup>

### 1.4.2 C-H Bond Activation of Indole and Quinoline derivatives

Widenhoefer has reported that the intramolecular alkylation of 2-alkenyl indoles with a catalytic amount of  $PtCl_2(CH_2=CH_2)$  in dioxane that contained a trace of HCl at 60 °C for 24 h led to tetrahydrocarbazoles as cyclization products (eq 28).<sup>39</sup> Platiumcatalyzed cyclization of 2-(4-pentenyl)indoles showed good generality and tolerated
substitution at each position of the 4-pentenyl chain including the internal and *cis*- and *trans*-terminal position. Also, this synthetic method was applicable to the synthesis of tetrahydro- $\beta$ -carbolinones and was effective for cyclization of unprotected indoles. 2-(3-Butenyl)indoles were carried out platinum-catalyzed cyclization with 6-*endo*-trig regioselectivity. Mechanistic studies established a mechanism for the platinum-catalyzed cyclization of 2-alkenyl indoles involving a nucleophilic attack of the indole on a platinum-complexed olefin.

$$(28)$$

In contrast, the reaction of 1-methyl-2-(4-pentenyl)indole with a catalytic amount of PdCl<sub>2</sub>(CH<sub>3</sub>CN)<sub>2</sub>, and a stoichiometric amount of CuCl<sub>2</sub> in methanol under CO atmosphere at room temperature for 0.5 h, led to the cyclization and carboalkoxylation to form the corresponding tetrahydrocarbazole as a single regioisomer in 83% yield (eq 29).<sup>40</sup> Palladium-catalyzed cyclization/carboalkoxylation of 2-(4-pentenyl)indoles tolerated substitution along the alkenyl chain and at the internal and cis-terminal positions. А variety of alcohols were also tolerated for palladium-catalyzed cyclization/carboalkoxylation. This cyclization/carboxylation catalyzed by palladium complex was also effective for cyclization of 2-(3-butenyl)indoles, 3-(3-butenyl)indoles, 3-(4-pentenyl)indoles, and 2-(5-hexenyl)indoles.

$$R = Me, Et, n-Octyl, i-Pr, Cy$$

$$\begin{array}{c}
PtCl_2(CH_3CN)_2 \\
ROH, CuCl_2 \\
THF, 25 \ \ C, \ 0.5 \ h \\
CO
\end{array}$$

$$73 \sim 83\% \quad O$$

$$(29)$$

Bergman and Ellman have reported the first intermolecular coupling of unactivated alkenes to heterocycles by using Rh(I) catalyst (eq 30).<sup>16(b)</sup> A wide range of heterocycles were employed in the reaction, and a variety of functional groups could be incorporated, including esters, nitriles, and acetals. The intermolecular coupling were found to be promoted by weak Lewis acids such as MgBr<sub>2</sub>, which dramatically increased the reaction rate of both the inter- and intramolecular reactions. Different functional groups can be introduced from readily available starting materials, leading to the potential use of this reaction in drug discovery and process chemistry.

Aryl-rhodium(III) complexes generated *in situ* from the reaction of  $[RhCl(COE)_2]_2$  (COE = *cis*-cyclooctene),  $[p-(CF_3)C_6H_4]_3P$ , and CsOPiv efficiently catalyzed the direct C-arylation of free (NH)-indoles and (NH)-pyrroles in presence of mild base in good yields and with high regioselectivity (eq 31).<sup>41</sup> The reaction displayed excellent functional group compatibility and low moisture sensitivity. Kinetics studies

support a mechanism involving phosphine displacement by an indole in Ar-Rh(III) pivalate complexes (resting state of catalyst), followed by a rate-limiting C-H bond metalation.

 $L = [p-(CF_3)C_6H_4]_3P$ 

An Ar-Rh(III) pivalate complex is assembled in situ and represents the resting state of the catalyst. Subsequently, displacement of the phosphine ligand by indole takes place in a pre-equilibrium step to form indole-Rh(III) complex, which is followed by the slow C-H bond metalation step as shown Scheme 11. The resulting intermediate **20** then undergoes reductive elimination, furnishing the desired coupling product. The rhodium(I) complex formed in this step is rapidly converted back to the resting state via an oxidative addition of iodobenzene and a halide-pivalate exchange. Thus, the oxidative addition of haloarene precedes the slow C-H transformation step. The model is further supported by the large kinetic isotope effect ( $k_{\rm H}/k_{\rm D} = 3.0$ ) at the 2-position of indole. Although the intimate mechanistic details of the C-H metalation step are unclear, they propose that the pivalate ligand assists the C-H bond dissociation as an internal base. This is consistent with the fact that the initial rates of the reaction between indole and Ar-Rh(III) pivalate complex were not affected by the addition of CsOPiv.

# Scheme 11



As shown in eq 32, Li has developed a new type efficient C-C bond formation method via a cross-dehydrogenative coupling (CDC) reaction of indoles and tetrahydroquinolines catalyzed by copper bromide in the presence of an oxidizing reagent, like *t*-BuOOH (eq 32).<sup>42</sup> The CDC reaction provides a simple and efficient catalytic pathway to construct indolyl tetrahydroisoquinolines via combination of sp<sup>3</sup> C-H and aryl sp<sup>2</sup> C-H bond activation followed by C-C bond formation.



#### 1.4.3 C-H Bond Activation of Pyridine derivatives

Moore reported that  $Ru_3(CO)_{12}$  was found to be a selectively active catalyst for the *ortho* positions in pyridine (eq 33).<sup>43</sup> For instance, pyridine is used as solvent and the reaction is conducted at 150 °C under 10 atm of carbon monoxide. Conversion of 1hexene to pyridinyl ketone mixture results in the high regioselectivity of the reaction. In addition, no other pyridine containing products are observed.

The cationic ruthenium vinylidene complex  $[CpRu(=C=CHR)(PPh_3)_2]PF_6$  (21) has been reported to be an effective catalyst for the direct alkenylation reaction of pyridines (eq 34).<sup>44</sup> Heating the cationic ruthenium vinylidene complex in pyridine at 100-125 °C for 24 h gives (*E*)-2-akenylpyridine. Initially, pyridine coordinates to ruthenium center by displacement of one of the phosphine ligands. Then [2+2] heterocycloaddition occurs to form a 4-membered ruthenacyclic complex. Deprotonation of the  $\beta$ -hydrogen affords a neutral  $\pi$ -azaallyl complex. Protonolysis produces the product. As a result, a vinylidene group is inserted into the  $\alpha$ -C-H bond of pyridine. The alkenylation reaction is made catalytic in ruthenium by the use of (alkyn-1-yl)silane as the vinylidene source. Treatment of (alkyn-1yl)trimethylsilane with pyridine in the presence of cationic ruthenium complex affords the corresponding (E)-2-alkenylpyridine not only in a good yield but also in a regio- and stereoselective manner.



Recently, Kakiuchi has reported a new method for producing  $\pi$ -conjugated aromatic compounds by the ruthenium catalyzed alkenylation of an aromatic C-H bond in arylpyridines and related compounds with alkenyl esters (eq 35).<sup>45</sup> This reaction provides a new halogen-free reaction condition that can be used in the synthesis of  $\pi$ -conjugated aromatic compounds. For example, the reaction of 2-(2-tolyl)pyridine with styryl acetate in presence of Ru(COD)(COT) (COD =1,5-cyclooctadiene, COT = 1,3,5-cyclooctatriene) as a catalyst, provides the *E*-isomer as the sole product in 98% yield.



Chang has reported two new types for the oxidative C-C bond formation of pyridine *N*-oxides; selective alkenylation and direct cross coupling reaction with unactivated arenes (eq 36).<sup>46</sup> For selective alkenylation, reaction proceeds highly efficiently when of  $Pd(OAc)_2$  was used in combination with  $Ag_2CO_3$  (1.5 equiv.) and pyridine (1.0 equiv.) with pyridine *N*-oxides and ethyl acrylate affords highly site selective coupling product in good yield.

Chang also discovered the direct arylation of pyridine *N*-oxides with unactivated arenes catalyzed by palladium complex. For instance, reaction of pyridine *N*-oxide or its *para* derivative with benzene as unactivated arene afforded the corresponding *ortho* phenylated products with 3:1 ratio in high yields (eq 37).

$$\begin{array}{c} R \\ \hline \oplus N \\ \ominus O \end{array} + \end{array} \begin{array}{c} Pd(OAc)_{2} \\ Ag_{2}CO_{3} \\ \hline 1,4-dioxane, 130 \ \ C, 16 \ h \end{array} \begin{array}{c} R \\ \oplus N \\ \ominus O \end{array} \begin{array}{c} Ph \\ Ph \\ \Theta O \\ 3:1 \end{array} \begin{array}{c} R \\ \ominus O \\ \end{array} \begin{array}{c} R \\ \oplus O \\ \end{array} \end{array} \begin{array}{c} R \\ \oplus O \\ \end{array} \end{array}$$

A cobalt-catalyzed, chelate-assisted hydroarylation reactions of styrenes, described by Yoshikai demonstrated that the chelate-assisted C-H bond activation at

*ortho* position of phenylpyridine derivatives allowed for tuning of the branched/linear regioselectivity by ligand control (eq 38).<sup>47</sup> The reaction of 2-phenylpyridine with styrene in presence of CoBr<sub>2</sub>, PCy<sub>3</sub>, and Me<sub>3</sub>SiCH<sub>2</sub>MgCl at 60 °C in THF for 12 h gave a branched product in 88% yield, with high regioselectivity (b/l = 96:4). In contrast, when the reaction of 2-phenylpyridine with styrene was performed using IMes·HCl (1,3-dimesitylimidazolium chloride), a precursor of an *N*-heterocyclic carbene (NHC) ligand, instead of PCy<sub>3</sub> a near complete switch of the reioselectivity was observed. Thus the reaction afforded the linear product in 84% yield with a regioselectivity of 97:3.



#### 1.4.4 C-H Bond Activation of Amide, Amine, and Imine Derivatives

Recently, Kondo has reported the novel ruthenium-catalyzed regio- and stereoselective codimerization of *N*-vinylamides with alkenes or alkynes. Treatment of *N*-methyl-*N*-vinylacetamide with ethyl acrylate in the presence of a catalytic amount of  $[Ru(\eta^6-COT)(\eta^2-DMFM)_2]$  (COT = 1,3,5-cyclooctatriene, DMFM = dimethyl fumarate) (22) in *N*,*N*-dimethylacetamide (DMA) at 160 °C for 3 h gave the linear codimer, ethyl-5-(*N*-methylacetylamino)-pent-4-enoate, in 90% yield with 95% *E* selectivity (eq 39).<sup>48</sup>

$$\overset{O}{\longrightarrow}_{N} \overset{V}{\longrightarrow} + \overset{CO_{2}Et}{\longrightarrow} \overset{22}{\xrightarrow{DMA, 160 \ C, 3h}} \overset{O}{\longrightarrow}_{N} \overset{CO_{2}Et}{\xrightarrow{N}} \overset{(39)}{\xrightarrow{72\% (95\% \ E \ form)}}$$

To learn more about this reaction mechanism, they performed a deuterium labeling study. The reaction of *N*-methyl-*N*-vinylacetamide with dimethyl fumarate- $d_2$  was carried out in the presence of [Ru( $\eta^6$ -COT)( $\eta^2$ -DMFM)<sub>2</sub>] catalyst in DMA at 160 °C for 3 h to give the deuterium incorporated product (eq 40). The results of deuterium labeling studies could be explained by the formation of a ruthenium hydride species at an early stage in the catalytic cycle.

-

They proposed a reaction mechanism for the codimerization of *N*-vinylamides with alkene (Scheme 12). The reaction starts with the removal of a COT ligand in  $[Ru(\eta^6-COT)(\eta^2-DMFM)_2]$  by *N*-vinylamide through metallacycle formation of **23** and reductive elimination to generate coordinatively unsaturated zero-valent ruthenium hydride species through activation of sp<sup>2</sup> C-H bonds in alkenes or a DMFM ligand. Alkenes are inserted into a Ru-H bond, followed by the successive chelate-assisted insertion of *N*-vinylamides

into a Ru-C bond. Subsequent  $\beta$ -hydride elimination gives the products with regeneration of an active ruthenium hydride species.



Scheme 12

Chatani has discovered the ruthenium-catalyzed cyclocarbonylation of aromatic amides leading to phthalimides through C-H bond activation. The reaction of N-(2pyridinylmethyl)benzamide with CO and ethylene as a H<sub>2</sub> acceptor and H<sub>2</sub>O in presence of Ru<sub>3</sub>(CO)<sub>12</sub> in toluene at 160 °C did not give carbonylation product; instead, the

phthalimide was formed in 77% yield (eq 41). A wide variety of functional groups at the *para* position, including methoxy, amino, ester, ketone, cyano, chloro, and even bromo substituents, were tolerated in this cyclocarbonylation.<sup>49</sup>



The catalytic reactions which involve the cleavage of sp<sup>3</sup> C-H bonds are much more difficult than that of sp<sup>2</sup> C-H bonds. It is known that the cleavage of sp<sup>3</sup> C-H bond is kinetically and thermodynamically unfavorable. Recently, Yu has reported Pd(II) catalyzed reaction for the direct olefination of sp<sup>3</sup> C-H bonds. After  $\beta$ -C-H olefination, the amide products underwent 1,4-conjugate addition to give the corresponding lactam compounds (eq 42).<sup>15(a)</sup> Electron-withdrawing substitutents on the *N*-aryl group dramatically increase the reaction rate. Especially, the most electron-withdrawing CONHAr<sub>2</sub> group improved the yield further to 88%. The choice of solvent was also critical for this reaction, with polar and strongly coordinating amide solvents such as DMF, DMA and NMP giving the best reactivity.



Shibata has also found that a cationic Ir(I)-BINAP complex showed high catalytic activity in sp<sup>3</sup> C-H bond alkenylation of amide with alkynes through carbonyl-directed C-H bond activation, which prevailed over aromatic sp<sup>2</sup> C-H bond activation. Alkenylation of amides with alkynes via carbonyl directed sp<sup>3</sup> C-H bond activation was proceeded with Ir(I)-BINAP.<sup>50</sup> For example, the reaction of *N*,*N*-dimethylbenzamide with diphenylacetylene in the presence of in situ formed cationic Ir(I)-BINAP complex gave the allylamide product as an E-isomer in good yield (eq 43).

Tanaka reported that cationic rhodium(I)/BIPHEP complex catalyzes 1pyrrolidinecarbonyl-directed regioselective alkenylations of olefinic or aromatic sp<sup>2</sup> C-H bonds in good yields under mild reaction conditions.<sup>51</sup> The reaction of crotonamide with diphenylacetylene catalyzed by  $[Rh(COD)_2]BF_4/BIPHEP$  afforded stereoselective coupling products in good yield (eq 44). A 1-pyrrolidinecarbonyl group as a directing group is essential for this transformation. Dimethylcarbamoyl and 1-piperidinecarbonyl groups significantly decreased the reaction rate, and ester and ketone carbonyl groups completely shut down the reaction. The reaction between benzamide and alkynes also proceeded in presence of the same rhodium catalyst (eq 45). Indeed, the desired alkenylation proceeded at ambient temperature, and the 1-pyrrolidinecarbonyl group is also the best directing gourp.



Recently, Ellman and Bergman have reported the oxidative coupling of aryl *O*-methyl oximes with unactivated alkenes via C-H bond functionalization catalyzed by Rh(III) complex (eq 46).<sup>52</sup> A wide range of unactivated alkenes were employed in the reaction, and a variety of functional groups could be in corporated, including phenyl, ester, and halogen groups. The oxidative coupling of aryl *O*-methyl oximes with alkenes

using  $[Cp*RhCl_2]_2$  as the catalyst in the presence of  $Cu(OAc)_2$  as an oxidant and with THF as the solvent gave their corresponding oxidative products.

$$MeO_{N} = \begin{bmatrix} Cp^{*}RhCl_{2}]_{2} & MeO_{N} \\ Cu(OAc)_{2} & \\ AgSBF_{6} \\ \hline THF, 75 \ C, 20 \ h \end{bmatrix}$$

$$(46)$$

R = n-Bu, *i*-Pr, Cy, Ph, CO<sub>2</sub>Et, OAc

Li and coworkers has developed the C-H olefination of such bezamides at the *C*-aryl ring. When electron-poor olefins are utilized, the coupled products may futher undergo in situ Michael addition to give  $\gamma$ -lactams (eq 47).<sup>53</sup> This reaction would proceed by the oxidative olefination of the *ortho* C-H bond in the C-aryl ring, followed by intramolecular hydroamination. Benzamides with electron-donating or –withdrawing groups affords lactams in high isolated yield when electron-poor olefins are employed.



Recently, Glorius has reported that the challenging allylic sp<sup>3</sup> C-H activation and also an alternative vinylic sp<sup>2</sup> C-H activation of enamines and the subsequent coupling with unreactivated alkynes yielding pyrroles.<sup>54</sup> The reaction of *N*-acetyl enamine

derivatives with internal alkyens in precence of  $[Cp*RuCl_2]_2$  and AgSbF<sub>6</sub> as the catalyst together with Cu(OAc)<sub>2</sub> as the oxidant in 1,2-dichloroethane resulted in the formation of pyrroles (**24**) as a single regiosiomer (eq 48). On the other hand, the change from ester to a nitrile resulted in the  $\alpha$ -functionalization of the enamine and, consequently, the formation of a regoisomeric pyrrole (**25**). This important results indicate the crucial role of the ester group to activate the allylic sp<sup>3</sup> C-H bond.



# **Chapter II**

# Synthetic and Mechanistic Studies of Chelate-Assisted Oxidative Coupling Reaction of Arylamides and Unactivated Alkenes

Chelate-assisted catalytic coupling reactions involving C-H activation have emerged as one of the most powerful C-C bond formation methods for arene compounds.<sup>1</sup> Among the chelate-directed catalytic C-H activation and functionalization reactions, oxidative coupling methods have been found to be particularly effective in forming new C-C and carbon-to-heteroatom bonds to sp<sup>2</sup> arene C-H bonds.<sup>1d,e</sup> Since Fujiwara's seminal reports on the oxidative coupling reactions of arene compounds,<sup>55</sup> oxidative C-N, C-O and C-halogen bond-forming reactions of arene compounds have been achieved by using chelate directing groups.<sup>56</sup> Fagnou reported regioselective cross coupling reactions of unactivated arene compounds by using amino and amide directing groups.<sup>57</sup> Late transition metal catalysts have been found to be most versatile in mediating Heck-type C-H oxidative alkenylation of heteroarene compounds, where the regioselectivity has often been found to be dictated by both steric and electronic nature of the arene substituents and chelate directing groups.<sup>58</sup> By screening amino acid ligands, Yu recently developed a remarkably selective Pd-catalyzed C-H olefination of carboxysubstituted arene compounds.<sup>59</sup> Still, most of these oxidative coupling methods either require stoichiometric amounts of metal oxidants or reactive reagents. Thus development of direct oxidative C-H coupling methods which do not require strong oxidizing agents or reactive substrates with increased the efficiency of the coupling reaction would be beneficial from both synthetic and, environmental points of view.

# **Results and Discussion**

The regioselective coupling reactions of nitrogen containing compounds catalyzed by late transition metal catalysts have been developed by several research groups, as well as  $sp^2$  and  $sp^3$  C-H bond activation of nitrogen heterocycles and amide derivatives.<sup>15, 48-51,</sup> <sup>53</sup> The selective C-H bond activation of amide compounds is a highly attractive synthetic tool because organic compounds which contain amide functional groups are very useful materials in natural products and pharmaceutical agents. Our research group has reported that the *in-situ* formed cationic ruthenium-hydride complex is a highly effective catalyst for the coupling reaction of arylketones and linear alkenes involving C-H bond activation.<sup>60(a)</sup> Recently, our research group also developed a convenient and efficient method synthesize cationic ruthenium-hydride to а complex  $[(\eta_{6} C_6H_6$ )(PCy<sub>3</sub>)(CO)RuH]<sup>+</sup>BF<sup>-</sup><sub>4</sub> (26) from the protonation reaction of tetrameric ruthenium complex {[(PCy<sub>3</sub>)(CO)RuH]<sub>4</sub>( $\mu$ -O)( $\mu$ -OH)<sub>2</sub>} (4) which was synthesized in two steps from ruthenium-hydride complex (PCy<sub>3</sub>)<sub>2</sub>(CO)RuHCl (19) (Scheme 13).<sup>60(b), 60(c)</sup> Thus, the treatment of tetrameric ruthenium complex 4 (200 mg, 0.12 mmol) with HBF<sub>4</sub>·OEt<sub>2</sub> (64  $\mu$ L) in C<sub>6</sub>H<sub>6</sub> at room temperature cleanly afforded the cationic ruthenium-hydride complex 26, which was isolated as an ivory-colored solid in 95% yield (Scheme 13). The ruthenium-hydride signal was observed at  $\delta$  -10.39 (d,  $J_{\rm PH}$  = 25.9 Hz) by <sup>1</sup>H NMR spectroscopy of 26 in CD<sub>2</sub>Cl<sub>2</sub>, and a single phosphine signal was detected at  $\delta$  72.9 ppm by  ${}^{31}P{}^{1}H$  NMR spectroscopy. The molecular structure of cationic ruthenium-hydride 26, characterized by X-ray crystallography, showed a three-legged piano-stool geometry, which is capped by an  $\eta^6$ -benzene moiety (Figure 1).<sup>48</sup> The Ru<sub>1</sub>-P<sub>1</sub> distance for complex

**26** is 2.32 Å and the average bond length of  $Ru_1$ - $C_{1-6}$  is 2.32 Å. The  $C_7$ - $Ru_1$ - $H_{1M}$  angles is 88° and  $P_1$ - $Ru_1$ - $H_{1M}$  angles is 83°.

#### Scheme 13



In an effort to extend the synthetic utility, we have begun to explore the coupling reactions of arylamides and alkenes. For example, the treatment of  $C_6H_5CONMe_2$  (0.5 mmol) with cyclopentene (10 mmol) in the presence of cationic ruthenium-hydride complex **26** in CH<sub>2</sub>Cl<sub>2</sub> at 80 °C for 5 h gave an 89 : 11 ratio of the olefination product **27a** and the C-H insertion product **28a** in 52% conversion (eq 49).



The reaction of arylamide substrates with cyclic alkenes in the presence of cationic ruthenium-hydride complex **26** proceeds via a different reaction pathway compared to the reaction of arylketone with alkenes. This result means that C-H activation proceeded selectively at the *ortho*-position of the aromatic ring of arylamide. Furthermore, the major product is olefination product, which is more synthetically useful, compared to the C-H insertion product. To confirm the structure of the coupling products, the hydrogenation reaction was performed on the product mixture using the same catalyst. Thus, the treatment of a crude reaction solution with H<sub>2</sub> (1 atm) at 80 °C in the presence of cationic ruthenium-hydride complex **26** in CH<sub>2</sub>Cl<sub>2</sub> led to **28a** which was isolated by column chromatography on silica gel, and its structure was completely established by spectroscopic methods. Intrigued by this initial discovery, we surveyed a number of different coupling reaction of arylamides and alkenes.



*Figure 1*. Molecular Structure of Cationic Ruthenium-Hydride Complex 26.

# 2.1 Optimization Studies for Coupling of Arylamide with cyclopentene

#### **2.1.1 Catalyst Survey**

Initially, the coupling reaction of a benzamide and cyclopentene was used to screen for catalyst activity (Table 1). Thus, the treatment of  $C_6H_5CONMe_2$  (0.5 mmol) with an excess amount of cyclopentene (2.5 mmol) in the presence of a metal catalyst (5 mol%) in CH<sub>2</sub>Cl<sub>2</sub> at 80 °C was analyzed by GC after 5 h of reaction time. Among the surveyed ruthenium catalysts, complex 26 exhibited uniquely high activity and selectivity for the oxidative coupling product 27a over the simple ortho-C-H insertion product 28a (entries 2 and 3). The addition of HBF<sub>4</sub>•OEt<sub>2</sub> to complex 26 was found to slightly decrease the activity compared to no additive (entry 4).  $RuHCl(CO)(PCy_3)_2$ , which is known to be an effective catalyst for both hydrovinylation and silvlation reaction of alkenes and alkynes, showed no activity for the coupling reaction (entries 5 and 6).<sup>61</sup> It is noteworthy that the reaction catalyzed by RuH<sub>2</sub>(CO)(PPh<sub>3</sub>)<sub>3</sub>, which is well known to be highly efficient in a similar coupling reaction of arylketones with alkenes, did not proceed even when it was carried out in the presence of HBF<sub>4</sub>•OEt<sub>2</sub> (entries 9 and 10). Also, commonly available ruthenium complexes such as  $RuCl_2(PPh_3)_3$ ,  $RuCl_3 \cdot 3H_2O$ ,  $Ru_3(CO)_{12}$  and  $[Ru(p-1)]_2$ cymene)Cl<sub>2</sub>]<sub>2</sub>, which are representative ruthenium catalysts for C-H bond activation, showed no activity for the coupling reaction under the standard reaction conditions.

	$\frac{O}{V}$ $\frac{[cat] (5 \text{ mol}\%)}{CH_2Cl_2, 80 \degree} (1)$	NMe <sub>2+</sub>	NMe <sub>2</sub>
Ť		27a 🚺	28a 🔇
entry	catalyst	ratio ( <b>27a : 28a</b> )	conversion(%) <sup>b</sup>
1	{[(PCy <sub>3</sub> )(CO)RuH] <sub>4</sub> (µ-O)(µ-OH) <sub>2</sub> } ( <b>4</b> )		0
2	4/HBF <sub>4</sub> ·OEt <sub>2</sub>	86 : 14	52
3	$[(\eta^6-C_6H_6)(PCy_3)(CO)RuH]^+BF_4^-$ (26)	88 : 12	52
4	<b>26</b> /HBF <sub>4</sub> ·OEt <sub>2</sub>	80 : 20	48
5	RuHCI(CO)(PCy <sub>3</sub> ) <sub>2</sub>		0
6	RuHCI(CO)(PCy <sub>3</sub> ) <sub>2</sub> /HBF <sub>4</sub> ·OEt <sub>2</sub>		0
7	RuCl <sub>2</sub> (PPh <sub>3</sub> ) <sub>3</sub>		0
8	RuCl <sub>2</sub> (PPh <sub>3</sub> ) <sub>3</sub> /HBF <sub>4</sub> ·OEt <sub>2</sub>		0
9	RuH <sub>2</sub> (CO)(PPh <sub>3</sub> ) <sub>3</sub>		0
10	RuH <sub>2</sub> (CO)(PPh <sub>3</sub> ) <sub>3</sub> /HBF <sub>4</sub> ·OEt <sub>2</sub>		0
11	[RuH(CO)(PCy <sub>3</sub> ) <sub>2</sub> (CH <sub>3</sub> CN) <sub>2</sub> ] <sup>+</sup> BF <sub>4</sub> <sup>-</sup>		0
12	RuCl <sub>3</sub> ·3H <sub>2</sub> O/HBF <sub>4</sub> ·OEt <sub>2</sub>		0
13	Ru <sub>3</sub> (CO) <sub>12</sub> /NH <sub>4</sub> PF <sub>6</sub>		0
14	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub> /PPh <sub>3</sub>		0
15	Re(CO) <sub>3</sub> (THF) <sub>2</sub> Br/HBF <sub>4</sub> ·OEt <sub>2</sub>		0
16	AuCl(PPh <sub>3</sub> ) <sub>3</sub>		0
17	AuCl(PPh <sub>3</sub> ) <sub>3</sub> /HBF <sub>4</sub> ·OEt <sub>2</sub>		0
18	HBF <sub>4</sub> ·OEt <sub>2</sub>		0
19	Cy <sub>3</sub> PH <sup>+</sup> BF <sub>4</sub> <sup>-</sup>		0

Table 1. Catalyst Suvery on the Reaction of C<sub>6</sub>H<sub>5</sub>CONMe<sub>2</sub> with Cyclopentene.<sup>a</sup>

<sup>*a*</sup> Reaction conditions:  $C_6H_5CONMe_2$  (0.5 mmol), cyclopentene (2.5 mmol), catalyst (5 mol%),  $CH_2Cl_2$  (2 mL), 5 h. <sup>*b*</sup> The conversion of  $C_6H_5CONMe_2$  as determined by GC analysis using  $C_6Me_6$  as an internal standard.

## 2.1.2 Solvent Effect

In order to find optimal solvent, the solvent effect on the activity and selectivity was examined for the coupling reaction of arylamides and alkenes. The reaction of  $C_6H_5CONMe_2$  (0.5 mmol) with cyclopentene (2.5 mmol) in the presence of cationic ruthenium-hydride complex **26** was performed in a variety of solvents and the yield of coupling products was determined by GC (Table 2). It was found that the nature of the solvents considerably affects the activity of the coupling reaction.

Both  $CH_2Cl_2$  and PhCl were found to be most suitable for the coupling reaction among the screened organic solvents (entries 1 and 2). A stable 18-electron complex was formed in  $CH_3CN$  because of the solvent's strong coordinating ability, which ultimately leads to no catalytic activity (entry 4). Weakly coordinating solvents such as THF showed lower activity compared to non-coordinating solvents (entry 5). Other solvents such as  $C_6H_6$ , PhCH<sub>3</sub> and Et<sub>2</sub>O also were found to show moderate activity for the coupling reaction. The formation of an equivalent amount of cyclopentane was detected in the crude reaction mixture.

**Table 2**. Solvent Effect on the Coupling Reaction of  $C_6H_5CONMe_2$  with Cyclopentene Catalyzed by **26**.<sup>*a*</sup>

26 (5 mol %) solvent, 80 °C, 5 h	0 NMe <sub>2</sub> +	0 NMe <sub>2</sub> 28a
solvent	ratio ( <b>27a : 28a</b> )	conversion(%) <sup>b</sup>
CH <sub>2</sub> Cl <sub>2</sub>	88 : 12	52
PhCI	89 : 11	51
CICH <sub>2</sub> CH <sub>2</sub> CI	90 : 11	50
CH <sub>3</sub> CN		0
THE	93 : 7	28
Et <sub>2</sub> O	93 : 7	33
C <sub>6</sub> H <sub>6</sub>	92:8	39
$PHOH_3$		33
	$\begin{array}{c} { \begin{array}{c} 26 \ (5 \ mol \ \%) \\ \hline \text{solvent}, \ 80 \ ^{\circ}\text{C}, \ 5 \ h \end{array}} \\ \hline \\ \\ \hline \\ \\ \hline \\$	$\begin{array}{c} & \begin{array}{c} & \begin{array}{c} 26 \ (5 \ mol \ \%) \\ \hline solvent, \ 80 \ ^{\circ}C, \ 5 \ h \end{array} & \begin{array}{c} & \begin{array}{c} & \begin{array}{c} & \begin{array}{c} & \\ & \\ & \end{array} \\ \hline & \\ & \end{array} \\ \hline & \\ & \begin{array}{c} ratio \\ (27a: 28a) \end{array} \end{array} \\ \hline \\ & \begin{array}{c} \\ & \\ & \end{array} \\ \hline \\ & \begin{array}{c} \\ & \\ & \end{array} \\ \hline \\ & \begin{array}{c} \\ & \\ \\ & \end{array} \\ \hline \\ & \\ & \\ & \end{array} \\ \hline \\ & \begin{array}{c} \\ \\ \\ \\ \\ & \end{array} \\ \hline \\ & \\ & \end{array} \\ \hline \\ & \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\$

<sup>*a*</sup> Reaction conditions:  $C_6H_5CONMe_2$  (0.5 mmol), cyclopentene (2.5 mmol), **26** (15 mg, 5 mol%), 5 h. <sup>*b*</sup> The conversion of  $C_6H_5CONMe_2$  as determined by GC analysis using  $C_6Me_6$  as an internal standard.

# **2.2 Reaction Scope**

The scope of the coupling reaction was explored by using the catalyst **26** (Table 3). Both secondary and tertiary arylamides were found to react smoothly with cyclic olefins to give the oxidative coupling products predominantly, but the primary arylamide benzamide was less reactive for coupling reaction with cyclopentene (entry 6). The secondary amides with *N*-electron withdrawing group were found to promote the oxidative coupling product **27** over the insertion product **28** (entries 10-12). Cyclic alkenes generally give the oxidative coupling products **27** preferentially, but cyclohexene resulted in a low yield of the coupling products (entry 13). Steric and electronic environments on the amide group were found to be less important with 1,1-disubtituted and terminal olefins in yielding a mixture of **27** and **28** for these cases (entries 17-24). In contrast, the reaction with 1-hexene yielded a complex mixture of **27** and **28** that resulted from the coupling with both terminal and internal olefins. 3-Acetyl-*N*-methylpyrrole is also a suitable substrate for the oxidative coupling reaction (entry 26).

	•••••••••••••••••••••••••••••••••••••••	- 8- Piii 8			
entry	y substrate	alkene	products	temp (℃)	yield (%) <sup>b</sup>
	X NMe <sub>2</sub>		X NMe <sub>2</sub> X NMe <sub>2</sub> Y Y Y Y	:	
1 2 3	X = H X = Me X = CI		<b>27a 28a</b> (88 : 12) <b>27b 28b</b> (82 : 18) <b>27c 28c</b> (80 : 20)	80 80 80	52 54 62
	NMe <sub>2</sub>	$\bigcirc$		:	
4	Y = OMe		<b>27d 28d</b> (91:9)	80	76
5	Y = CI		27e 28e (88:12)	80	73

**Table 3.** Oxidative C-H Coupling Reaction of Arylamides with Alkenes.<sup>a</sup>



<sup>*a*</sup> Reaction conditions: carbonyl compound (0.5 mmol), cyclopentene (2.5 mmol), **26** (15 mg, 5 mol%), 5 h. <sup>*b*</sup> Isolated yield.

To examine the steric effect for the coupling reaction, substrates with different size alkyl group at nitrogen site have been explored. A change from methyl to isopropyl led to a decrease in the initial reaction rate, but the isolated yield of each substrate was comparable over long reaction time (eq 50). Also we attempted the coupling reaction with more sterically demanding substrate such as N,N,2-trimethylbenzamide, but exceedingly low amounts of product were formed (eq 51). Methyl group at the 2-position in aromatic ring inhibited the coupling reaction with alkenes due to steric repulsion. This suggests that chelation of carbonyl group to ruthenium center is important step to proceed the coupling reaction.



< 5%

## **2.3 Mechanistic Studies**

#### 2.3.1 Phosphine Inhibition

The coupling reaction was strongly inhibited by the addition of phosphines. The amount of coupling products was determined for the coupling reaction of  $C_6H_5CONEt_2$ with cyclopentene in presence of cationic ruthenium-hydride complex 26 (5 mol %) and various amounts of  $PCy_3$ . The conversion dramatically decreased with increasing amount of added PCy<sub>3</sub> ( $0 \sim 6.3 \text{ mol } \%$ ) (Figure 2). For example, addition of 5 mol % of PCy<sub>3</sub> to the reaction mixture led to <10% conversion of substrate to product after 5 h, while 6.3 mol % of PCy<sub>3</sub> was sufficient to effect complete shutdown of the coupling reaction. In an effort to further gain insights of the role of additional phosphine, we monitored the reaction mixture of 26,  $C_6H_5CONEt_2$  with cyclopentene at 80 °C. A new Ru-H peak at  $\delta$  -23.4 (t,  $J_{\rm PH} = 17.2$  Hz) and free benzene peak were observed by <sup>1</sup>H NMR and a new single phosphine signal was detected at  $\delta$  49.3 ppm by <sup>31</sup>P{<sup>1</sup>H} NMR spectroscopy at 80 °C. The hydride peak  $\delta$  -23.4 ppm and the phosphine signal  $\delta$  49.3 ppm were assigned to a new cationic Ru-H complex which has two phosphine ligands, by comparing the spectroscopic data with a similar ruthenium complex  $(PCy_3)_2(CO)RuHCl$  (19) (<sup>1</sup>H NMR  $\delta$  -24.7 (t,  $J_{PH} = 18$  Hz),  ${}^{31}P{}^{1}H$  NMR  $\delta$  46.6). <sup>61b</sup> The phosphine addition study indicated that  $\eta^6$ -benzene ligands are easily dissociated by substrate compared to phosphine ligand under mild reaction conditions.



*Figure 2.*  $PCy_3$  Inhibition Study for the Coupling Reaction of  $C_6H_5CONEt_2$  with Cyclopentene Catalyzed by 26.

# 2.3.2 Isotope Effect Study

A negligible deuterium isotope effect was observed for the reaction of  $C_6H_5CONEt_2$  versus  $C_6D_5CONEt_2$ . The pseudo-first order plots for the catalytic reaction of both  $C_6H_5CONEt_2$  and  $C_6D_5CONEt_2$  with cyclopentene at 80 °C gave  $k_{obs} = 5.3 \times 10^{-3}$  min<sup>-1</sup> and  $k_{obs} = 4.9 \times 10^{-3}$  min<sup>-1</sup>, respectively, from which a negligible isotope effect of  $k_H/k_D = 1.1 \pm 0.1$  was derived (Figure 3). These results indicate that cleavage of the aryl C-H bond is not the rate-limiting step for the coupling reaction.



**Figure 3**. First-Order Plots of  $-ln([C_6H_5CONEt_2]_t/[C_6H_5CONEt_2]_0)$  vs Time for Coupling Reaction.

The carbon isotope effects of  $C_6H_5CONEt_2$  were measured by <sup>13</sup>C NMR analysis following Singleton's carbon isotope measurement technique at natural abundance (eq 52).<sup>62</sup> The virgin  $C_6H_5CONEt_2$  was used as the standard. The <sup>13</sup>C NMR signal intensities were compared between virgn and recovered starting materials using *para*-carbon signal as an internal standard. The carbon isotope effect was observed on *ortho* carbon when the <sup>13</sup>C ratio of recovered  $C_6H_5CONEt_2$  at 78-82% conversion was compared to that the virgin sample (average of 3 runs). As shown in eq 52, the relative proportions of the <sup>13</sup>C isotope composition at *ortho* carbon was the most significantly increased by 2.3%. No other signals showed any significant changes in the <sup>13</sup>C isotopic composition. Using the equations of Melander and Saunder, the  ${}^{12}C/{}^{13}C$  isotope effects are calculated and summarized in eq 52 and Table 4.<sup>63</sup> These isotope effects indicate that the rate-limiting step is the C-C bond formation with arylamides and alkenes.



(recovered at 80% conversion)

*Table 4.* Average <sup>13</sup>C Integration of the Recovered and Virgin Sample of C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub>.

$ \begin{array}{c}                                     $	$23h+24h + 0 1 \\ 8 23h + 24h + 2 3 \\ 5 4 3 \\ 4 3 \\ 4 3 \\ 4 3 \\ 4 3 \\ 4 3 \\ 5 4 \\ 4 3 \\ 4 3 \\ 3 \\ 4 3 \\ 5 \\ 4 \\ 3 \\ 4 \\ 4$
--	--

C #	virgin	recovered (78 % conv.)	recovered/virgin	change (%)
1	1.034	1.041	1.007	0.70
2(ref)	1.000	1.000	1.000	0.00
3	1.975	2.020	1.023	2.27
4	1.986	1.985	0.999	-0.10
5	0.956	0.954	0.998	-0.20

C #	virgin	recovered (80 % conv.)	recovered/virgin	change (%)
1	1.034	1.041	1.007	0.70
2(ref)	1.000	1.000	1.000 1.000 0.	
3	1.975	2.024	2.024 1.023	
4	1.986	1.984	1.984 0.999	
5	0.956	0.957	1.001	0.10
C #	virgin	recovered (82 % conv.)	recovered/virgin	change (%)
1	1.034	1.041	1.007	0.70
2(ref)	1.000	1.000	1.000	0.00
3	1.975	2.023	1.024	2.43
4	1.986	1.988	1.001	0.10
5	0.956	0.953	0.997	-0.30

#### 2.3.3 Deuterium Labeling Study

The following deuterium labeling experiments were performed in order to understand about the reaction mechanism. The treatment of *N*,*N*-diethyl benzamide- $d_5$  (0.5 mmol) and cyclopentene (2.5 mmol) in the presence of **26** (5 mol %) in CH<sub>2</sub>Cl<sub>2</sub> was monitored by <sup>1</sup>H and <sup>2</sup>H NMR spectroscopy. Initially, a rapid and extensive H/D exchange was observed on the *ortho*-arene positions after 1 h at 20 °C. After 1 h at 80 °C, both the coupling products (55% D) along with both unreacted cyclopentene and the amide substrates evidenced significant deuterium incorporation (Scheme 14). The peak intensity at  $\delta$  7.02 ppm, which corresponds to the *ortho* position of the coupling products significantly increases due to rapid H/D exchange with cyclopentene (Figure 4 (a) and (b)). The decreased deuterium signal of *ortho* position of coupling products was further confirmed by <sup>2</sup>H NMR spectroscopy.

Recovered starting materials showed the proton signal with strong intensities at *ortho*-arene positions in its <sup>1</sup>H NMR spectrum. Recovered cyclopentene and cyclopentane exhibited strong deuterium signals in their <sup>2</sup>H NMR spectrum due to intensive deuterium incorporation. Figure 5 (a) showed the <sup>1</sup>H NMR spectrum of *N*,*N*-diethyl benzamide- $d_5$  and while Figure 4 (b) shows the <sup>1</sup>H NMR spectrum of recovered *N*,*N*-diethyl benzamide- $d_n$ . The peaks at  $\delta$  7.28 ppm, which are assignable to the aromatic protons of the arylamide substrate, considerably increased compared to virgin *N*,*N*-diethyl benzamide- $d_5$ . Rapid H/D exchange with cyclopentene in recovered *N*,*N*-diethyl benzamide- $d_5$  (c) and (d)). As shown in Figure 5 (e), significant amounts of deuterium were observed for cyclopentene- $d_n$  and cyclopentane- $d_n$  in the <sup>2</sup>H NMR spectrum of recovered cyclopentene and cyclopentane. These results indicate that C-H bond activation step is reversible at *ortho* position of aromatic ring in substrate.







**Figure 4.** (a) <sup>1</sup>H NMR Spectrum of Mixture of  $23h-d_n$  and  $24h-d_n$ , (b) <sup>2</sup>H NMR Spectrum of Mixture of  $23h-d_n$  and  $24h-d_n$ , (c) <sup>1</sup>H NMR Spectrum 24h, (d) <sup>1</sup>H NMR Spectrum of  $24h-d_n$ .



**Figure 5.** (a) <sup>1</sup>H NMR Spectrum of *N*,*N*-Diethyl Benzamide- $d_5$ , (b) <sup>1</sup>H NMR Spectrum of Recovered *N*,*N*-Diethyl Benzamide- $d_n$ , (c) <sup>2</sup>H NMR Spectrum of Recovered *N*,*N*-Diethyl Benzamide- $d_n$ , (d) <sup>2</sup>H NMR Spectrum of Recovered Cyclopentene- $d_n$  and Cyclopentane- $d_n$ .

#### 2.3.4 Isolation of Catalytically Relevant Species

In an effort to gain insights into the nature of the intermediate species, the reaction of tetrameric ruthenium complex **4** with amide were examined. The protonation reaction between tetrameric ruthenium complex **4** (200 mg, 0.12 mmol) and *N*,*N*-dimethyl-2-naphthamide (100 mg, 0.50 mmol) with HBF<sub>4</sub>·OEt<sub>2</sub> (64 µL) in CH<sub>2</sub>Cl<sub>2</sub> at room temperature cleanly afforded the cationic ruthenium-hydride complex [RuH(PCy<sub>3</sub>)(CO)( $\eta^6$ -*N*,*N*-dimethyl-2-naphthamide)]BF<sub>4</sub> (**29a**), which was isolated as a green-colored solid in 80% yield (Scheme 15). The broad ruthenium-hydride signal was observed at  $\delta$  -21.70 by <sup>1</sup>H NMR spectroscopy in CD<sub>2</sub>Cl<sub>2</sub>, and a broad single phosphine signal was detected at  $\delta$  74.4 ppm by <sup>31</sup>P{<sup>1</sup>H} NMR spectroscopy. Subsequently, the cationic ruthenium-hydride complex **30a** was formed by adding cyclopentene (5 equiv) to the cationic ruthenium-hydride complex **29a** in CH<sub>2</sub>Cl<sub>2</sub> at room temperature. The new ruthenium-hydride signal appeared in the <sup>1</sup>H NMR of **30a** in CD<sub>2</sub>Cl<sub>2</sub> at  $\delta$  -19.58 (d, *J*<sub>PH</sub> = 21.8 Hz) by and a new single phosphine signal exhibited at  $\delta$  65.4 ppm in its <sup>31</sup>P{<sup>1</sup>H} NMR spectrum.

The cationic ruthenium-hydride complex **30a** was isolated in 82% yield from a reaction of **29a** with cyclopentene in CH<sub>2</sub>Cl<sub>2</sub>, and its structure was established by X-ray crystallography (Figure 6). The molecular structure of **30a** showed a distorted square-pyramidal geometry with hydride ligand occupying an axial position. Amide group is *trans* arrangement to carbonyl and olefin group. The C<sub>11</sub>-C<sub>12</sub> bond distance of the olefin group is 1.39 Å, which is comparable to cyclic olefin bond distance. Catalytic activities of the isolated complex **30a** for the coupling reaction of *N*,*N*-dimethyl-2-naphthamide with cyclopentene were found to be identical to that of **26** (eq 53).



Complex **30a** represents a rare example of a structurally characterized cationic ruthenium hydride species which is catalytically active for chelate-assisted C-H activation reactions. Though *ortho*-metalated complexes have been commonly invoked as the key species for chelate-assisted C-H activation chemistry,<sup>1</sup> structurally well-characterized metalated ruthenium complexes were often found to be catalytically inactive for the C-H insertion reactions.<sup>64</sup> More recently Kakiuchi and Yu independently reported the synthesis of catalytically active *ortho*-metalated Ru and Pd complexes and their activity for both C-O and C-H bond activation and insertion reactions of arene compounds, respectively.<sup>65</sup> It is also noteworthy that both Ru(0)-enone and Rh(I)-olefin complexes were found to be active for the C-H insertion of aryl ketones.<sup>66</sup> In our case, the successful isolation of **30a** further suggests that the coupling reaction could be inhibited by the products. Indeed, the coupling reaction C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> with cyclopentene in the presence of 2.5 equiv **27a** showed virtually no activity under otherwise similar conditions as stipulated in eq 49 ( $\leq$ 3% conversion).

Scheme 15



Figure 6. Molecular Structure of 30a.
#### 2.3.5 VT NMR Study

In an attempt to better understand the mechanism of the reaction, the reaction of complex 26 with different arylamides was monitored by NMR. The most noticeable change was observed in the reaction with N,N-dimethyl-2-naphthamide. In a heavywalled J-Young NMR tube, N,N-dimethyl-2-naphthamide (87 µmol) was added into a  $CD_2Cl_2$  solution containing complex 26 (50 mg, 87µmol). When the mixture was monitored at room temperature, the formation of a new species was detected by <sup>31</sup>P NMR spectroscopy ( $\delta$  74.4), and this new species was found to exist as an equilibrium mixture with 26 (26:29a = 9:1) (eq 54). Since no other peak could be assigned, due to the presence of PCy<sub>3</sub> signals, the structure of new species was established from the protonation reaction between the tetrameric ruthenium complex 4 and N,N-dimethyl-2naphthamide with HBF<sub>4</sub>•OEt<sub>2</sub>. Both <sup>31</sup>P NMR signals were identical and all of the spectroscopic data were consistent with the complex 29a. The equilibrium constants between 26 and 29a were estimated from VT <sup>31</sup>P NMR spectroscopy in the temperature range of 30 to 60 °C. The van't Hoff analysis of these data led to the thermodynamic parameters of  $\Delta H^{\circ} = 0.6 \pm 0.1$  kcal/mol and  $\Delta S^{\circ} = -0.5 \pm 0.1$  eu for the equilibrium reaction (Figure 7). These results indicate that the ligand exchange between benzene and arylamides are very facile under the catalytic reaction condition.



*Figure 7.* van't Hoff Plot of *ln K* vs 1/T.

### 2.4 Proposed Mechanism

These results provide support for a mechanism involving vinyl C–H bond activation as depicted in Scheme 16. We propose that the initial arene exchange/ $\pi$ -coordination of an arylamide from 26 would form the arene-coordinated cationic Ru-H species 29. The equilibrium constants between 26 and 29a were measured from VT NMR

experiments using phosphorus signal for both complexes in different temperature ( $K_{eq} = 0.3 \text{ at } 30 \text{ °C}$ ). The phosphine inhibition study implicates that the reactive species would be formed from a reversible dissociation of  $\eta^6$ -benzene ligand. These results indicate that the reversible ligand exchange reaction of **26** with arylamides readily proceeds in the reaction mixture, which are consistent with our proposed mechanism.

Complex 29b further undergoes chelate-directed ortho-C-H activation and dehydrogenation to form the *ortho*-metallated species **31**. The extensive H/D exchange at ortho-position of arylamide as well as deuterated cyclopentene and cyclopentane is observed by both <sup>1</sup>H and <sup>2</sup>H NMR. The negligible isotope effect of  $k_{\rm H}/k_{\rm D} = 1.1 \pm 0.1$  is observed for the coupling reaction of C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> and C<sub>6</sub>D<sub>5</sub>CONEt<sub>2</sub> with cyclopentene. These mechanistic features indicate that the cleavage of aryl C-H bond is reversible and not the rate-limiting step for the coupling reaction. The subsequent vinyl C–H activation and the turnover limiting aryl-to-vinyl reductive elimination steps would form the cationic ruthenium-hydride species **32**. Both carbon isotope effect study which shows the most significantly increased ratio at *ortho* carbon by comparison with virgin sample. The successful isolation of the complex **30b**, which shows identical catalytic activity with respect to the coupling reaction in comparison with cationic ruthenium hydride complex **26**, provide the supporting evidence for the rate-limiting C–C bond formation step. The cationic Ru-H complex 32 would be easily converted to cationic Ru-H complex 29b by dissociation of product/ $\pi$ -coordination of an arylamide. On the other hand, the formation of the insertion product 28 can be readily explained by invoking a direct olefin insertion and the subsequent ortho C-H activation and reductive elimination subsequence from **29b**.

One of the major reasons the oxidative C-H coupling pathway is favored over the

Murai-type C-H insertion pathway may be due to the catalyst's ability to promote facile vinyl C-H activation. Cyclic alkenes are favor to form the metal-vinyl species instead of metal-alkyl species and generate the oxidative C-H coupling product predominantly. While the formation of metal-vinyl species has been well documented in the C–H bond activation literature,<sup>67</sup> its synthetic utility has been rarely exploited in catalytic coupling reactions.





### Conclusions

The cationic ruthenium-hydride complex **26** was found to be an effective catalyst for coupling reaction with arylamides and alkenes via C-H activation. The C-H bonds at *ortho* position of arylamides is selectively cleaved and added to alkenes. The olefination products were produced in high yield and high selectivity when unactivated cyclic alkenes such as cyclopentene, cyclohexane, cycloheptene and cyclooctene, were employed. A catalytically active cationic ruthenium-hydride complex **30** was isolated and characterized. A normal deuterium isotope effect ( $k_{CH}/k_{CD} = 1.1 \pm 0.1$ ) and deuterium labeling studies showed that the C-H bond activation is not the rate-determining step. <sup>13</sup>C isotope effect results indicated that the rate determining step was C-C bond formation which was consistent with our proposed mechanism.

## **Chapter III**

# Stereoselective Catalytic Conjugate Addition and Oxidative Coupling Reaction of Simple Alkenes to α,β-Unsaturated Carbonyl Compounds

Considerable research to the development of the expeditious catalytic synthesis of tetrasubstituted olefins has in part been stimulated by growing demand for developing generally applicable methods for biologically active compounds such as tamoxifen **33** (drug for breast cancer) and rofecoxib **34** (nonsteroidal anti-inflammatory drug) as well as for light/electron responsive organic materials and molecular devices including sensors and switches (Figure 8).<sup>68</sup>



Figure 8. Structure of Tamoxifen 33 and Rofecoxib 34.

Since classical olefination methods are generally not suitable for the synthesis of highly substituted olefins, extensive research has been devoted over the years to develop catalytic olefination methods that do not generate copious byproducts.<sup>4, 33-34, 68-75</sup> Recent notable examples for the synthesis of tetrasubstituted olefins that utilize transition metal catalysis include: Suzuki-type Pd-catalyzed coupling reactions,<sup>69</sup> Ni- and Rh-catalyzed

exocyclization methods,<sup>70</sup> Ni- and Pd-catalyzed nucleophilic coupling reactions of silylacetylenes<sup>71</sup> and of alkyne-to-arylboronic acids,<sup>72</sup> Ti-catalyzed tandem alkyneepoxide-ethyl acetate coupling,<sup>73</sup> and the ring-closing olefin metathesis by using Grubbs' catalyst.<sup>74</sup> Though chelate-assisted catalytic C–H insertion method to  $\alpha$ , $\beta$ -unsaturated carbonyl and imine compounds has successfully been utilized for the synthesis of tri- and tetrasubstituted olefins,<sup>4, 33-34</sup> the general applicable catalytic conjugate addition of simple olefins to  $\alpha$ , $\beta$ -unsaturated carbonyl compounds has been largely elusive due to the lack of reactivity of alkene substrates and the formation of homocoupling and other byproducts. In this regard, the recently reported Ni-catalyzed enone-to-alkene cross coupling reaction and allylic substitution are a notable advance as described below.<sup>75</sup>

Jamison has reported a nickel-catalyzed reaction of enones with alkenes in the presence of  $Et_3SiOTf$  and  $Et_3N$  to give enol silyl ethers, which is more straightforward method for introducing an alkenyl group at the  $\beta$ -position of enones (eq 55).<sup>75a</sup> He has also described the catalytic intermolecular allylic substitution of unactivated, simple alkene catalyzed by nickel complex, which can accommodate a wide range of allylic alcohol derivatives and simple terminal alkenes (eq 56).<sup>75b</sup> Ogoshi has developed a first example of a direct conjugate addition of simple alkenes to enones catalyzed by Ni(0) complex (eq 57).<sup>75c</sup> This reaction is a powerful synthetic tool for the introduction of an alkenyl group at the  $\beta$ -position of enones without any pre-step, since the conventional conjugate addition method requires the preparation of alkenyl metals and its metal must be discarded after reaction. Recently, Montgomery has reported that regioselective catalytic reductive coupling of enones and terminal allenes provides the preparation of



Even though several research groups have reported notable advances for the catalytic synthesis of tetrasubstituted olefins, examples of generally applicable synthetic methods of tetrasubstituted olefins catalyzed by transition metal complexes are relatively rare.

#### **Results and Discussion**

Inspired by recent reports on the C-C bond forming reactions catalyzed by late transition metal complexes,<sup>76</sup> we have begun to explore the synthetic utility of a welldefined cationic ruthenium hydride complex  $\left[\eta^{6}-(C_{6}H_{6})(CO)(PCy_{3})RuH\right]^{+}BF_{4}^{-}$  (26) for the coupling reaction of a variety of carbonyl compounds with simple alkenes.<sup>77</sup> As describe in Chapter 2, we recently discovered that the cationic ruthenium hydride complex 26 is a highly effective catalyst precursor for the oxidative coupling reaction of arylamides with simple alkenes.<sup>77c</sup> While studying the ruthenium-catalyzed C-C bond forming reactions, we recently discovered that novel catalytic synthesis of tetrasubstituted olefins from the intermolecular and intramolecular conjugate addition reaction of simple olefins to  $\alpha,\beta$ -unsaturated carbonyl compounds.<sup>77</sup> For example, propene (2.9 mmol) was added to a  $CH_2Cl_2$  solution of (E)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (0.58 mmol) and a cationic ruthenium complex 26 (3 mol%) and the reaction mixture was stirred at 70 °C for 2 h to obtain stereoselective tetrasubstituted olefin product with 95% yield (eq 59). This catalytic method efficiently produces tetrasubstituted olefin products from the intermolecular coupling of  $\alpha,\beta$ -unsaturated carbonyl compounds and simple alkenes without generating of any byproducts.

$$\begin{array}{c} O \\ Ph \end{array} + CH_3 \xrightarrow{26 (3 \text{ mol}\%)} O \\ CH_2Cl_2, 70 \ \ \ O \\ H_3 \end{array} \xrightarrow{O} O \\ Ph \xrightarrow{O} OEt \end{array}$$
(59)

#### **3.1 Catalyst Survey**

Initial feasibility of the conjugate addition reaction was screened by using a cinnamic acid derivative and an  $\alpha$ -olefin. Thus, the treatment of ethyl cinnamate (0.58 mmol) with propene (2.9 mmol) in the presence of a Ru catalyst (3 mol %) in CH<sub>2</sub>Cl<sub>2</sub> was stirred at 70 °C for 2 h, after which the product conversion was analyzed by GC. Among the screened ruthenium catalysts, the complex **26**, both in isolated form and *in-situ* formed from the of the tetranuclear complex {[(PCy<sub>3</sub>)(CO)RuH]<sub>4</sub>( $\mu$ -O)( $\mu$ -OH)<sub>2</sub>} (**4**) with HBF<sub>4</sub>•OEt<sub>2</sub>, exhibited uniquely high activity in yielding the coupling product **35c** (Table 5). RuH<sub>2</sub>(CO)(PPh<sub>3</sub>)<sub>3</sub>, which was reported by Muari and Trost for the alkylation of  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with alkenes, did not proceed even in presence of an additive HBF<sub>4</sub>•OEt<sub>2</sub> (entry 5). Other readily available ruthenium complexes such as RuCl<sub>2</sub>(PPh<sub>3</sub>)<sub>3</sub>, RuCl<sub>3</sub>•3H<sub>2</sub>O, Ru<sub>3</sub>(CO)<sub>12</sub> and [Ru(*p*-cymene)Cl<sub>2</sub>]<sub>2</sub> also showed no activity for this coupling reaction under the similar conditions.

Ph	$\begin{array}{c} O \\ O \\ O \\ O \\ O \\ C \\ H_2 \\ C \\ C \\ H_2 \\ C \\ C \\ C \\ H_2 \\ C \\ C \\$	at 70 ℃ Ph	O 35c OEt
entry	catalyst	additive	conversion (%) <sup>b</sup>
1	[(η <sup>6</sup> -C <sub>6</sub> H <sub>6</sub> )(CO)(PCy <sub>3</sub> )RuH] <sup>+</sup> BF <sub>4</sub> <sup>-</sup> ( <b>26</b> )	-	99
2	{[(PCy <sub>3</sub> )(CO)RuH] <sub>4</sub> (µ-O)(µ-OH) <sub>2</sub> } ( <b>4</b> )	-	0
3	{[(PCy <sub>3</sub> )(CO)RuH] <sub>4</sub> (µ-O)(µ-OH) <sub>2</sub> } ( <b>4</b> )	HBF <sub>4</sub> ·OEt <sub>2</sub>	99
4	RuHCI(CO)(PCy <sub>3</sub> ) <sub>2</sub>	-	0
5	RuH <sub>2</sub> (CO)(PPh <sub>3</sub> ) <sub>3</sub>	-	0
6	RuCl <sub>2</sub> (PPh <sub>3</sub> ) <sub>3</sub>	-	0
7	RuCl <sub>2</sub> (PPh <sub>3</sub> ) <sub>3</sub>	HBF <sub>4</sub> ·OEt <sub>2</sub>	0
8	[RuH(CO)(PCy <sub>3</sub> ) <sub>2</sub> (S) <sub>2</sub> ] <sup>+</sup> BF <sub>4</sub> <sup>-</sup>	-	0
9	RuCl <sub>3</sub> ·3H <sub>2</sub> O/HBF <sub>4</sub> ·OEt <sub>2</sub>	HBF <sub>4</sub> ·OEt <sub>2</sub>	0
10	Ru <sub>3</sub> (CO) <sub>12</sub>	-	0
11	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub>	-	0
12	HBF <sub>4</sub> ·OEt <sub>2</sub>	-	0

**Table 5.** Catalyst Survey for the Coupling Reaction of (E)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et and Propene.<sup>*a*</sup>

<sup>*a*</sup> Reaction conditions: (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (0.58 mmol), propene (2.9 mmol), catalyst (3 mol%),CH<sub>2</sub>Cl<sub>2</sub> (2 mL), 70 °C, 2-5 h, <sup>*b*</sup> The conversion of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et as determined by GC analysis using C<sub>6</sub>Me<sub>6</sub> as an internal standard. <sup>*c*</sup> S = CH<sub>3</sub>CN.

#### **3.2 Reaction Scope**

The scope of the coupling reaction of carbonyl compounds with alkenes was examined by using the catalyst **26**. Enones,  $\alpha$ , $\beta$ -unsaturated esters and amides were found to react smoothly with simple  $\alpha$ -olefins to give the tetrasubstituted olefin products. Moreover,  $\alpha$ -substituted  $\alpha$ , $\beta$ -unsaturated compounds and intramolecular compounds are also suitable substrates for the coupling reaction with high yield. In general, cinnamic esters and amides with *para*-electron donating group were found to promote the coupling reaction, but neither cyclic enones nor pyrrolinones gave the coupling products under the similar reaction conditions.

#### **3.2.1** Coupling Reactions of α,β-Unsaturated Ester and Enones with Alkenes

The scope of the coupling reaction of enones and  $\alpha,\beta$ -unsaturated esters with simple alkenes was examined by using the catalyst 26 (Table 6). Enones and  $\alpha,\beta$ unsaturated esters were found to react easily with a wide variety of simple olefins to yield the tetrasubstituted olefin products. The reaction of (E)-p-X-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et (X = OEt, Me, H, F, Cl, CO<sub>2</sub>Me, CF<sub>3</sub>) with propene gave the coupling products 35 in high yields (entries 1-7). Both electron donating and withdrawing group at para position in enones and  $\alpha$ , $\beta$ -unsaturated esters gave corresponding tetrasubstituted olefin products in high yields under the similar reaction conditions. A mixture of E/Z olefin products were formed from the coupling reaction of  $\alpha,\beta$ -unsaturated esters with ethylene or 1-pentene, while the coupling reaction with 1-butene and 2-butene gave the same product 35n (entries 13-16). The coupling reaction with cyclopentene showed good activity but poor selectivity due to olefin isomerization (entry 17). A linear olefin products 35q was produced with 4-chlorostyrene (entry 18). Heterocyclic compounds such as (E)-ethyl Nmethyl-2-pyrroleacetate and (E)-ethyl-3-(furan-2-yl) acrylate were found to readily undergo the coupling reaction with propene to give the stereoselective product 35t and **35u** (entries 20 and 21). A cyclic lactone with propene also gave the tetrasubstituted olefin coupling products 35v with a low yield (entry 22).



**Table 6.** Conjugate Addition Reaction of Simple Olefins to  $\alpha$ . $\beta$ -Unsaturated Ester and Enones.<sup>*a*</sup>



<sup>*a*</sup> Reaction conditions: carbonyl compound (0.58 mmol), propene (2.9 mmol), catalyst (3 mol%),  $CH_2Cl_2$  (2 mL), 70 °C, 2-14 h, <sup>*b*</sup> Isolated yield. <sup>*c*</sup> Less than 5% of branched olefin product is formed.

#### **3.2.2** Coupling Reactions of α,β-Unsaturated Amides with Alkenes

In an effort to extend the scope, we next surveyed the conjugate addition of  $\alpha$ , $\beta$ unsaturated amides with simple olefins (Table 7). The reaction of the primary amides such as (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCONH<sub>2</sub> with propene gave a slightly lower yield of the coupling product compared to secondary and tertiary amides (entries 1-11). Steric and electronic environments on the amide group were found to be less important with propene in yielding stereoselective products **36** for these cases (entries 1-11). The fact that both 1- and 2-butenes gave the same product **360** suggested that the rate of olefin isomerization is much faster than the coupling reaction rate (entries 15, 16). It should be emphasized that the  $\alpha$ -olefins with sterically non-demanding group were found to give similar (*Z*)-selective tetrasubstituted olefins (entries 17-20), whereas the coupling reaction with cyclopentene afforded a highly diastereoselective formation of the coupling product **36t** (entry 21). The structure of **36t** was tentatively assigned on the basis of <sup>1</sup>H NMR spectroscopic data. Assignment of the signals for diastereotopic CH proton (H<sub>1</sub>) was established by the magnitude of its vincinal coupling to H<sub>2</sub> (Figure 9). For activated olefins, a mixture of linear and branched olefin products **36u** and **36v** was produced with styrene (entry 22), while ethyl acrylate gave an exclusive formation of the (*Z*)-selective linear coupling product **36w** (entry 23). (*E*)-*N*,*N*-Dimethyl-(naphthalen-2-yl)-2propenamide was also found to react smoothly with propene to give the corresponding tetrasubstituted olefin product **36x** (entry 24).



Figure 9. Newman Projection of Compound 36t.

entry	substrate	alkene	product (s)	t (h) te	emp ( <sup>o</sup> C)	yield (%) <sup>b</sup>
	Ph NR <sub>2</sub>	CH3	Ph NR <sub>2</sub>			
1 2 3 4 5 6 7 8 9 10 11	$R = H, H$ $R = H, Me$ $R = Me, Me$ $R = Et, Et$ $R = i \cdot Pr, i \cdot Pr$ $R = n \cdot Bu, n \cdot Bu$ $R = H, p \cdot Me \cdot C_6 H$ $R = H, Ph$ $R = H, Ph$ $R = H, Bz$ $R = Me, Ph$	4	36a 36b 36c 36d 36e 36f 36g 36h 36i 36j 36k	2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	70 70 70 70 70 70 70 70 70 70 70	75 93 94 95 95 95 94 96 94 94 94
	Ph N		Ph N			
12 13	n = 1 n = 2		36l 36m	2 2	70 70	94 95
14	Ph NHMe	H <sub>2</sub> C=CH <sub>2</sub>	Ph NHMe ( <i>Z</i> )/( <i>E</i> )- <b>36n</b> = 3:1	14	50	78
15 16			Ph NHMe 360	14 14	50 50	64 65
17 18 19 20		$R = (CH_2)_2CH_3$ $R = (CH_2)_4CH_3$ $R = Cy$ $R = CH_2Ph$	R O Ph NHMe 36p 36q 36r 36s	14 14 14 14	50 50 50 50	90 81 90 85
21			H <sup>WF</sup> O H Ph 36t	14	50	89
		Ph Ph Ph	O NHMe Ph	O NH	Ме	
22			<b>36u/36v</b> = 3:2	14	50	70

**Table 7.** Conjugate Addition Reaction of Simple Olefins to α.β-Unsaturated Amides.<sup>a</sup>



<sup>*a*</sup> Reaction conditions: carbonyl compound (0.58 mmol), propene (2.9 mmol), catalyst (3 mol%), CH<sub>2</sub>Cl<sub>2</sub> (2 mL), 70 °C, 2-14 h. <sup>*b*</sup> Isolated yield.

# 3.2.3 Coupling Reactions of $\alpha$ -Substituted $\alpha$ , $\beta$ -Unsaturated Carbonyl Compounds with Alkenes

The scope of the coupling reaction of  $\alpha$ -substituted  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with simple olefins was explored by using the cationic ruthenium hydride complex 26 (Table 8). Generally,  $\alpha$ -substituted  $\alpha$ , $\beta$ -unsaturated carbonyl compounds were found to undergo the coupling reaction with simple olefins to give tetrasubstituted olefin products. However, the tertiary cinnamides which are sterically demanding substrates showed no activity for the coupling reaction with alkenes. The reaction of  $\alpha$ phenyl  $\alpha,\beta$ -unsaturated amide with propene gave slightly lower yield of the coupling product in comparison with  $\alpha$ -methyl  $\alpha$ , $\beta$ -unsaturated amide substrates (entries 4-7). A mixture of E/Z olefin products was formed from  $\alpha$ -substituted  $\alpha,\beta$ -unsaturated amide with ethylene, while the coupling reaction with 1-butenes and 2-butenes gave the same product 37i (entries 8-10). Sterically less demanding  $\alpha$ -olefins such as 1-hexene and 4phenyl-1-butene were found to give tetrasubstituted Z-olefins selectively 37j and 37k, while the coupling reaction with cyclopentene gave highly diastereoselective coupling products which have three different chiral centers (entries 11-13). The structures of the coupling products 371 and 37m were assigned from the <sup>1</sup>H NMR spectroscopic data.

Assignment of the <sup>1</sup>H NMR spectra for the diastereotopic CH protons was established by examining the coupling constants with their vicinal protons (Figure 10). Heterocyclic compound such (*E*)-ethyl-3-(furan-2-yl) 2-methylacrylate with propene yielded tetrasubstituted olefin product **330** (entry 14).



Figure 10. Newman Projection of Compounds 371 and 37m.



**Table 8.** Conjugate Addition Reaction of Simple Olefins to  $\alpha$ -Substituted  $\alpha$ , $\beta$ -Unsaturated Compounds.<sup>*a*</sup>

<sup>&</sup>lt;sup>*a*</sup> Reaction conditions: carbonyl compound (0.58 mmol), propene (2.9 mmol), **26** (3 mol%),  $CH_2Cl_2$  (2 mL), 70 °C, 2-14 h, <sup>*b*</sup> Isolated yield.

## 3.2.4 Oxidative Coupling Reactions of α-Substituted α,β-Unsaturated Carbonyl Compounds with Aromatic Alkenes

The scope of the coupling reaction of  $\alpha$ -substituted  $\alpha,\beta$ -unsaturated carbonyl compounds with aromatic alkenes was further extended by using cationic ruthenium hydride complex 26 (Table 9).  $\alpha$ -Methyl  $\alpha,\beta$ -unsaturated esters (e.g. (E)-p-X- $C_6H_4CH=C(CH_3)CO_2Et$ , X = Me, H, Cl) and  $\alpha$ -methyl  $\alpha,\beta$ -unsaturated amides with sterically demanding group were found to give <5% of the coupling products. In contrast,  $\alpha$ -methyl  $\alpha$ ,  $\beta$ -unsaturated amide with less bulky alkyl group with styrene in the presence of 26 was found to easily afford oxidative coupling products under similar condition (entries 1-5). Styrene derivartives with electron donating groups were found to readily undergo the oxidative coupling reaction with  $(E)-C_6H_5CH=C(CH_3)CONHCH_3$  to give tetrasubstituted diene products (entries 7 and 8). In contrast, aromatic olefins with electron withdrawing group gave low yields for the coupling reaction with (E)- $C_6H_5CH=C(CH_3)CONHCH_3$  (entries 6 and 9). 2-Vinylnaphthalene derivatives were also suitable substrates for the oxidative coupling reaction with (E)- $C_6H_5CH=C(CH_3)CONHCH_3$  to give oxidative coupling products (entries 10 and 11).

entry	substra	ate	alkene	product (s)	t (h)	temp (°C)	yield (%) <sup>b</sup>
x		o ↓ z	<sup>₽h</sup>	Ph			
1 2 3 4 5	X = H X = Me X = Cl X = H X = H	$Z = OEt$ $Z = OEt$ $Z = OEt$ $Z = NMe_2$ $Z = NHMe$		38a 38b 38c 38d 38d 38e	14 14 14 14 14	50 50 50 50 50	<5 <5 <5 51
	Ph	D NHMe		C <sub>6</sub> H <sub>4</sub> -p-Y O Ph NHMe			
6 7 8 9	ο		Y = CI Y = OMe Y = Me Y = Ph	38f 38g 38h 38i Ar	14 14 14 14	50 50 50 50	14 62 56 29
	Ph	NHMe 🥢		Ph NHMe			
10 11			Y = H Y = OMe	38j 38k	14 14	50 50	70 71

**Table 9.** Oxidative Coupling Reaction of Aromatic Olefins to  $\alpha$ -Substituted  $\alpha$ . $\beta$ -Unsaturated Amides.<sup>*a*</sup>

<sup>*a*</sup> Reaction conditions: carbonyl compound (0.58 mmol), propene (2.9 mmol), catalyst (3 mol%), CH<sub>2</sub>Cl<sub>2</sub> (2 mL), 70 °C, 2-14 h, <sup>*b*</sup> Isolated yield.

# **3.4.5 Intramolecular Coupling Reaction of α,β-Unsaturated Esters with Simple Olefins**

To further extend the synthetic applicability of the coupling reaction, we next explored the intramolecular coupling reaction catalyzed by cationic ruthenium hydride complex **26** (Table 10). Thus, the treatment of (*E*)-methyl 3-(2-allylphenyl)acrylate with 5 mol% of **26** in CH<sub>2</sub>Cl<sub>2</sub> at 80 °C led to the formation of 1,2-disubstitued indene product **39a** in 78% isolated yield (entry 1). Both intramolecular substrates, which have 1-butenyl and 2-butenyl-substituted appendages gave the same 1,2-disubstitued indene product **39b** respectively (entries 2 and 3). The result indicates that the olefin isomerization is faster than the cyclization reaction. On the other hand, cyclic olefin substrates were found to readily undergo the oxidative cyclization reaction to form fulvene products **39c-39h** (entries 4-9). Both seven- to eight-membered cyclic olefin substrates showed the similar activity for cyclization (entries 5, 6, 8 and 9), but six-membered cyclic olefin substrates with olefin group exhibited relatively low yield (entries 4 and 7). Even though reaction temperature was increased up to 100 °C, five-membered cyclic olefin substrates exhibited no activity for cyclization.



**Table 10.** Intramolecular Coupling Reaction of  $\alpha,\beta$ -Unsaturated Esters with Simple Olefins Catalyzed by **26**.<sup>*a*</sup>

 $^a$  Reaction conditions: carbonyl compound (0.58 mmol), catalyst (5 mol%), CH<sub>2</sub>Cl<sub>2</sub> (2 mL), 80 °C, 24 h,  $^b$  Isolated yield.

In order to investigate steric effects, we employed a series of cinnamide derivatives with sterically demanding alkyl groups ( $R = CH_3$ ,  $(CH_3)_2$ ,  $(i-Pr)_2$ ) as the substrates for the coupling reaction. Increasing the steric bulk of amide nitrogen position

from methyl to isopropyl led to dramatically decreased yield after 0.5 h, but the coupling reactions with different alkyl groups at nitrogen site gave the comparable yield at 70 °C for 2 h (Eq 60). A sterically more demanding substrate (*E*)-ethyl-3-(2,6-dimethylphenyl)acrylate showed no activity under the similar reaction condition. 2,6-Dimethyl group inhibited the coupling reaction with propene due to steric repulsion by strongly blocking the substrate to coordinate to ruthenium center (eq 61). From these results, a steric effect toward active ruthenium species should play an important role in the coupling reaction.



We next compared the reactivity pattern between *trans* and *cis* conformation of  $\alpha,\beta$ -unsaturated carbonyl compounds. We hypothesized that the coupling reaction proceeds through *trans* conformation for the  $\alpha,\beta$ -unsaturated carbonyl compound and complexation of cationic ruthenium hydride complex *syn* to the ene group on the carbonyl double bond. On the other hand, the *cis* conformation of substrate would disfavor for coupling reaction because chelation of ruthenium to oxygen atom in carbonyl

substrates is key step for coupling reaction. This seems to be true because the coupling reaction of ethyl *cis*-2-furanacrylate with propene in presence cationic ruthenium complex at 20 °C for 0.5 h led to a fast transition from *cis* to *trans* conformation without forming the coupling product (Eq 62). We rationalized that the *cis* conformation would disfavor the the coupling reaction because the sterically crowded *Z*-form substrate would be difficult to coordinate ruthenium center. These results indicate that the chelate-assisted insertion of the ruthenium into the carbonyl substrate is an important step for the coupling reaction.



As mentioned above, we found that the cationic ruthenium complex **26** is a highly effective catalyst for the intermolecular conjugate addition reaction of simple olefins to  $\alpha$ , $\beta$ -unsaturated carbonyl compounds.<sup>77d</sup> The coupling reaction easily generates tetrasubstituted olefins after olefin isomerization step. We hypothesized that the coupling reaction of  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with simple alkenes can easily produce 1,1-disubstituted olefin products. This reaction would be synthetically useful because chiral compounds which can be easily generated from the coupling reaction and 1,1-disubstituted alkenes derived from the conjugate addition reaction are powerful intermediates in synthesis of a diverse range of important compounds. Treatment of (*E*)-*p*-ClC<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et with propene in the presence of a catalytic amount of cationc

ruthenium hydride complex **26** (3 mol%) in  $CH_2Cl_2$  at 0 °C for 48 h gave the coupling products in 80% yield with high diastereoselectivity in the formation of 1,1-disubstituted olefin product (**42**:**35a** = 9:1) (eq 63). (E)-*p*-ClC<sub>6</sub>H<sub>4</sub>CH=C(CH<sub>3</sub>)CO<sub>2</sub>Et was found to react with propene in the presence of **26** at 0 °C for 48 h. The reaction showed high stereoselectivity for 1,1-disubstituted olefin product albeit with a moderate yield (30%) (eq 64).



#### **3.3 Mechanistic Studies**

#### **3.3.1 Reaction Profile**

In order to probe the reaction mechanism of the coupling reaction, the disappearance of starting material as well as the appearance of products was monitored by <sup>1</sup>H NMR for the reaction of (*E*)-*p*-CF<sub>3</sub>C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et (**44**) with propene in the presence of **26** (2 mg, 3.5  $\mu$ mol) at 60 °C. The compound **44** was rapidly converted to compound **45** within 5 min along with the insertion of propene. The signals due to the tetrasubstituted olefin product **35g** gradually appeared at the expense of the compound **45** by isomerization. Eventually, the formation of the tetrasubstituted olefin product **35g** was observed after heating at 60 °C for 3 h (Figure 11).

The kinetics of the conversion of **45** to the tetrasubstitute olefin product **35g** was followed by <sup>1</sup>H NMR. In a J-Young NMR tube, **44** (28 mg, 0.12 mmol), propene (5 equiv), and **26** (2.0 mg, 3.5 µmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> (0.5 mL). The appearance of **45** and **35g** was monitored by <sup>1</sup>H NMR at 60 °C in 5 min intervals. The experimental data were successfully fitted to the kinetic equation for two consecutive reaction kinetics by using nonlinear regression techniques for the conversion of **44** to **35g** (Sigma-plot Version 10). The rate constants  $k_1 = 0.042 \text{ min}^{-1}$  and  $k_2 = 0.011 \text{ min}^{-1}$  were calculated from this analysis.



*Figure 11.* Kinetic Profile of the Conversion of **44** to **35g**. Notations: **44** (●), **45** (■), **35g** (▲).

#### **3.3.2 Hammett Study**

To discern the electronic effects on the product formation, Hammett  $\rho$  values were measured for the coupling reactions with different carbonyl compounds and alkenes. Thus, the correlation of relative rates with  $\sigma_p$  for series of *para*-substituted *p*-X-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et (X = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CO<sub>2</sub>Me, CF<sub>3</sub>) with propene in presence of **26** (3 mol%) at 20 °C led to positive  $\rho$  value ( $\rho = +1.1 \pm 0.1$ ) (Figure 12).



*Figure 12.* Hammett Plot of the Coupling Reaction of *para*-Substituted (*E*)-*p*-X- $C_6H_4CH=CHCO_2Et$  (X = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CO<sub>2</sub>Me, CF<sub>3</sub>) with Propene.

An analogous correlation of the reaction rates of (E)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et with series of *para*-substituted styrene *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y = OCH<sub>3</sub>, CH<sub>3</sub>, H, Cl, CF<sub>3</sub>) also resulted in the positive  $\rho$  value ( $\rho$  = +1.5 ± 0.1) in presence of **26** (5 mol%) at 50 °C in CH<sub>2</sub>Cl<sub>2</sub> (Figure 13). The reaction rates were found to be strongly accelerated by electronwithdrawing group of both cinnamate and styrene derivatives. The similar Hammett  $\rho$ values indicate the same mechanistic pathway for the coupling reaction of  $\alpha$ , $\beta$ unsaturated carbonyl compounds and alkenes. The positive  $\rho$  values indicate that an excess electron density on the transition state is influenced by coordination of both substrates to the ruthenium center. The Hammett  $\rho$  value observed from the correlation of *para*-substituted *p*-X-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et in CD<sub>2</sub>Cl<sub>2</sub> matches well with the chelatedirected regioselective alkene insertion, which is dictated by the nucleophilicity of a developing negative charge on the  $\beta$ -carbon of cinnamate derivatives.<sup>77(d)</sup>



*Figure 13.* Hammett Plot of the Coupling Reaction of (E)-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et with *para*-Substituted Styrene *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y = OCH<sub>3</sub>, CH<sub>3</sub>, H, Cl, CF<sub>3</sub>).

For the coupling reaction of (E)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with *para*substituted styrenes *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CF<sub>3</sub>), a difference in the selectivity pattern was observed depending on the substitution (Scheme 16). The coupling reactions of (E)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene derivatives with *para*-electron donating group (OCH<sub>3</sub>, CH<sub>3</sub>, H) gave the oxidative coupling products **39** predominantly (70 ~ 85 %). In contrast, the coupling reactions of (E)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y = F, Cl, CF<sub>3</sub>) with *para*-electron withdrawing group were found to give a mixture of the branched and linear olefin products **48** and **49** as the major products (75 ~ 85%).

To understand the electronic effects on the product formation for *para*-substituted group of styrene derivatives, the Hammett  $\rho$  value was measured for the coupling reaction of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene derivatives in CH<sub>2</sub>Cl<sub>2</sub>. The correlation of the relative rates with  $\sigma_p$  for (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with a series of *para*-substituted styrene *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CF<sub>3</sub>) in presence **26** ( 5 mol %) at 40 °C in CH<sub>2</sub>Cl<sub>2</sub> led to the positive  $\rho$  values both electron donation and electron withdrawing groups (for electron donating groups:  $\rho = +1.1 \pm 0.1$ , for electron withdrawing groups:  $\rho = +0.9 \pm 0.1$ ) (Scheme 16 and Figure 14). The reaction rate was found to be strongly accelerated by electron-withdrawing group of styrene derivatives, which is consistent with conjugate addition mechanism. The positive value of  $\rho$  indicates that there must be an excess of electron density built up at transition state which can be influenced by the substituent on the arene ring. These results also imply that the rate-limiting step is facilitated by electron withdrawing groups, but the coupling reactions were operated by different mechanistic pathway depending substrate employed.

#### Scheme 16



**Figure 14.** Hammett Plot of the Coupling Reaction of (E)-C<sub>6</sub>H<sub>4</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with *para*-Substituted Styrene *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CF<sub>3</sub>).

#### **3.3.3 Isotope Effect Studies**

The following kinetic experiments were performed in order to gain detailed mechanistic insights on the coupling reaction. The carbon isotope effect of the coupling reaction was measured by employing Singleton's NMR technique at natural abundance.<sup>62</sup> The <sup>13</sup>C NMR signal intensities were compared between the recovered (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCONMe<sub>2</sub> at 75 ~ 80% conversions and virgin sample by using aromatic C<sup>(7)</sup>H signal as an internal standard. As shown in Scheme 17, the most pronounced carbon isotope effect was observed on the β-carbon atom of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCONMe<sub>2</sub> when the <sup>13</sup>C ratio of recovered substrate at 78% conversion was compared to that of the virgin sample (<sup>13</sup>C(recovered)/<sup>13</sup>C(virgin) at C<sub>β</sub> = 1.019, average of three runs). Using the equations developed by Melander and Saunder,<sup>63</sup> the <sup>12</sup>C/<sup>13</sup>C isotope effects are calculated as summarized in Table 11. The results indicate that the olefin insertion to  $\alpha$ ,  $\beta$ -unsaturated carbonyl substrate is the rate-limiting step for the coupling reaction.

Scheme 17



In contrast, a negligible isotope effect of  $k_{\rm H}/k_{\rm D}$  was observed from ethylene and ethylene- $d_4$  with (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et at 60 °C in CD<sub>2</sub>Cl<sub>2</sub>. The pseudo-first-order plots of the catalytic reaction from both ethylene and ethylene- $d_4$  led to  $k_{\rm obs} = 8.3 \times 10^{-4}$  min<sup>-1</sup> and  $k_{\rm obs} = 7.3 \times 10^{-4}$  min<sup>-1</sup>, respectively, which translated to a  $k_{\rm H}/k_{\rm D} = 1.1 \pm 0.1$  (Figure 15). These results indicate that the C-C bond formation step is the rate-limiting step of coupling reaction of  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with simple olefins and subsequent  $\beta$ -hydride elimination and isomerization steps are relatively facile.<sup>77d</sup>

O N	Me <sub>2</sub> +CH <sub>3</sub>	26 (3 mol%) CH <sub>2</sub> Cl <sub>2</sub> , 70 ℃	50 + 36c +	
C #	virgin	recovered (75 % conv.)	recovered/virgin	change (%)
1	1.089	1.084	0.9952	-0.48
2	0.928	0.929	1.0011	0.11
3	1.035	1.054	1.0182	1.82
4	1.047	1.046	0.9990	-0.10
5	2.138	2.134	0.9980	-0.20
6	2.119	2.121	1.0010	0.10
7(ref)	1.000	1.000	1.0000	0.00
C #	virgin	recovered (78 % conv.)	recovered/virgin	change (%)
1	1.089	1.090	1.0005	0.05
2	0.928	0.929	1.0009	0.09
3	1.035	1.054	1.0186	1.86
4	1.047	1.047	0.9990	-0.10
5	2.138	2.139	1.0003	0.03
6	2.119	2.119	0.9999	-0.01
7(ref)	1.000	1.000	1.0000	0.00

**Table 11.** Average <sup>13</sup>C Integration of the Recovered and Virgin Samples of (*E*)- $C_6H_5CH=CHCONEt_2$ .

C #	virgin	recovered (80 % conv.)	recovered/virgin	change (%)
1	1.089	1.090	1.0010	0.10
2	0.928	0.929	1.0011	0.11
3	1.035	1.055	1.0192	1.92
4	1.047	1.045	0.9984	-0.16
5	2.138	2.140	1.0010	0.10
6	2.119	2.121	1.0010	0.10
7(ref)	1.000	1.000	1.0000	0.00





**Figure 15.** First-Order Plots of  $-ln[cinnamate]_t/[cinnamate]_0$  vs Time for the Coupling Reaction of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et with Ethylene ( $\blacklozenge$ ) or Ethylene- $d_4(\bullet)$ .

To discern the rate-limiting step of the coupling reaction of (E)- $C_6H_5CH=C(CH_3)CONHCH_3$  with styrene derivatives which can give different selectivity depending on *para*-substituted styrene derivatives, the carbon isotope effect of the coupling reactions of (E)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with 4-chlorostyrene were measured by employing Singleton's NMR technique at natural abundance. In this case, the most pronounced carbon isotope effect was observed on the  $\beta$ -carbon atom of (E)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> when the <sup>13</sup>C ratio of recovered substrate at 81% conversion was compared to that of the virgin sample  $({}^{13}C(recovered)/{}^{13}C(virgin)$  at  $C_{\beta} = 1.039$ , average of two runs). These results indicate that the olefin insertion into an  $\alpha$ -substituted  $\alpha,\beta$ -unsaturated carbonyl compounds is the rate-limiting step of the coupling reaction which is consistent with previous case.<sup>77d</sup> On the other hand, the negligible carbon isotope effect for coupling reaction of  $(E)-C_6H_5CH=C(CH_3)CONHCH_3$  with styrene in yielding the oxidative coupling product **38e** was observed on  $\beta$ -carbon atom of (E)- $C_6H_5CH=C(CH_3)CONHCH_3$ . The <sup>13</sup>C ratio of recovered substrate to that of the virgin sample[<sup>13</sup>C(recovered)/<sup>13</sup>C(virgin)] was  $C_{\beta} = 0.998$  (average of 2 runs, at 70% conversion). These results of <sup>13</sup>C isotope effect studies are calculated and summarized in Table 12 and Table 13. These observations indicate that the C-C bond forming step is not the rate-limiting step for the coupling reaction of  $C_6H_5CH=C(CH_3)CONHCH_3$  with styrene derivatives in forming oxidative product 38e.



**Table 12.** Average <sup>13</sup>C Integration of the Recovered and Virgin Samples of (*E*)- $C_6H_5CH=C(CH_3)CONHCH_3$ .

	Me + <i>p</i> -Cl-(	C <sub>6</sub> H₄ <mark>26</mark> (3 mol%) C <sub>6</sub> H₅Cl, 100 ℃	<b>52 + 53 +</b> 7	
C #	virgin	recovered (80 % conv.)	recovered/virgin	6 change (%)
1	1.014	1.014	1.0000	0.00
2	0.969	0.967	1.0010	0.10
3	0.970	1.038	1.0381	3.81
4	0.955	0.958	1.0030	0.30
5	2.066	2.069	1.0015	0.15
6	1.992	1.996	1.0020	0.20
7(ref)	1.000	1.000	1.0000	0.00
C #	virgin	recovered (82 % conv.)	recovered/virgin	change (%)
1	1.014	1.013	0.9990	-0.10
2	0.969	0.968	0.9990	-0.10
3	0.970	1.009	1.0402	4.02
4	0.955	0.950	0.9950	-0.50
5	2.066	2.068	1.0010	0.10
6	1.992	1.991	0.9990	-0.10
7(ref)	1.000	1.000	1.0000	0.00






(recovered at 70% conversion)

C #	virgin	virgin recovered (65 % conv.) recovered/virgin		change (%)	
1	1.014	1.015	1.0009	0.09	
2	0.969	0.969	0.9990	-0.10	
3	0.970	0.975	1.0020	0.20	
4	0.955	0.953	0.9980	-0.20	
5	2.066	2.068	1.0010	0.10	
6	1.992	1.991	0.9995	-0.05	
7(ref)	1.000	1.000	1.0000	0.00	

C #	virgin	recovered (67 % conv.) recovered/virgin		change (%)
1	1.014	1.013	0.9990	-0.10
2	0.969	0.970	1.0010	0.10
3	0.970	0.967	0.9970	-0.30
4	0.955	0.958	1.0030	0.30
5	2.066	2.067	0.9995	-0.05
6	1.992	0.993	1.0005	0.05
7(ref)	1.000	1.000	1.0000	0.00

A normal deuterium isotope effect  $k_{\rm H}/k_{\rm D} = 1.7 \pm 0.1$  was observed for coupling reaction of styrene and styrene- $d_8$  with (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> at 40 °C in CH<sub>2</sub>Cl<sub>2</sub>. Also, a negligible isotope effect of  $k_{\rm H}/k_{\rm D} = 1.1 \pm 0.1$  was observed from (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> and (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene under the similar reaction conditions. This result indicates that the cleavage of vinyl C-H bond of styrene is the rate-limiting step for oxidative coupling reaction, which means that this coupling reaction of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene derivatives which have electron donating groups has different mechanistic pathway in comparison with previous case.



**Figure 16.** First-Order Plots of  $-ln([cinnamide]_t/[cinnamide]_0)$  vs Time for the Coupling Reaction of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with Styrene ( $\blacklozenge$ ) or Styrene-*d*<sub>8</sub>( $\blacklozenge$ ).



**Figure 17.** First-Order Plots of  $-ln([cinnamide]_t/[cinnamide]_0)$  vs Time for the Coupling Reaction of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHMe ( $\blacklozenge$ ) or (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHMe ( $\blacklozenge$ ) with Styrene.

#### **3.3.4 Deuterium Labeling Studies**

The following deuterium labeling studies were performed to learn more about the reaction mechanism. To examine H/D exchange pattern on the carbonyl substrate, the reaction mixture of (*E*)-C<sub>6</sub>D<sub>5</sub>CD=CDCONMe<sub>2</sub> (0.58 mmol) with propene (2.9 mmol) in the presence of **26** (3 mol %) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) was stirred at 70 °C for 2 h. The isolated product **36c** was found to contain ~55% D on the  $\alpha$ -methylene position, but with only 6%

of D on the  $\delta$ -methyl positions (eq 65). Also, a significant deuterium incorporation was observed on unreacted propene.

Figure 18(a) shows the <sup>1</sup>H NMR spectrum of the compound **36c**- $d_n$ . The intensities of the peaks at  $\delta$  3.37 and 1.81 ppm which were assigned to methylene and methyl resonances, respectively, significantly decreased due to extensive deuterium incorporation. As shown in Figure 18(b), the existence of deuterium at each position was further confirmed by <sup>2</sup>H NMR. The recovered propene also observed an extensive deuterium incorporation on both vinyl and methyl positions (Figure 18(c)).



*Figure 18.* (a) <sup>1</sup>H NMR Spectrum of **36c**- $d_n$ , (b) <sup>2</sup>H NMR Spectrum of **36c**- $d_n$ , (c) <sup>2</sup>H NMR Spectrum of Recovered Propene.

To examine the H/D exchange pattern on the  $\alpha$ -substituted carbonyl substrate, the treatment of (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub> (0.07 mmol) and styrene (5 equiv) in the presence of **26** (5 mol%) in CD<sub>2</sub>Cl<sub>2</sub> was monitored by <sup>1</sup>H and <sup>2</sup>H NMR (eq 66). Initially, extensive H/D exchange was observed on both the  $\beta$ -position of carbonyl compound and vinyl position of styrene after 15 h at 20 °C without forming product. Such extensive H/D scrambling pattern is consistent with reversible C-H bond activation of  $\beta$ -position of (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub>. These results indicate that vinyl C-H bond activation of  $\alpha$ -substituted cinnamide substrates is not the rate limiting step of coupling reaction with styrene, for the styrenes with electron donating group at *para*-position.



Figure 19(b) shows the <sup>2</sup>H NMR spectrum of the coupling reaction mixture of (E)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene in presence of **26** after 15 h at 20 °C. The intensity of the peak at  $\delta$  7.32 assigned to two vinyl resonance noticeably decreased due to extensive deuterium incorporation. As shown in Figure 19(b), new peaks due to styrene signals were observed by <sup>2</sup>H NMR spectrum.



**Figure 19.** (a) <sup>2</sup>H NMR Spectrum of (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub>, (b) <sup>2</sup>H NMR Spectrum of the Coupling Reaction Mixture of (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with Styrene.

#### **3.3.5 Isolation of Catalytically Relevant Species**

In an effort to trap/isolate catalytically relevant species, the reaction mixture of complex **26** (10 µmol) with a naphthyl-substituted amide (10 µmol), cyclopentene (5 equiv) and H<sub>2</sub>O (10 equiv) in CD<sub>2</sub>Cl<sub>2</sub> was followed by NMR (Scheme 20). The formation of the Ru-allyl complex **56** was detected after 5 h at room temperature. In a preparatory scale reaction, the complex **56** was most conveniently isolated from the reaction mixture of the tetrameric ruthenium complex {[(PCy<sub>3</sub>)(CO)RuH]<sub>4</sub>(µ-O)(µ-OH)<sub>2</sub>} (**4**) with the amide (0.1 mmol), HBF<sub>4</sub>•OEt<sub>2</sub> (4 equiv) and cyclopentene (20 equiv) in wet CH<sub>2</sub>Cl<sub>2</sub>, and its structure was unequivocally established by X-ray crystallography (Figure 20). The X-ray crystal structure of **56** showed that ruthenium atom has a distorted octahedral coordination. The cation complex is totally disordered (in a 2:1 ratio) over non-crystallographic mirror plane. The Ru<sub>1</sub>-C<sub>11</sub>, Ru<sub>1</sub>-C<sub>11</sub> and Ru<sub>1</sub>-C<sub>11</sub> distances for the complex **56** are 2.22 Å, 2.17 Å and 2.14 Å, respectively. The bond lengths of Ru<sub>1</sub>-O<sub>2</sub> are 1.98 Å. The O<sub>1w</sub>-Ru<sub>1</sub>-O<sub>1</sub> angles are 87° and O<sub>2</sub>-Ru<sub>1</sub>-C<sub>11</sub> angles are 83°.

The complex **56** was found to exhibit virtually identical activity as **26** in mediating the coupling reaction of (E)-C<sub>6</sub>H<sub>5</sub>CD=CHCONHCH<sub>3</sub> and propene under the conditions stipulated in eq 59; the reaction gave **36b** in > 90% yield after 2 h. When this reaction was performed in the presence of 1.5 equiv of H<sub>2</sub>O, a substantially lower product conversion (75% after 2 h) was observed. To further establish the catalytic relevance of the complex **56**, the exchange reaction of **56** with (*E*)-PhCH=CHCONHMe (1 equiv) was monitored by <sup>1</sup>H NMR. The reversible coordination of the amide substrate was observed at room temperature to form a 2:1 ratio of **56** and carbonyl-coordinated complex, but no new Ru-H species has been detected even after heating at 60 °C. Though more careful

study is needed to establish the reaction step for coverting catalytically active species, these results suggest that the Ru-allyl complex 56 would be converted to catalytically active Ru-H species by ligand exchange with substrate and H<sub>2</sub>O.

### Scheme 20



Figure 20. Molecular Structure of 56.

#### **3.4 Empirical Rate Law Derivation**

The kinetics of the catalytic coupling reaction of (*E*)-ethyl cinnamate with propene was merasured. The treatment of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (0.07 mmol) and propene (5 equiv) in the presence of **26** (5 mol%) in CD<sub>2</sub>Cl<sub>2</sub> at 20 °C was monitored by <sup>1</sup>H NMR. Initially, the rate was merasured by monitoring the <sup>1</sup>H integration of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (OCH<sub>2</sub>CH<sub>3</sub> (1.21 ppm)) and the product (OCH<sub>2</sub>CH<sub>3</sub> (1.16 ppm)) signals, and these were normalized against the solvent resonance. The kinetic plot for the reaction rate as a function of catalyst concentration indicates that reaction rate is firstorder (Figure 21). The kinetic data revealed that the rate of coupling reaction is independent of the propene concentration (Figure 22). In contrast, the plot of kinetic data indicates that the rate of coupling reaction is inverse first-order of substrate concentration (Figure 23). The empirical rate law is given in eq 67.

$$v = k[\mathbf{26}]^{1}[\text{propene}]^{0}[\text{cinnamate}]^{-1}$$
(67)



*Figure 21.* Plot of Observed Reaction Rate vs Catalyst Concentration for the Coupling Reaction.



*Figure 22.* Plot of Observed Reaction Rate vs Propene Concentration for the Coupling Reaction.



*Figure 23.* Plot of Observed Reaction Rate vs (E)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et Concentration for the Coupling Reaction.

## **3.5 Thermodynamic Parameters**

The temperature dependence on the coupling reaction of (E)-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et with propene mediated by **26** was monitored by <sup>1</sup>H NMR. Both catalyst concentration

and the cinnamate molar ratio were fixed as the coupling reaction conditions. The excess propene concentration not only maintains approximately zero-order conditions throughout the reaction but also minimizes cinnamate inhibition. Standard Eyring (Figure 24) and Arrhenius analysis (Figure 25) derived from the kinetic data resulted the following activation parameters:  $\Delta H^{\Box} = 20.3$  kcal/mol,  $\Delta S^{\Box} = -42.1$  eu, and  $E_a = 20.9$ kcal/mol. A large, negative  $\Delta S^{\Box}$  value suggests that the transition state is highly organized.



*Figure 24.* Eyring Plot for the Coupling Reaction of (E)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et with Propene.



*Figure 25*. Arrhenius Plot for the Coupling Reaction of (E)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et with Propene.

## **3.6 Proposed Mechanism**

We propose three different mechanistic pathways to explain the coupling reactions of  $\alpha$ , $\beta$ -unsaturated carbonyl compounds with alkenes. The plausible mechanism of the conjugate addition reaction involves the cationic Ru-H species **57**, which is initially formed from the ligand exchange step of **26** with the carbonyl substrate (Scheme 21).<sup>11d</sup> The key features of this mechanistic pathway are the chelate-directed regioselective alkene insertion and  $\beta$ -hydride elimination steps to form the cationic Ru/alkene/hydride species **58**.<sup>77d</sup> Both the observation of the carbon isotope effect on  $\beta$ -carbon (<sup>13</sup>C ratio of recovered sample/virgin sample:  $C_{\beta} = 1.019$ ) and a negligible isotope effect of  $k_{\rm H}/k_{\rm D} = 1.1 \pm 0.1$  with ethylene and ethylene- $d_4$  support the rate-limiting olefin insertion step (Table 14). The positive Hammett  $\rho$  values for both carbonyl compounds and alkenes are also consistent with the formation of the carbonyl-chelated species **58** 

followed by the regioselective olefin insertion and  $\beta$ -hydride elimination. The insertion step is dictated by the nucleophilicity of a developing negative charge on the  $\beta$ -carbon of cinnamate derivatives. In light of the recent deuterium labeling study on the alkene dimerization and isomerization reactions,<sup>77c</sup> the subsequent olefin isomerization step is expected to be facile in forming the tetrasubstituted olefin products with the regeneration of **57**. The successful isolation of the catalytically relevant species **56** suggests complex **57** as a possible intermediate, which can undergo dehydrogenation and then trapping by a water molecule.<sup>77d</sup> The catalytically relevant species **56** would be converted to catalytically active species **57** by ligand exchange with hydrogen form water.

	Conjugate Addition Reaction	Oxidative Coupling Reaction
Hammett $\rho$ value	$\rho = + 1.12^{a}$	$\rho = + 1.12^{c}$
	$\rho = +1.47^b$	$\rho = + 0.88^d$
<sup>13</sup> C Isotope Effect	$C_{\beta} = 1.018^{e}$	$C_{\beta} = 0.998^{f}$
Deuterium Kinetic Isotope Effect	$k_{\rm H}/k_{\rm D} = 1.1 \pm 0.1^{g}$	$k_{\rm H}/k_{\rm D} = 1.7 \pm 0.1^{h}$
Rate Limiting Step	Alkene Insertion	Vinyl C-H Activation

**Table 14.** Kinetics and Isotope Effects for the Coupling Reaction of  $\alpha$ , $\beta$ -Unsaturated Carbonyl Compounds with Alkene.

<sup>*a*</sup> Hammett  $\rho$  value ((*E*)-*p*-X-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et with propene). <sup>*b*</sup> Hammett  $\rho$  value ((*E*)-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et with Y-*p*-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub>). <sup>*c*</sup> Hammett  $\rho$  value ((*E*)-C<sub>6</sub>H<sub>5</sub>CH=CH(CH<sub>3</sub>)CONHMe with Y-*p*-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y=OCH<sub>3</sub>, CH<sub>3</sub> H)). <sup>*d*</sup> Hammett  $\rho$  value ((*E*)-C<sub>6</sub>H<sub>5</sub>CH=CH(CH<sub>3</sub>)CONHMe with Y-*p*-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (Y=F, Cl, CF<sub>3</sub>)). <sup>*e*</sup> <sup>13</sup>C ratio (recovered/virgin) ((*E*)-C<sub>6</sub>H<sub>4</sub>CH=CH(CH<sub>3</sub>)CONHMe with styrene). <sup>*f*</sup> <sup>13</sup>C ratio (recovered/virgin) ((*E*)-C<sub>6</sub>H<sub>4</sub>CH=CH(CH<sub>3</sub>)CONHMe with styrene). <sup>*g*</sup> Kinetic isotope effect ((*E*)-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et with ethylene/ethylene-*d*<sub>4</sub>). <sup>*h*</sup> Kinetic isotope effect ((*E*)-C<sub>6</sub>H<sub>4</sub>CH=CH(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene/styrene-*d*<sub>8</sub>).



The formation of oxidative coupling products **38** can be explained by a mechanistic pathway involving vinyl C-H bond activation of arylalkenes (Scheme 22). The insertion of arylalkenes to the electrophilic ruthenium center followed by the ligand exchange reaction of **26** with carbonyl substrate and the  $\beta$  C-H bond activation of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> would form the ruthenium hydride species **59**. Subsequent vinyl C-H activation of arylalkenes and reductive elimination would give *E* selectivity for oxidative coupling products.

We performed a variety of kinetic experiments to elucidate the mechanism of the coupling reaction. Both the normal deuterium kinetic isotope effect of  $k_{\rm H}/k_{\rm D} = 1.7 \pm 0.1$  with styrene and styrene- $d_8$  and negligible isotope effect of  $k_{\rm H}/k_{\rm D} = 1.1 \pm 0.1$  with (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> and (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub> are consistent with the vinyl C-H bond cleavage rate-limiting step. These results indicate that the different rate-limiting step compared to the conjugate addition reaction (Table 14). Both negligible carbon isotope effect from the coupling reaction with styrene and deuterium labeling study also provide supporting evidences for the rate-limiting vinyl C-H bond activation step. The reaction rate and product selectivity were found to be increased by styrene containing electron donating groups, depending on electron donating abilities of *para* functional group.

In contrast, the coupling reaction of (E)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with styrene containing electron withdrawing groups gave a mixture of the branched and linear olefin insertion products predominantly which were similar fashions with alkene insertion pathway.<sup>11d</sup> This phenomenon indicates that the coupling reaction depending on the substrate used exhibits different product selectivity. The most pronounced carbon isotope effect was observed on the  $\beta$  carbon atom of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with 4chlorostyrene. These results also indicate that the formation of C-C bond via olefin insertion into a carbonyl substrate is the rate-limiting step, and are consistent with alkene insertion pathway instead of the oxidative coupling pathway. These results suggest that two different reaction mechanisms are competing operational dependent on the substrate employed due to electron effect.

## Scheme 22



In case of the coupling reaction of cyclic cinnamic acid derivatives with propene, we observed the formation of tetrasubstituted olefin products via a direct C-H bond cleavage (eq 68). Initially, the cationic Ru-H complex **61** is formed form the ligand exchange reaction of **26** with cyclic  $\alpha,\beta$ -unsaturated compounds. The chelate-assisted insertion of the ruthenium center into  $\beta$ -position by C-H bond cleavage gives the cationic Ru-H species **62** after hydrometallation of **61**. Reductive elimination from **62** provides products **60a** and **60b**, regenerating the catalytically active species **62**. The stereochemistry around the double bond is retained in each step. Trost and Murai also reported that the Ru-catalyzed coupling reactions of 1-acetylcyclohexene and  $\alpha,\beta$ unsaturated esters with alkenes such as silylethylenes and styrene by direct C-H bond cleavage.<sup>8</sup> The apparent lack of the formation of conjugate addition products or oxidative products can be readily explained by the rigidity of the cyclic moieties, which would prohibit the insertion of the  $\alpha,\beta$ -unsaturated ester by hydride migration or vinyl C-H activation of alkenes.





## Conclusions

An effective catalytic conjugate addition reaction protocol has been developed for the synthesis for tetrasubstituted olefin products. The kinetic and spectroscopic data are consistent with three different mechanistic pathways depending on substrate employed. The reaction of an  $\alpha,\beta$ -unsaturated carbonyl compounds with simple olefins predominantly gives the tetrasubstituted olefin products. The mechanistic studies suggest that the olefin insertion into an  $\alpha,\beta$ -unsaturated carbonyl substrate is the rate-limiting step for conjugate addition reaction. In contrast, the coupling reaction of (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> with aromatic alkenes gives oxidative coupling products. The mechanistic results support the different mechanistic pathway, which involves the rate-limiting vinyl C-H activation step, in comparison with the conjugate addition reaction. This synthetic methodology promises to provide a straightforward route to the valuable organic compounds such as tetrasubstituted olefins which are difficult to synthesize by using traditional synthetic methods.

## **Chapter IV**

# Aqueous Phase C-H Bond Oxidation Reaction of Arylalkanes Catalyzed by a Water-Soluble Cationic Ru(III) Pymox Complex

C-H oxidation reactions play an important role in organic chemistry and there is an increasing demand for selective and mild oxidation methods in modern organic synthesis. Saturated hydrocarbon compounds account for the largest sources of natural petroleum feedstock materials, which traditionally have been used as fuels, lubricants, and solvents. During the past few decades, there has been an extensive research devoted on the catalytic oxidation of inexpensive hydrocarbons to more commercially valuable products such as pharmaceuticals, flavors, fragrances, plasticizers, and polymer precursors.<sup>78</sup> Many methods for catalytic oxidations with transition metal catalysts have been investigated extensively because of their particular importance in view of synthetic, mechanistic, and industrial aspects.<sup>79</sup> A number of transition metal catalyzed reaction have been extensively investigated including iron-catalyzed oxidation of alkanes called Gif systems.

The ruthenium catalysts have been used to be oxidation of hydrocarbons with *t*-BuOOH to give corresponding ketones and alcohols efficiently.  $RuCl_2(PPh_3)_3$  (63) was reported to be effective catalyst for oxidation of the benzylic position of hydrocarbons. For example, the catalytic oxidation of fluorine gives fluorenone in 87% (eq 69).<sup>80</sup>



The mechanism of this reaction has been studied by Murahashi. Hammett  $\rho$  value showed -1.39 which was similar to those obtain for the oxidation with cytochrome P450 (-1.6)<sup>81</sup> and Fe(TPP)Cl-PhIO (-1.69)<sup>82</sup> but the observed  $\rho$  value (-1.39) is quite different from the *t*-BuO• radical (-0.4)<sup>83</sup> and the *t*-BuOO• radicals (-0.6)<sup>84</sup> (Table 1). These kinetic data implicate the existence of a metal-dependent species such as oxo-ruthenium, which is similar to Fe<sup>V</sup>=O of cytochrome P450, rather than *t*-BuO• and *t*-BuOO• intermediates. The intramolecular deuterium isotope effect ( $k_{\rm H}/k_{\rm D}$ ) was examined. The  $k_{\rm H}/k_{\rm D}$  value was determined to be 9.0 and this value was similar to that obtained for oxidation with cytochrome P450 ( $k_{\rm H}/k_{\rm D} = 11$ ),<sup>85</sup> This data indicates that the C-H bond breaking is the rate determining step. Furthermore, the intermolecular isotope effect of the oxidation of cyclohexanes was determined to be 9.2. This is contrast to the smaller isotope effect ( $k_{\rm H}/k_{\rm D} = 4.2$ ) which was obtained from the reaction with *t*-BuO• (Table 15).<sup>86</sup>

The kinetic results, isotope effects, and other studies indicate that the oxidation is not due to *t*-BuO• or *t*-BuOO• radical but due to the oxo ruthenium species derived from low valent ruthenium and *t*-BuOOH (Scheme 24). The ruthenium(II) complex **A** reacts with *t*-BuOOH to give the alkylperoxoruthenium(II) complex **B**, which subsequently undergoes heterolytic cleavage of the O-O bond to give oxo-ruthenium(IV) species **C**. The reaction with *t*-BuO• or *t*-BuOO• is unlikely, because the oxidation of alkanes proceeds efficiently even in the presence of radical scavengers such as 2,6-di-*tert*-butyl4-methylphenol. Abstraction of a hydrogen atom from an alkane by intermediate **C** would form the caged complex **D**, bearing an alkyl radical and a hydroxo-ruthenium(III) species. Transfer of the hydroxyl ligand to the caged alkyl radical (path **A**) would afford an alcohol and the Ru(II) intermediate **F** to complete the catalytic cycle. Further oxidation of secondary alcohol formed the corresponding ketones under the reaction conditions. When the alkyl radical **D** has low oxidation potential, fast single-electron transfer would take place to give cation **E** (path **B**), which undergoes reaction with the second molecule of *t*-BuOOH to give the corresponding *t*-butylperoxy product and water. From all data such as  $\rho$  values,  $k_{\rm H}/k_{\rm D}$  ratios, and relative oxidation rates in the various substrates, they proposed mechanism for RuCl<sub>2</sub>(PPh<sub>3</sub>)<sub>3</sub>/*t*-BuOOH system which was containing the oxo-ruthenium species (Scheme 24).

system	ρ	k <sub>H</sub> /k <sub>D</sub> <sup>a</sup>	k <sub>H</sub> ∕k <sub>D</sub> <sup>b</sup>
RuCl <sub>2</sub> (PPh <sub>3</sub> ) <sub>3</sub> / <i>t</i> -BuOOH <sup>c</sup>	-1.39	9.0	9.2
cytochrome P450	-1.6 <sup>d</sup>	11 <sup>e</sup>	
[Ru(dmp) <sub>2</sub> (OH <sub>2</sub> ) <sub>2</sub> ][[PF <sub>6</sub> ] <sub>2</sub> /H <sub>2</sub> O <sub>2</sub>			4.0 <sup><i>f</i></sup>
[Ru(6,6'-Cl <sub>2</sub> bpy) <sub>2</sub> (OH <sub>2</sub> ) <sub>2</sub> ][CF <sub>3</sub> SO <sub>3</sub> ] <sub>2</sub> / <i>t</i> -BuOOH <sup>g</sup>			4.8
<i>t-</i> BuO∙	-0.4 <sup>h</sup>		4.2
t-BuOO∙	$-0.6^{i}$		

**Table 15.** Kinetics, Intra- and Intermolecular Deuterium Isotope Effects for the Ruthenium Catalyzed Oxidation of Hydrocarbons with *t*-BuOOH.

<sup>*a*</sup> Intramolecular isotope effect for the oxidation of 1,1-dideuterio-1,3-diphenylpropane.

<sup>b</sup> Intermolecular isotope effect for the oxidation of cyclohexane/cyclohexane- $d_{12}$ .

<sup>c</sup> Reference 80. <sup>d</sup> Reference 81. <sup>e</sup> Reference 85. <sup>f</sup> Reference 88. <sup>g</sup> Reference 87.

<sup>*h*</sup> Reference 83. <sup>*i*</sup> Reference 84.



In contrast, Che reported that the ruthenium(II) complex *cis*-[Ru(6,6'-Cl<sub>2</sub>bpy)<sub>2</sub>(OH<sub>2</sub>)<sub>2</sub>][CF<sub>3</sub>SO<sub>3</sub>], (**64**) is a robust catalyst for C-H bond oxidations of hydrocarbons. They proposed different mechanism in the reaction which was called to be a free radical system. <sup>87</sup> *cis*-[Ru(6,6'-Cl<sub>2</sub>bpy)<sub>2</sub>(OH<sub>2</sub>)<sub>2</sub>][SF<sub>3</sub>SO<sub>3</sub>]<sub>2</sub>, (**64**) was found to efficient catalyst for oxidation of the alkanes, including linear alkanes such as *n*-hexane. Typically, the catalytic oxidation of cyclohexane gives cyclohexanol and cyclohexanone in 85% yield (ratio = 1:1.6) (eq 70).

A smaller kinetic isotope effect ( $k_{\rm H}/k_{\rm D} = 4.8$ ) and tertiary to secondary C-H bond relative reactivites ( $k_{tert}/k_{sec} = 7.4$ ) have been observed in the catalytic cyclohexane and adamantine oxidations, respectively. Also Drago and coworkers found very similar data for the catalytic  $[Ru(dmp)_2(OH_2)_2][PF_6]_2/H_2O_2$  system  $(k_H/k_D = 4.0$  for cyclohexane;  $k_{tert}/k_{sec} = 7.1$  for adamantane; dmp = 2,9-dimethyl-1,10-phenanthroline).<sup>88</sup> From the results of cyclohexane oxidation, they suggested that the active species has substantial radical character: (a) addition of 2,6-di-*tert*-butyl-4-methylphenol as a radical trap suppressed the catalytic oxidation; (b) a substantial amount of cyclohexyl chloride was produced in the presence of CCl<sub>4</sub>; (c) when the terminal oxidant was changed from *t*-BuOOH to cumene hydroperoxide, the product profile is dominated by acetophenone, which is generated by  $\beta$ -scission of cumyloxy radicals. Following these kinetic and mechanistic data, they proposed a free-radical process mechanism which generates active alkyl radical from metal-oxo intermediate.

It is remarkable that the allylic positions of hydrocarbons were oxidized selectively by dirhodium caprolactamate  $(Rh_2(cap)_4)$ , (65) with *t*-BuOOH (eq 71).<sup>89,</sup> Most of cyclohexene and their derivatives were rapidly converted to enones in 1 h with 0.1 mol% 65.



Mechanistically, **65** undergoes a l-electron oxidation in the presence of *t*-BuOOH to form a  $Rh^{5+}$  species **66** which can be converted under the reaction conditions to

dirhodium alkylperoxo species **67** (Scheme 25). Furthermore, evolution of oxygen during the course of the reaction was observed and is consistent with the formation of *t*-BuOO• radical which is known to di-*tert*-butyltetraoxide<sup>90</sup> and it is able for selective hydrogen atom abstraction to produce radical **68**. Ligand transfer of the metal-bound peroxide to the carbon-centered radical forms the mixed peroxide **69** and regenerates the catalyst. This proposed sequence thus constitutes a redox chain catalytic cycle.<sup>91</sup> Finally, rapid decomposition of **55** yields products.





Recently, White and coworkers have reported that an iron-based catalyst ([Fe(*S*,*S*-PDP)(CH<sub>3</sub>CN)<sub>2</sub>][SbF<sub>6</sub>]<sub>2</sub>), (**70**) that reacts selectively with C-H bonds which has traditionally used to add directing groups, require extra steps.<sup>33</sup> The iron-based catalyst makes it possible to oxidize the unreactive aliphatic C–H bonds at tertiary carbons in

complex molecules without the need for directing or activating groups. Predictable selectivity is achieved solely based on the electronic and steric properties of the C-H bonds, without the need for directing groups. Three modes of selectivity enable the predictable oxidation of complex natural products and their derivatives such as (+)-aremisinin at specific C-H bonds with preparatively useful yields (eq 72). This type of general and predictable reactivity stands to enable aliphatic C-H oxidation to streamline the process of complex molecule synthesis.



EWG : electronic withdrawing group BG : bulky group DG : directing group



The use of transition metals for catalyzing reactions is of growing importance in modern organic chemistry.<sup>93</sup> From economic and environmental points of view, the development of novel organic transformation systems in aqueous solution has become one of the most attractive research topics in modern chemistry.<sup>94</sup> Generally, transition metal catalysis is carried out under an inert gas atmosphere and the exclusion of moisture has been essential. The reason could be that water as a ligand often occupies metals' coordination sites. On the other hand, metal-carbon bonds are easily attacked by water either via proton transfer or via the oxygen (nucleophilic reaction). However, recent studies have shown that, in water, such catalysts are both facilitated and can be operated in open air.<sup>95</sup> Another predictable advantage of these water soluble catalysts can be reused after separation from water insoluble organic products.

In the 1970s Shilov et al. developed a catalytic oxidation in which methane was converted to methanol and chloromethane in acidic aqueous solution (eq 73).<sup>96</sup> Since Shilov's pioneering report on methane oxidation catalyzed by  $PtCl_4^{2-}$  under mild conditions, a number of well-defined, transition metal catalytic systems have been shown to catalyze methane functionalization. Notable recent example of methane oxidation have been reported by Periana using (bpym)PtCl<sub>2</sub> (bpym = 2,2'-bipyrimidine).<sup>97</sup> Periana system converts methane to methyl bisulfate in concentrated sulfuric acid, a potential precursor for methanol (eq 74). The conversion of methane to methyl bisulfate is greater than 70% based on methane and the selectivity is also over 90%. This system accomplishes C-H activation, oxidation, and functionalization in the best way so far achieved for a homogeneous system (Scheme 26).<sup>97</sup>

$$RH \xrightarrow{PtCl_6^{2^\circ}, PtCl_4^{2^\circ}cat}_{H_2O(Cl^\circ), 120^\circ C} ROH + RCI$$
(73)

$$CH_4 + H_2SO_4 \xrightarrow{(bpym)PtCl_2} CH_3OSO_3H + H_2O + SO_2$$
(74)



Muzart reported an efficient and recyclable water-soluble system to promote the allylic oxidation of olefins with organic peroxyesters in water.<sup>98</sup> The mixture of the

Scheme 26

hydrophilic nitrogen donor ligand with  $Cu(CH_3CN)_4BF_4$  in water led to an efficient and reusable catalytic system for the allylic oxidation of olefins with *tert*-butyl perbenzoate (eq 75). The aqueous phase recovered after extraction of the reaction products remained catalytically active, and oxidation of cyclohexene was repeated four times with the same aqueous phase without a drop in the yield.



Xia found that Mn(salen) complex (**76**) was effective catalysts for the oxidation of secondary alcohols to ketones in the presence of the co-oxidant diacetoxyiodobenzene (PhI(OAc)<sub>2</sub>) under aqueous condition (eq 76).<sup>99</sup> To increase the substrate and catalyst's solubility in water, the phase-transfer catalyst (N(C<sub>2</sub>H<sub>5</sub>)<sub>4</sub>Br) was included in the aqueous system and the product was formed with an high 84.1% ee in 62.5% yield.



Recently, Li and coworkers reported a efficient asymmetric aldehyde-alkyneamine coupling reaction in water (eq 77).<sup>100</sup> Use of the tridentate bis(oxazolinyl) pyridines, **77** with Cu(OTf) afforded the product with both high yield (48-93%) and enantioselectivity in organic solvent (up to 99.6% ee) and in water (up to 84% ee). In most cases, imines were formed *in situ* and the addition was very simple: mixing an aldehyde, aniline, and alkyne with the catalyst in one pot.



Fukuzumi have recently reported that the formation of novel ruthenium(IV)-oxo complex and its reactivity toward efficient and selective catalytic oxygenation and

oxidation reaction of various hydrocarbons in water, which can be used as an oxygen source.<sup>101</sup> In this system, water is not only the solvent but also the sole oxygen source in the efficient oxidation of organic compounds catalyzed by a Ru(II)-pyridylamine-aqua complex, **78** with Ce(IV) as the oxidant. An intermediate-spin Ru(IV)-oxo complex, **79** is formed as the reactive species.<sup>101</sup> This catalytic system has been shown high turnover numbers for various substrates: cyclohexane is converted into the corresponding dicarboxylic acid and the water soluble sodium 4-sulfonate-1-ethylbenzene forms the corresponding acetophenone derivative selectivity (eqs 78-79).



Doyle reported that  $Rh_2(cap)_4$ , **65** with *t*-BuOOH can effectively catalyze allylic,<sup>89</sup> benzylic oxidation.<sup>102</sup>. They have recently reported the oxidation of alkynes to  $\alpha$ , $\beta$ -acetylenic ketones by *t*-BuOOH, catalyzed by **65** with water as the reaction solvent.<sup>103</sup> The catalytic oxidation of 4-octyne by **65** with *t*-BuOOH in water afforded the

corresponding ynone with high yield (89%) and short reaction time (1 h) (eq 80). The aqueous phase recovered after extraction of the reaction products can be used to another reaction, and oxidation of other substrate was repeated three times with the same aqueous phase without decreasing catalytic activity.

$$\begin{array}{c} 65 \\ \underline{t\text{-BuOOH}} \\ H_2O, 40 \ ^{\circ}C \end{array}$$
(73)

#### **Results and Discussion**

The catalytic C-H bond oxidation of alkanes under mild conditions is of central importance in view of synthetic and industrial aspects.<sup>78</sup> Generally, catalytic oxidation of alkanes under mild conditions is quite difficult, because of the lack of the reactivity of alkanes, which stems from their high C-H bond dissociation energy. Efficient metal catalysts are required for unactivated C-H bond oxidation in organic synthesis. Recently, transition metal catalyzed oxidations have been reported; ruthenium and iron complexes with chelating nitrogen ligands have been shown to exhibit high activity and selectivity for oxidative reactions under organic and aqueous condition,<sup>87,92,101,104</sup> but selective catalytic oxidation of alkane under mild condition are relatively rare.

From environmental and economic points of view, the development of organic transformations in aqueous media has become one of the major advantages in modern chemistry.<sup>105</sup> Following the general trend, the design of novel transition-metal catalysts for organic reaction in water has arisen a growing interest in recent years, disclosing up to

date a wide variety of highly efficient and selective synthetic approaches.<sup>95</sup> Recently, Fukuzumi and co-workers has reported highly efficient and selective catalytic oxygenation and oxidation reaction by  $[Ru(tpa)(H_2O)_2](PF_6)_2$  which contain tetradentate nitrogen based ligand under aqueous condition.<sup>101</sup> Even though catalytic reaction under aqueous condition has a lot of advantages, it is difficult to use water as solvent in metal catalyzed chemical reactions because the majority of transition metal catalysts are unstable and insoluble in aqueous condition.

Since the first reports of the use of chiral oxazoline-based ligands in asymmetric catalysis,<sup>106</sup> a diverse range of oxazoline ring-based ligands with transition metals have been developed as catalysts in a wide range of chemical reactions, such as cyclopropanation, epoxidation, alcohol oxidation. Initially we thought that oxazoline ligands are suitable candidates for highly active and selective catalytic oxidation because of their readily accessibility, modular nature, and applicability in a wide range of metal catalyzed transformations.<sup>106,107</sup> We have recently developed cationic ruthenium (III) complex  $[(pymox-Me_2)_2RuCl_2]^+BF_4^-$ (80) (pymox-Me<sub>2</sub> 4,4-dimethyl-2-(2-= pyridyl)oxazoline). We have decovered that 80 was found to be efficient catalyst for C-H oxidation reaction of arylalkanes in water. In this chapter, we describe the synthesis and characterization of 80 and their applications for C-H oxidation reactions in aqueous condition.

#### **4.1** Synthesis of Water-Soluble Ruthenium Pymox Complexes

As part of an ongoing effort to develop ruthenium-catalyzed C-H bond activation reactions, we initially screened several chelating nitrogen ligands to synthesize water soluble ruthenium catalysts. Thus, the treatment of  $[(COD)RuCl_2]_x$  with 1.2 equivalents of pymox-Me<sub>2</sub> ligand in 1,2-dichloroethane at 50 °C produced an orange-yellow colored complex (pymox-Me<sub>2</sub>)Ru(COD)Cl<sub>2</sub> (**81**), which was isolated in 65% yield after recrystallization in *n*-hexanes/CH<sub>2</sub>Cl<sub>2</sub> (Scheme 27). The treatment of **81** (0.4 mmol) with pymox-Me<sub>2</sub> (1.9 mmol) in 1,2-dichloroethane at 100 °C led to the isolation of a deep blue-purple colored complex (pymox-Me<sub>2</sub>)<sub>2</sub>RuCl<sub>2</sub> (**82**) in 55% yield. Alternatively, the complex **82** could be directly produced from the treatment of  $[(COD)RuCl_2]_x$  with excess amount of pymox-Me<sub>2</sub> in 1,2-dichloroethane at 100 °C with 65% yield.

The subsequent treatment of **82** with NaBF<sub>4</sub> and *t*-BuOOH in CH<sub>2</sub>Cl<sub>2</sub> led to the cationic Ru(III) complex [(pymox-Me<sub>2</sub>)<sub>2</sub>RuCl<sub>2</sub>]<sup>+</sup>BF<sub>4</sub><sup>-</sup> (**80**) in 73% isolated yield. The structure of these ruthenium complexes was completely established by both spectroscopic and X-ray crystallographic methods. The molecular structure of both **82** and **80** showed an octahedral geometry with *cis* coordination of the chloride and anti-pyridine ligands. The average Ru-Cl bond distance of the cationic Ru(III) complex **80** (2.33 Å) was found to be considerably shorter than the neutral complex **82** (2.41 Å). The magnetic moment of **80** ( $\mu_{eff} = 1.55 \text{ B}_{M}$ ) as determined by using the Evans NMR method was also consistent with a paramagnetic Ru(III) complex.

$$(81)$$

In a strikingly different reactivity pattern, only complex **80** was found to exhibit high catalytic activity for C-H bond oxidation reaction in aqueous solution, even though both **82** and **80** are soluble in water. Thus, the treatment of ethylbenzene (1.0 mmol) with *t*-BuOOH (3 mmol, 70 wt% in aqueous solution) in the presence of 1 mol% of **80** in water (3 mL) cleanly produced acetophenone in >95% conversion within 16 h at room temperature (eq 81).

Scheme 27




Figure 27. Molecular Structure of Complex 80.



Figure 28. Molecular Structure of Complex 81.



Figure 29. Molecular Structure of Complex 82.

# 4.2 Optimization Studies for C-H Oxidation Reaction of Arylalkanes

## **4.2.1 Catalyst Survey**

Initially, we used the reaction of ethylbenznene with *t*-BuOOH as an oxidizing agent to survey the catalytic activity of ruthenium complexes. However, commonly available ruthenium complexes such as  $RuCl_2(PPh_3)_3$ ,  $RuO_2$ ,  $RuCl_3 \cdot 3H_2O$ , (*p*-cymene) $RuCl_2$  and  $Bu_4N^+RuO_4^-$  were not soluble in aqueous solution. Thus, catalytic activity could not be directly compared to the complex **80** under the similar reaction condition.

Ethylbenzene was readily oxidized to acetophenone by **80** at 20 °C in aqueous condition giving 71% yield in 5 h at 20 °C and over 95% yield in 16 h. On the other hand, oxidation of ethylbenzene with 1.0 mol%  $RuCl_2(PPh_3)_3$  (**83**) and *t*-BuOOH in benzene at

20 °C for 2 h afforded acetophenone in only 53% yield.<sup>80</sup> Che reported that the reaction was performed at 20 °C for 24 h in the presence of *cis*-[Ru(6,6'-Cl<sub>2</sub>bpy)<sub>2</sub>(OH<sub>2</sub>)<sub>2</sub>][SF<sub>3</sub>SO<sub>3</sub>]<sub>2</sub> (bpy = 2,2'-bipyridine) (**84**) (0.1 mol%) with *t*-BuOOH in acetone and obtained in 71% yield.<sup>87</sup> Doyle and coworker obtained that ethylbenzene is reacted with dirhodium caprolactamate (Rh<sub>2</sub>(cap)<sub>4</sub>) (**65**), *t*-BuOOH and NaBF<sub>4</sub> in DMF at 40 °C for 16 h and gave its corresponding product in 20% yield.<sup>102</sup> Even though our catalytic system could not be directly compared with these catalytic systems, our system was comparable to or better than the reported catalytic systems (Table 16).

	+	<i>t-</i> BuOOH (3 equiv)	[cat] H <sub>2</sub> O, 20 °C		Ĭ,
entry	cat		solvent	t (h)	yield (%)
1 <sup>a</sup>	none		H <sub>2</sub> O	5	0
2 <sup>b</sup>	80		H <sub>2</sub> O	5	71
3 <sup>c</sup>	83		$C_6H_6$	2	53
4 <sup><i>d</i></sup>	84		acetone	24	71
5 <sup>e</sup>	65		DMF	16	20

**Table 16.** Catalyst Survey for Oxidation Reaction of Ethylbenzene with *t*-BuOOH.

Reaction Conditions: <sup>*a*</sup> *t*-BuOOH (3.0 equiv). <sup>*b*</sup> **80** (1 mol%), *t*-BuOOH (3.0 equiv). <sup>*c*</sup> **83** (1 mol%), *t*-BuOOH (4.0 equiv), reference 18. <sup>*d*</sup> **84** (0.1 mol%), *t*-BuOOH (3.2 equiv), reference 25. <sup>*e*</sup> **65** (1.0 mol%), *t*-BuOOH (3.2 equiv), NaBF<sub>4</sub> (0.5 equiv), reference 31.

 $\sim$ 

# 4.2.2 Survey of Oxidizing Agents

A variety of oxidizing agents was examined in order to optimize the reaction condition. The reaction of ethylbenzene (1.0 mmol) with an oxidizing agent (3.0 mmol) in the presence of the complex **80** was performed in a variety of oxidizing agents and the yield of oxidizing products was determined by GC. It was found that the nature of oxidizing agents significantly affected the efficiency of the oxidation of arylalkanes.

	+ [ox]	80 (1 mol%) additive (3 mol%)		
	(3 equiv)	H₂O, 20 ℃, 16 h		
entry	[ox]	additive(s)	conversion <sup>b</sup> (%)	
1	02	none	9	
2	$O_2$	NaBF <sub>4</sub>	10	
3 <sup><i>c</i></sup>	$O_2$	CH <sub>3</sub> CHO	18	
4 <sup><i>c</i></sup>	$\overline{O_2}$	CH <sub>3</sub> CHO + NaBF <sub>4</sub>	ı 20	
5	NalO <sub>4</sub>	none	0	
6	NalO <sub>4</sub>	NaBF <sub>4</sub>	2	
7	<i>m</i> -CPBA	none	18	
8	<i>m</i> -CPBA	NaBF <sub>4</sub>	22	
9	$H_2O_2$	none	21	
10	$H_2O_2$	NaBF <sub>4</sub>	23	
11	CH <sub>3</sub> CO <sub>3</sub> H	none	35	
12	CH <sub>3</sub> CO <sub>3</sub> H	NaBF <sub>4</sub>	38	
13	<i>t-</i> BuOOH	none	>95	
14	<i>t-</i> BuOOH	NaBF <sub>4</sub>	>95	

Table 17. Oxidizing Agent and Additive Effects on Oxidation of Ethylbenzene.<sup>a</sup>

<sup>*a*</sup> Reaction conditions: ethylbenzene (1 mmol), [ox] (3.0 equiv), additive (3 mol%), **80** (1 mol%), H<sub>2</sub>O (3 mL), 20 °C, 16 h. <sup>*b*</sup> Determined by GC. <sup>*c*</sup> Acetaldehyde (0.1 equiv).

*t*-BuOOH was found to be the best oxidizing agent for this reaction (Table 17). As in the case of molecular oxygen, low yields were obtained (entry 1). In the presence of additives such as NaBF<sub>4</sub>, acetaldehyde which generates peracetic acid in situ with molecular oxygen, or both NaBF<sub>4</sub> and acetaldehyde, product yield was increased to a little bit higher compared with no additive (entry 2-4). Sodium periodate (NaIO<sub>4</sub>) showed no reaction with complex 80 under the similar reaction condition (entry 5-6). 3-Chloroperoxybenzoic acid (m-CPBA) only or with NaBF<sub>4</sub> as additive also led to low yields (entry 7-8). Hydrogen peroxide ( $H_2O_2$ ) gave low yields. Addition of NaBF<sub>4</sub> to H<sub>2</sub>O<sub>2</sub> reaction gave similar yield (entry 9 and 10). Peracetic acid (CH<sub>3</sub>CO<sub>3</sub>H) was found to be slightly increase activity compared to other oxidants, but in lower yield compared to t-BuOOH (entry 11-12). When t-BuOOH was used as oxidizing agent in this reaction, it exhibited highest activity with complex 80 under similar reaction condition. When  $NaBF_4$ was added as additive, product yield was not changed compared to the case of catalyst only. From these results, t-BuOOH was the best oxidizing agent with complex 80 in aqueous solution and oxidation reaction with t-BuOOH in presence of 80 was not affected from NaBF<sub>4</sub>.

## 4.2.3 Reusability of the Cationic Ruthenium Complex in Aqueous Solution

Salient features of water-soluble catalyst **80** are that it retains significant activity after repeated runs, and it can be readily separated from the reaction mixture by simple extraction. In this series of oxidations, ethylbenzene was treated under standard conditions (1.0 mol% of **80**, 3.0 mol% NaBF<sub>4</sub>, 3.0 equiv of *t*-BuOOH, in 3.0 mL water) to afford their corresponding oxidizing products in 90% yield (eq 82). Following extraction of products with diethyl ether, the aqueous layer from the first reaction was reused with additional substrate and oxidizing agent. The second and third reactions were performed with the similar reaction condition and these reaction gave comparable yields  $(2^{nd}: 71\%, 3^{rd}: 61\%)$ . Subsequently, we found that resultant aqueous layer from the second reaction was able to catalyze another oxidation. Even though the yields of second and third time were diminished, the oxidation of ethylbenzene by complex **80** in aqueous condition showed reusability of the catalyst.



# **4.3 Reaction Scope**

The scope of the oxidation of alkanes was explored in order to demonstrate the synthetic utility of complex **80**. We examined various benzylic compounds and cyclic alkanes by using these optimized conditions. In general, the C-H bond oxidation of benzylic compounds occurred smoothly at room temperature to give the ketone products (Table 18).

Ethylbenzene and its derivatives are oxidized at the benzylic C-H bonds to give acetophenone and its derivatives selectively (entry 1-3). Propylbenzene was converted to

its corresponding  $\alpha$ -position ketone **85d** with good yield (entry 4). The formation of C-C bond cleavage product for isobutylbenzene is reminiscent of the oxidation reaction promoted by transition metal complexes (entry 5), where benzyloxy radical species has been implicated for the C-C bond cleavage reactions of alkylbenzenes.<sup>10</sup> The oxidation of tertiary benzylic C-H bond is favored over the primary ones to give the alcohol products (entry 6). 2-Ethylnaphthalene produced to their corresponding ketone in higher yield than ethylbenzene and their derivatives (entry 7). Indan was oxidized with high conversion and converted selectively to their corresponding ketones, which are 1-indanone and 2,3-dihydro-3-hydroxyindan-1-one, with moderate yield in 2 h (entry 8). Fluorene and diphenylmethane can be readily converted into the corresponding ketones in high yield in short reaction time (entries 9 and 11).

In contrast, the dehydrogenation product **851** was favored over the oxidation product **85m** for the 9,10-anthracene case (entry 10). Functionalized alkylated arene such as xanthene was also oxidized with 95% yield (entry 12). The oxidation of cyclic alkenes was found to be sluggish, giving only modest conversions under the similar reaction conditions (entries 13-15).

entry	substrate	products	t (h)	convn (%)	yield (%) <sup>b</sup>
1 2 3	x	O 85a X = H 86b X = OMe 85c X = Cl	16 16 16	95 97 87	83(90) 87(95) 77(83)
4		85d	16	88	72(80)
5		85e	24	56	41(47)
6		$OH$ $OH$ $OH$ $HO_2C$ $O$ $SF:85g = (3:1)$	16	90	74(80) <sup>c</sup>
7		85h	16	94	89(90)
8		85i:85j = (8.5:	1) 2	>99	80(90) <sup>d</sup>
9 <sup>e</sup>		85k 0	2	93	86(92)
10 <sup>e</sup>			2	>99	88(95)
11		0 0 851:85fm 0 85n	= (4:1) 2	90	82(88)
12 <sup>e</sup>		850	2	>99	87(95)
13	$\bigcirc$	O OH 85p:85q = (3:1	24 )	33	(29) <sup>f</sup>
14	$\bigcirc$	O O HO 85r:85s = (1:1	24 )	31	(28) <sup>f</sup>
15	$\bigcirc$	<b>85t:85u = (15:</b>	24 1)	66	54(62)

**Table 18.** Aqueous Phase C-H Bond Oxidation of Arylalkanes.<sup>a</sup>

<sup>*a*</sup> Reaction conditions: substrate (1.0 mmol), *t*-BuOOH (3.0 mmol, 70 wt% in water), **80** (1.0 mol%), H<sub>2</sub>O (3 mL), 20-22 °C. <sup>*b*</sup> Isolated product yields. The GC product yields are listed in parenthesis. <sup>*c*</sup> Less than 5% of benzaldehyde derivative is formed. <sup>*d*</sup> Five present of 1,3-indandione is formed. <sup>*e*</sup> Substrate was dissolved in 1 mL of CH<sub>2</sub>Cl<sub>2</sub>. <sup>*f*</sup> Products were not isolated due to low conversion and difficulty in separation.

## **4.4 Mechanistic Studies**

#### 4.4.1 Isotope Effect Study

As mention in above, the  $k_{\rm H}/k_{\rm D}$  value between radical-caged and free radical mechanism showed different feature (Table 15). The radical-caged mechanism was usually obtained by higher isotope effect ( $k_{\rm H}/k_{\rm D}$ ) than free radical mechanism. But, both mechanisms indicate that the C-H bond breaking is the rate determining step. To learn more about the isotope effect of our system, the oxidation of ethylbenzene with *t*-BuOOH and ethylbenzene- $d_{10}$  with *t*-BuOOH were performed separately, and the reaction rates were determined by GC. A significant isotope effect was observed for the C-H bond oxidation reaction of ethylbenzene and ethylbenzene- $d_{10}$ . The pseudo-first order plots of the catalytic reaction of both ethylbenzene and ethylbenzene- $d_{10}$  with *t*-BuOOH, at room temperature led to a  $k_{\rm obs} = 2.1 \times 10^{-2} \, \text{h}^{-1}$  and  $k_{\rm obs} = 1.5 \times 10^{-3} \, \text{h}^{-1}$ , respectively, from which a  $k_{\rm H}/k_{\rm D} = 14 \pm 0.1$  was obtained (Figure 30). This result indicates that the cleavage of the C-H bond is the rate-limiting step in the overall reaction mechanism and this value is similar with radical caged mechanism.



*Figure 30.* First-Order Plots of  $-ln([ethylbenzene]_t/[ethylbenzene]_0)$  vs Time for C-H Bond Oxidation Reaction of Ethylbenzene ( $\blacklozenge$ ) or Ethylbenzene- $d_{10}$  ( $\blacklozenge$ ) with *t*-BuOOH.

# 4.4.2 Hammett Study

Hammett studies of *para*-substituted ethylbenzene substrates were performed to determine the electronic effects on arylalkane substrate during C-H bond oxidation reaction. The reaction rate was found to be considerably accelerated by ethylbenzene containing electron donating groups. The Hammett correlation of *para*-substituted ethylbenzene substrates *p*-X-C<sub>6</sub>H<sub>4</sub>CH<sub>2</sub>CH<sub>3</sub> (X = OMe, CH<sub>3</sub>, H, F, Cl) led to  $\rho = -1.1 \pm 0.1$  (Figure 31). The negative value of  $\rho$  indicates considerable cationic character in the transition state. The observed  $\rho$  value is substantially higher than the oxidation reaction catalyzed by free radical species such as *t*-BuO• and *t*-BuOO• ( $\rho = -0.4$  to -0.6), but

somewhat lower than the ones catalyzed by  $(PPh_3)RuCl_2/t$ -BuOOH and cytochrome P-450 and their synthetic model systems ( $\rho = -1.3$  to -1.6).<sup>12</sup> A relatively high - $\rho$  value suggested of a substantial charge transfer from a metal-oxo species to the substrate during the C-H bond cleavage step.



*Figure 31.* Hammett plot of the C-H Bond Oxidation Reaction of *para*-Substituted Ethylbenzene Substrates p-X-C<sub>6</sub>H<sub>4</sub>CH<sub>2</sub>CH<sub>3</sub> (X = OMe, CH<sub>3</sub>, H, F, Cl) with *t*-BuOOH.

# 4.4.3 Detection of Catalytically Active Species

The initially inactive neutral ruthenium complex **82** became an active catalyst upon addition of NaBF<sub>4</sub> for the oxidation reaction. This fact and a relatively low Ru(II)/Ru(III) redox potential ( $E_0 = +0.22$  V) clearly indicate that the cat

ionic Ru(III) complex **80** is catalytically active species for the oxidation reaction (Figure.30).<sup>108</sup>



*Figure 32.* Cyclic Voltammogram of **80** in CH<sub>2</sub>Cl<sub>2</sub>.

The observation of a strong metal-to-ligand charge transfer (MLCT) band at 360 nm ( $d\pi$ - $\pi^*$ ) from the reaction mixture of **82** with *t*-BuOOH and NaBF<sub>4</sub> also supports the formation of a Ru(III) speceies which gives similar metal-to-ligand charge transfer band at 360 nm ( $d\pi$ - $\pi^*$ ) (Figure 33). The data are most consistent with a "solvent-carged" oxygen rebound mechanism of rate-limiting C-H oxidation step from a Ru(IV)-oxo species.<sup>80,87,108</sup> From UV-vis spectra and relatively low redox potential, we expected that when complex **82** was added in the reaction solution with *t*-BuOOH and NaBF<sub>4</sub>, complex **80** was generated *in situ* from **82** and cationic Ru(III) complex **80** acted as catalyst precursor in oxidation of alkanes.



*Figure* **33.** UV-vis spectra of **82** (20  $\mu$ M), **82** (20  $\mu$ M)/*t*-BuOOH (30 equiv)/NaBF<sub>4</sub>, and **80** (20  $\mu$ M) in water.

# 4.5 Proposed Mechanism

Regarding ongoing controversy surrounding the reaction mechanism of Gif-type oxidations, pathways involving metal-oxo species or oxygenated free radicals have been proposed. It is difficult to differentiate these two mechanistic possibilities, partly because many reports describing the relative reactivity of discrete metal-oxo complexes toward different saturated C-H bonds. For our catalytic system, we proposed that ruthenium-oxo species **87** which is similar with Gif-type metal oxo species, generated from alkylperoxo-Ru(III) complex **86** by heterolytic cleavage of the O-O bond, is the active species for C-H bond oxidation of alkanes. This mechanism is similar with Murahashi's RuCl<sub>2</sub>(PPh<sub>3</sub>)<sub>3</sub>'s mechanism.<sup>18</sup> Another possibility is concerted mechanism without formation of radical intermediate based on proton tunneling. But, former mechanism is widely accepted in the literature.

While the detailed reaction mechanism for the oxidation of alkanes catalyzed by **80** still remains unclear at this stage, a plausible mechanism is complied as shown in Scheme 28. Initially, the reaction of Ru(II) complex **82** with *t*-BuOOH and NaBF<sub>4</sub> led to generation of catalytically active cationic Ru(III) complex **80**. We were able to isolate cationic Ru(III) complex **80** from the oxidation reaction and confirmed their molecular structure by X-ray diffraction analysis. These results indicate that cationic Ru(III) complex **80** is an active catalyst for the C-H bond oxidation reaction.

We propose that the Ru(III) complex 80 would react with *t*-BuOOH to give the alkylperoxo-Ru(III) complex 86. This is consistent with previous cases containing alkylperoxo-metals which can promote oxidation of hydrocarbons and are viable intermediate in catalytic oxidation of hydrocarbons with ROOH.<sup>18,71</sup> Alkylperoxo-Ru(III) complex 86 subsequently undergoes heterolytic cleavage of the O-O bond to give Ru(IV)-oxo species 87. The deuterium isotope effect was examined for oxidation of ethylbenzene and ethylbenzene- $d_{10}$  catalyzed by 80. The  $k_{\rm H}/k_{\rm D}$  value was determined to be  $14 \pm 0.1$  by means of GC analysis and this value indicates that C-H bond breaking is the rate-determining step. This large  $k_{\rm H}/k_{\rm D}$  value is similar to that obtained for the oxidation with cytochrome P450 and Murahashi system (Table 15). Hammett value ( $\rho = -$ 1.1) is slightly lower than the ones catalyzed by  $(PPh_3)RuCl_2/t$ -BuOOH and cytochrome P-450 and their synthetic model systems ( $\rho = -1.3$  to -1.6), but substantially higher than the oxidation reaction catalyzed by free radical species such as t-BuO• and t-BuOO• ( $\rho =$ -0.4 to -0.6) (Table 15).<sup>12</sup> A relatively high -o value suggested of a substantial charge transfer from a metal-oxo species to the substrate during the C-H bond cleavage step. These proposed mechanisms invoked metal-oxo complexes as reactive intermediates in

oxidation of hydrocarbons. Abstraction of a hydrogen atom from an alkane by the intermediate **87** would form the caged complex **88**, bearing an alkyl radical and a hydroxo-Ru(IV) intermediate **88**. The fact that a radical scavenger TEMPO (10 mol%) did not significantly affected the rate of the oxidation reaction also supports the notion of a solvent-caged mechanism (eq 83).

ROO*t*-Bu which is usually generated at free radical mechanism was not detected from oxidation of alkanes catalyzed by 80.<sup>30</sup> Transfer of the hydroxyl ligand to the caged alkyl radical would afford an alcohol and the ruthenium(II) species 89 to complete the catalytic cycle. Further oxidation of secondary alcohol thus formed gives the corresponding ketones under the reaction condition. From these data, we suggest that the active species of the 80/t-BuOOH system is oxo-ruthenium species, not free radical species.

The detailed reaction mechanism for the oxidation of alkanes catalyzed by **80** still remains to be established. Detailed mechanistic studies of these reactions study are currently underway.

Scheme 28



# **5.** Conclusions

We have successfully synthesized ruthenium complexes containing oxazoline ligands which have been fully characterized both spectroscopically and analytically. The cationic Ru(III) complex **80** was found to be an effective catalyst for the oxidation of saturated C-H bonds in aqueous condition. While high valent metal-oxo species have been invoked in both nonheme and Gif-type oxidation, catalytic C-H bond oxidation reactions mediated by well-defined cationic Ru(III) complexes have been rarely reported.<sup>13</sup> Efforts to extend the scope of oxidation reaction as well as the nature of active species are currently underway.

# Chapter V Experimental Section

**General Information.** All operations were carried out in an inert-atmosphere glove box or by using standard high vacuum and Schlenk techniques unless otherwise noted. Tetrahydrofuran, benzene, hexanes and Et<sub>2</sub>O were distilled from purple solutions of sodium and benzophenone immediately prior to use. The NMR solvents were dried from activated molecular sieves (4 Å). All organic substrates were received from commercial sources and used without further purification. The <sup>1</sup>H, <sup>2</sup>H, <sup>13</sup>C and <sup>31</sup>P NMR spectra were recorded on a Varian 300 or 400 MHz FT-NMR spectrometer. Mass spectra were recorded from a Agilent 6850 GC/MS spectrometer. The conversion of organic products was measured from a Hewlett-Packard HP 6890 GC spectrometer. FT-IR spectra were recorded on Perkin Elmer Spectrum 100. High resolution EI mass spectrum was obtained at the Center of Mass Spectrometry, Washington University, St. Louis, MO. Elemental analysis was performed at the Midwest Microlab, Indianapolis, IN.

### For Chapter 2

**Representative Procedure of the Catalytic Reaction.** In a glove box, complex **26** (15 mg, 25  $\mu$ mol), an arylamide (0.5 mmol) and an alkene (2.5 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The tube was brought out of the box, and was stirred for 5 h in an oil bath set at 80 °C. After the tube was cooled to room temperature, the solution was filtered through a short silica plug (hexanes/EtOAc = 2:1) in air, and the filtrate was analyzed by GC. Typically, the product mixture of **27** and **28** was not separable by column chromatography, and was subjected to the hydrogenation reaction to obtain isolated yield of the products. The treatment of the crude product mixture with H<sub>2</sub> (1 atm) in the presence of **26** (15 mg, 5 mol %) in chlorobenzene at 110 °C for 2 h led to the

of 29. **Synthesis** In a glove box, the tetrameric ruthenium complex  $\{[(PCy_3)(CO)RuH]_4(\mu-O)(\mu-OH)_2\}$  (4) (200 mg, 0.12 mmol) and N,N-dimethyl-2naphthamide were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The tube was brought out of the box, and HBF<sub>4</sub>·OEt<sub>2</sub> (64 µL, 0.48 mmol) was added under N<sub>2</sub> stream. The color of the solution was changed from dark red to green immediately. After stirring for 1 h at room temperature, the solvent was removed under vacuum, and the residue was crashed by adding hexanes (10 mL). Filtering the resulting solid through a fritted funnel and recrystallization from CH<sub>2</sub>Cl<sub>2</sub>/hexanes yielded the product as a light green powder (210 mg, ca. 60% yield, estimated purity by <sup>1</sup>H NMR ~80%; contained 2 other minor isomers (~20%)).

Spectroscopic data for the major isomer of **29**: <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 400 MHz)  $\delta$  7.9-7.4 (m, 7H, Ar), 3.18 and 3.02 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.0-1.2 (m, PCy<sub>3</sub>), -21.2 (br, Ru-H); <sup>13</sup>C{<sup>1</sup>H} NMR (CD<sub>2</sub>Cl<sub>2</sub>, 100.5 MHz)  $\delta$  197.6 (d,  $J_{CP}$  = 18.2 Hz, Ru-CO), 178.7 (CON(CH<sub>3</sub>)<sub>2</sub>), 134.8, 132.9, 129.6, 129.2, 128.9, 128.4, 128.0 and 123.9 (Ar), 38.9 (N(CH<sub>3</sub>)<sub>2</sub>), 38.6 (CH), 35.1 (N(CH<sub>3</sub>)<sub>2</sub>), 32.1, 30.9, 30.4, 29.9, 28.0, 26.8, 26.2 (CH<sub>2</sub>); <sup>31</sup>P{<sup>1</sup>H} NMR (CD<sub>2</sub>Cl<sub>2</sub>, 161.8 MHz)  $\delta$  74.4 (PCy<sub>3</sub>); IR (CD<sub>2</sub>Cl<sub>2</sub>) v<sub>CO</sub> = 1930, 1585 cm<sup>-1</sup>.

Synthesis of 30a. In a glove box, complex 29 (100 mg, 0.14 mmol) and cyclopentene (100 mg, 1.5 mmol) were dissolved in  $CH_2Cl_2$  (5 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The color of the solution was changed from green to orange immediately. After stirring for 1 h at room temperature, the solvent was removed under vacuum, and *n*-hexanes (10 mL) was added to the residue. The resulting solid was filtered through a fritted funnel and recrystallized

from  $CH_2Cl_2$ /hexanes to yield the product as a pale yellow powder (90 mg, 82% yield). Single crystals of **30a** suitable for X-ray crystallography were obtained from  $CH_2Cl_2/n$ -pentane solution.

For **30a**: <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 400.0 MHz)  $\delta$  8.45 (s, 1H, Ar), 8.06 (m, 1H, Ar), 7.94 (m, 2H, Ar), 7.72 (m, 2H, Ar), 5.46 (s, CC*H*CH<sub>2</sub>) 3.38 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 3.31 (m, 1H, CH<sub>2</sub>), 3.17 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 3.08 (m, 1H, CH<sub>2</sub>), 2.81 (m, 2H, CH<sub>2</sub>), 2.0-1.8 (br, 6H, PCy<sub>3</sub>), 1.82 (m, 2H, CH<sub>2</sub>), 1.8-0.6 (br, 27H, PCy<sub>3</sub>), -19.6 (d, *J*<sub>PH</sub> = 21.8 Hz, Ru-H); <sup>13</sup>C{<sup>1</sup>H} NMR (CD<sub>2</sub>Cl<sub>2</sub>, 100.5 MHz)  $\delta$  201.2 (d, *J*<sub>CP</sub> = 15.4 Hz, Ru-CO), 174.3 (CON(CH<sub>3</sub>)<sub>2</sub>), 136.4, 135.2, 132.4, 130.7, 130.5, 130.3, 129.4, 129.2, 128.3 and 117.8 (Ar), 115.5 (d, *J*<sub>CP</sub> = 5.1 Hz, Ru-C), 104.7 (d, *J*<sub>CP</sub> = 11.6 Hz, Ru-C), 42.4 (N(CH<sub>3</sub>)<sub>2</sub>), 37.6 and 37.5 (CH<sub>2</sub>), 37.3(N(CH<sub>3</sub>)<sub>2</sub>), 34.6 and 34.4 (CH), 30.4, 30.0, 28.1, 28.0, 27.9, 27.8, 26.4 and 21.0 (CH<sub>2</sub>); <sup>31</sup>P{<sup>1</sup>H} NMR (CD<sub>2</sub>Cl<sub>2</sub>, 161.8 MHz)  $\delta$  65.4 (PCy<sub>3</sub>); IR (CD<sub>2</sub>Cl<sub>2</sub>) v<sub>CO</sub> = 1948, 1600 cm<sup>-1</sup>; Anal. Calcd for C<sub>38</sub>H<sub>55</sub>BCl<sub>2</sub>F<sub>4</sub>NO<sub>2</sub>PRu: C, 53.85; H, 6.54. Found: C, 53.51; H, 6.48.

**Phosphine Inhibition Study.** In a glove box, complex **26** (15 mg, 25  $\mu$ mol), an C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> (0.5 mmol) and an alkene (2.5 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2.0 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. For phosphine inhibition study, PCy<sub>3</sub> (0 ~ 6.3 mol%) was added to the reaction mixture. The tube was brought out of the box, and was stirred for 5 h in an oil bath set at 80 °C. After the tube was cooled to room temperature, the solution was filtered through a short silica plug (hexanes/EtOAc = 2:1) in air, and the filtrate was analyzed by GC.

**Deuterium Isotope Effect Study.** In a glove box, complex **26** (17 mg, 30  $\mu$ mol), C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> (177 mg, 1.0 mmol) or C<sub>6</sub>D<sub>5</sub>CONEt<sub>2</sub> (182 mg, 1.0 mmol) and cyclopentene (0.09 g, 5 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (5.0 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. After the solution was

stirred at room temperature for 10 min, an equal amount of the solution (1.4 mL) was divided and placed in 5 different Schlenk tubes. The tubes were brought out of the box, and were stirred for 25 min in an oil bath set at 80 °C. Each reaction tube was taken out from the oil bath in 5 min intervals, and was immediately cooled in a dry ice/acetone bath. After filtering through a small silica gel column (hexanes/EtOAc = 2:1), the solution was analyzed by GC. The  $k_{obs}$  was determined from a first-order plot of ln[27h + 28h] vs time as measured by the appearance of the products 27h and 28h by GC.

**Deuterium Labeling Study.** In a glove box, complex **26** (15 mg, 25 µmol), *N*,*N*-diethyl benzamide- $d_5$  (C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub>, 91 mg, 0.50 mmol) and cyclopentene (0.17 g, 2.5 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The tube was brought out of the box, and was stirred for 1 h in an oil bath set at 80 °C. The tube was immediately cooled and was open to air. After filtering through a short silica gel column (hexanes/EtOAc = 2:1), the conversion was determined by GC. Both unreacted cyclopentene- $d_n$  and cyclopentane- $d_n$  were collected separately via a vacuum transfer. The product mixture of **27h**- $d_n$  and **28h**- $d_n$  and unreacted *N*,*N*-diethyl benzamide- $d_n$  was separated by a column chromatography on silica gel (hexanes/EtOAc), and each was analyzed by both <sup>1</sup>H and <sup>2</sup>H NMR.

**Carbon Isotope Effect Study.** In a glove box, complex **26** (0.23 g, 0.4 mmol),  $C_6H_5CONEt_2$  (1.4 g, 8.0 mmol) and cyclopentene (2.72 g, 0.04 mol) were dissolved in  $CH_2Cl_2$  (32 mL) in a 100 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The tube was brought out of the box, and was stirred in an oil bath at 80 °C for 5 h. The tube was cooled to room temperature and was open to air. After filtering through a small silica column (hexanes/EtOAc = 2:1), the conversion was determined by GC (78%, 80% and 82% conversion). Unreacted  $C_6H_5CONEt_2$  was separated by a column chromatography on silica gel (hexanes/EtOAc) for <sup>13</sup>C{<sup>1</sup>H} NMR

analysis.

The <sup>13</sup>C NMR analysis of the recovered and virgin samples of C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> was performed by following Singleton's <sup>13</sup>C NMR method (ref. 9 in the main text). The NMR sample of virgin and recovered C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> was prepared identically by dissolving C<sub>6</sub>H<sub>5</sub>CONEt<sub>2</sub> (100 mg) in DMSO- $d_8$  (0.5 mL) in a 5 mm high precision NMR tube. The <sup>13</sup>C{<sup>1</sup>H} NMR spectra were recorded with H-decoupling and 45 degree pulses. A 60 s delay between pulses was imposed to minimize  $T_1$  variations (d1 = 60 s, at = 5.0 s, np = 245098, nt = 706).

VT NMR Study for the Reaction of 26 with *N*,*N*-dimethyl-2-naphthamide. In a glove box, complex 26 (50 mg, 87 µmol) and *N*,*N*-dimethyl-2-naphthamide (17 mg, 87 µmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> (0.5 mL) in a thich wall J-Young NMR tube. The tube was allowed to equilibrate for 30 min before the NMR analysis. The sample tube was inserted into the NMR probe. The equilibrium constants were determined from the phosphorus integration of 26 and 29 by <sup>31</sup>P NMR in the temperature range 30 to 60 °C (10 °C intervals). The sample was allowed to equilibrate for 10-15 min before the data acquisition at each temperature.

#### **Characterization Data of Organic Products**

For **28a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 4H, Ar), 3.04 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.90 (m, 1H, ArCH), 2.72 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.0-1.4 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 142.8, 136.4, 128.9, 126.3, 125.7 and 125.4 (Ar), 42.3 (ArCH), 38.9 (N(CH<sub>3</sub>)<sub>2</sub>), 35.4 (CH<sub>2</sub>), 34.5 (CH<sub>2</sub>), 34.4 (N(CH<sub>3</sub>)<sub>2</sub>), 25.7 (CH<sub>2</sub>); GC-MS *m*/*z* = 217 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>19</sub>NO: C, 77.38; H, 8.81. Found: C, 77.49; H, 8.70.

For **28b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-6.9 (m, 3H, Ar), 3.08 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.91 (m, 1H, CH<sub>2</sub>C*H*CH<sub>2</sub>), 2.78 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.25 (s, 3H, CH<sub>3</sub>), 1.7-1.3 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.9 (CO), 139.6, 136.4, 135.3, 129.7, 126.3 and 125.9 (Ar), 42.1 (CH<sub>2</sub>CHCH<sub>2</sub>), 38.9 (N(CH<sub>3</sub>)<sub>2</sub>), 35.5 (CH<sub>2</sub>), 34.5 (CH<sub>2</sub>), 34.4 (N(CH<sub>3</sub>)<sub>2</sub>), 25.6 (CH<sub>2</sub>), 20.8 (CH<sub>3</sub>); GC-MS *m*/*z* = 231 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>18</sub>ClNO: C, 77.88; H, 9.15. Found: C, 78.57; H, 9.32.

For **28c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 3H, Ar), 3.03 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.85 (m, 1H, CH<sub>2</sub>CHCH<sub>2</sub>), 2.75 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.1-1.4 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.1 (CO), 141.5, 138.0, 134.4, 129.0, 128.1 and 125.4 (Ar), 41.9 (CH<sub>2</sub>CHCH<sub>2</sub>), 38.9 (N(CH<sub>3</sub>)<sub>2</sub>), 35.5 (CH<sub>2</sub>), 34.6 (N(CH<sub>3</sub>)<sub>2</sub>), 34.5 (CH<sub>2</sub>), 25.7 (CH<sub>2</sub>); GC-MS *m*/*z* = 251 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>18</sub>ClNO: C, 66.79; H, 7.21. Found: C, 67.20; H, 7.34.

For **28d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.0-6.6 (m, 3H, Ar), 3.70 (s, 3H, OCH<sub>3</sub>), 3.02 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.92 (m, 1H, ArCH), 2.74 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 1.9-1.4 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (CO), 159.8, 144.8, 129.0, 126.8, 112.0 and 110.6 (Ar), 55.0 (OCH<sub>3</sub>), 42.3 (ArCH), 38.9 (N(CH<sub>3</sub>)<sub>2</sub>), 35.0 (CH<sub>2</sub>), 34.5 (N(CH<sub>3</sub>)<sub>2</sub>), 34.4 (CH<sub>2</sub>), 25.6 (CH<sub>2</sub>); GC-MS *m*/*z* = 247 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>21</sub>NO<sub>2</sub>: C, 72.84; H, 8.56. Found: C, 72.68; H, 8.55.

For **28e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 3H, Ar), 3.02 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.91 (m, 1H, ArCH), 2.72 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.0-1.4 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.5 (CO), 145.5, 134.8, 134.6, 126.9, 126.6 and 125.8 (Ar), 42.1 (ArCH), 38.7 (N(CH<sub>3</sub>)<sub>2</sub>, 35.2 (CH<sub>2</sub>), 34.4 (N(CH<sub>3</sub>)<sub>2</sub>), 34.3 (CH<sub>2</sub>), 25.6 (CH<sub>2</sub>); GC-MS *m*/*z* = 251 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>18</sub>ClNO: C, 66.79; H, 7.21. Found: C, 66.83; H, 7.30.

For **28f**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 4H, Ar), 6.73 and 5.98 (s, NH<sub>2</sub>), 3.32 (m, 1H, CH<sub>2</sub>C*H*CH<sub>2</sub>), 2.0-1.4 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  173.6 (CO), 144.4, 136.2, 130.1, 126.7, 126.5 and 125.5 (Ar), 41.8 (CH<sub>2</sub>CHCH<sub>2</sub>), 35.3 (CH*C*H<sub>2</sub>), 25.9 (CH<sub>2</sub>); GC-MS *m*/*z* = 189 (M<sup>+</sup>); Anal. Calcd for C<sub>12</sub>H<sub>15</sub>NO: C, 76.16; H, 7.99. Found: C, 76.31; H, 7.88.

For **28g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 4H, Ar), 6.28 (d, J = 3.9 Hz, 1H, NH), 3.18 (m, 1H, ArCH), 2.71 (d, J = 4.9, Hz, NHCH<sub>3</sub>), 2.0-1.4 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.5 (CO), 144.3, 140.0, 129.7, 126.41, 126.40 and 125.4 (Ar), 41.8 (ArCH), 35.1 (CHCH<sub>2</sub>), 26.5 (NCH<sub>3</sub>), 25.8 (CH<sub>2</sub>); GC-MS m/z = 203 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>17</sub>NO: C, 76.81; H, 8.43. Found: C, 76.58; H, 8.18.

For **28h**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 4H, Ar), 3.68 (m, 1H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 3.37 (m, 1H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 3.03 (m, 2H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 2.92 (m, 1H, ArCH), 2.0-1.4 (br, 8H, CH<sub>2</sub>), 1.15 (t, J = 6.8 Hz, 3H, CH<sub>2</sub>CH<sub>3</sub>), 0.93 (t, J = 6.8 Hz, 3H, CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.9 (CO), 142.6, 136.8, 128.7, 126.1, 125.5, and 125.1 (Ar), 42.7 (N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 42.2 (ArCH), 38.4 (N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 35.5 (CH<sub>2</sub>), 34.4 (CH<sub>2</sub>), 25.8 (CH<sub>2</sub>), 25.6 (CH<sub>2</sub>), 13.8 (N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 12.6 (N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); GC-MS m/z = 245(M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>22</sub>NO: C, 78.32; H, 9.45. Found: C, 78.10; H, 9.23.

For **28i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (Ar), 6.97 (m, 1H, Ar), 6.35 (br, 1H, NH), 4.38 (m, 2H, NHC*H*<sub>2</sub>Ar), 3.17 (m, ArCH), 1.9-1.3 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.5 (CO), 144.2, 138.3, 136.8, 129.8, 128.6, 127.7, 127.4, 126.5, 126.4 and 125.4 (Ar), 43.6 (*C*H<sub>2</sub>Ar), 41.8 (ArCH), 35.1 (CH<sub>2</sub>), 25.7 (CH<sub>2</sub>); GC-MS *m*/*z* = 279 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO: C, 81.68; H, 7.58. Found: C, 81.72; H, 7.38. For **28j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.66 (s, 1H, NH), 7.4-7.0 (m, 10H, Ar), 3.30 (m, 1H, ArCH), 1.98 (br, 2H, CH<sub>2</sub>), 1.7-1.5 (br, 6H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.8 (CO), 144.8, 138.2, 137.1, 130.4, 129.2, 126.9, 126.5, 125.7, 124.5 and 119.9 (Ar), 42.0 (ArCH), 35.3 (CH<sub>2</sub>), 25.9 (CH<sub>2</sub>); GC-MS *m*/*z* = 265 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO<sub>2</sub>: C, 81.47; H, 7.22. Found: C, 81.61; H, 7.11.

For **28k**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-6.8 (m, 9H, Ar), 7.35 (br, 1H, NH), 3.73 (s, 3H, OCH<sub>3</sub>), 3.34 (m, 1H, ArCH), 2.05 (br, 2H, CH<sub>2</sub>), 1.78-1.5 (br, 6H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.7 (CO), 156.7, 144.9, 137.2, 131.3, 130.4, 127.0, 126.5, 125.8, 121.8 and 114.4 (Ar), 55.5 (OCH<sub>3</sub>), 42.0 (ArCH), 35.4 (CH<sub>2</sub>), 26.0 (CH<sub>2</sub>); GC-MS m/z = 295 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO<sub>2</sub>: C, 77.26; H, 7.17. Found: C, 77.05; H, 7.02.

For **281**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.87 (s, 1H, NH), 7.4-7.2 (m, 9H, Ar), 7.05 (m, 1H, Ar), 3.25 (m, 1H, ArCH), 1.94 (br, 2H, CH<sub>2</sub>), 1,72-1.5 (br, 6H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.1 (CO), 144.7, 136.8, 136.6, 130.4, 129.3, 129.0, 126.9, 126.4, 125.7 and 121.3 (Ar), 42.0 (ArCH), 35.2 (CH<sub>2</sub>), 25.9 (CH<sub>2</sub>); GC-MS *m*/*z* = 299 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>18</sub>ClNO: C, 72.11; H, 6.05. Found: C, 71.84; H, 6.10.

For **28m**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 3.11 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.79 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.47 (m, 1H, CH<sub>2</sub>CHCH<sub>2</sub>), 1.9-1.2 (br, 10H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 144.1, 136.1, 128.9, 126.7, 126.0 and 125.8 (Ar), 41.5 (CH<sub>2</sub>CHCH<sub>2</sub>), 39.0 (N(CH<sub>3</sub>)<sub>2</sub>), 34.8 (CH<sub>2</sub>), 34.7 (N(CH<sub>3</sub>)<sub>2</sub>), 34.0 (CH<sub>2</sub>), 27.1 (CH<sub>2</sub>), 26.9 (CH<sub>2</sub>), 26.2 (CH<sub>2</sub>); GC-MS *m*/*z* = 231 (M) <sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>21</sub>NO: C, 77.88; H, 9.15. Found: C, 77.84; H, 9.01.

For **28n**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.3-7.0 (m, 4H, Ar), 3.11 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.77 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.58 (m, 1H, CH<sub>2</sub>C*H*CH<sub>2</sub>), 1.9-1.4 (br, 12H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100

MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 146.0, 135.3, 129.1, 126.7, 125.7 and 125.6 (Ar), 43.3 (CH<sub>2</sub>CHCH<sub>2</sub>), 38.9 (N(CH<sub>3</sub>)<sub>2</sub>), 37.1 (CH<sub>2</sub>), 36.0 (CH<sub>2</sub>), 34.6 (N(CH<sub>3</sub>)<sub>2</sub>), 28.2 (CH<sub>2</sub>), 27.7 (CH<sub>2</sub>), 27.3 (CH<sub>2</sub>); GC-MS *m*/*z* = 245 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>23</sub>NO: C, 78.32; H, 9.45. Found: C, 78.10; H, 9.23.

For **280**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 10H, Ar), 6.17 (br, 1H, NH), 4.45 (d, J = 6.0 Hz, 2H, NHC $H_2$ Ar), 2.95 (m, 1H, ArCH), 1.73 (m, 2H, CH<sub>2</sub>), 1.6-1.3 (m, 10H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.5 (CO), 147.6, 138.5, 135.5, 130.0, 128.8, 128.0, 127.6, 126.9, 126.4 and 125.4 (Ar), 43.8 (CH<sub>2</sub>Ar), 42.3 (ArCH), 36.9 (CH<sub>2</sub>), 27.8 (CH<sub>2</sub>), 27.4 (CH<sub>2</sub>); GC-MS m/z = 307 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>21</sub>NO: C, 82.04; H, 8.20. Found: C, 82.01; H, 7.99.

For **28p**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 3.07 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.77 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.74 (m, 1H, ArCH), 1.8-1.5 (br, 14H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 146.4, 135.3, 128.9, 126.9, 125.6 and 125.4 (Ar), 40.0 (ArCH), 38.8 (N(CH<sub>3</sub>)<sub>2</sub>), 35.3 (CH<sub>2</sub>), 34.5 (N(CH<sub>3</sub>)<sub>2</sub>), 26.8 (CH<sub>2</sub>), 26.7 (CH<sub>2</sub>), 26.6 (CH<sub>2</sub>), 26.5 (CH<sub>2</sub>), 25.6 (CH<sub>2</sub>); GC-MS *m*/*z* = 259 (M<sup>+</sup>); Anal. Calcd for C<sub>17</sub>H<sub>25</sub>NO: C, 78.72; H, 9.71. Found: C, 78.67; H, 9.67.

For **28q**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 3.02 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.71 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.41 (br, 2H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 1.81 (m, 1H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 0.80 (d, J = 6.7 Hz, 6H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.4 (CO), 138.0, 136.6, 130.1, 128.5, 126.0, and 125.8 (Ar), 42.2 (CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 38.7 (N(CH<sub>3</sub>)<sub>2</sub>), 34.5 (N(CH<sub>3</sub>)<sub>2</sub>), 29.3 (CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 22.6 (CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>); GC-MS m/z = 205 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>19</sub>NO: C, 76.06; H, 9.33. Found: C, 75.82; H, 9.34.

For **28r**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 9H, Ar), 3.12 (br, 2H, CH<sub>2</sub>CH(CH<sub>3</sub>)Ph), 3.01 (br, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.77 (t, *J* = 6.5 Hz, 1H, CH<sub>2</sub>CH(CH<sub>3</sub>)Ph), 2.65 (br, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 1.21 (d, *J* = 6.5 Hz, 3H, CH<sub>2</sub>CHCH<sub>3</sub>(Ph)); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.0 (CO), 146.6, 137.2, 136.3, 130.2, 128.3, 128.1, 126.8, 126.0, 125.9 and 125.8 (Ar), 41.9 (CH<sub>2</sub>CHCH<sub>3</sub>(Ph)), 40.8 (CH<sub>2</sub>CHCH<sub>3</sub>(Ph)), 38.3 (N(CH<sub>3</sub>)<sub>2</sub>), 343 (N(CH<sub>3</sub>)<sub>2</sub>), 21.4 (CH<sub>3</sub>); GC-MS *m*/*z* = 267 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>21</sub>NO: C, 80.86; H, 7.92. Found: C, 80.57; H, 7.90.

For **28s**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 4H, Ar), 3.04 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.73 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.47 (br, 2H, CH<sub>2</sub>), 1.5-1.3 (br, 2H, CH<sub>2</sub>), 0.82 (s, 9H, C(CH<sub>3</sub>)<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.4 (CO), 139.6, 136.4, 129.5, 128.8, 125.9, and 125.8 (Ar), 45.7 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>2</sub>), 38.8 (N(CH<sub>3</sub>)<sub>2</sub>), 34.5 (N(CH<sub>3</sub>)<sub>2</sub>), 30.5 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 29.2 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 28.4 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 233 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>23</sub>NO: C, 77.21; H, 9.93. Found: C, 76.97; H, 10.08.

For **28t**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.6-7.3 (m, 6H, Ar), 3.04 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.68 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.47 (br, 2H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 1.86 (m, 1H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 0.81 (d, J = 6.6 Hz, 6H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.9 (CO), 135.4, 135.0, 132.9, 131.1, 128.4, 127.3, 127.0, 126.3, 125.5 and 125.2 (Ar), 42.3 (CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 38.5 (N(CH<sub>3</sub>)<sub>2</sub>), 34.3 (N(CH<sub>3</sub>)<sub>2</sub>), 28.8 (CH<sub>2</sub>), 22.4 (CH<sub>3</sub>); GC-MS *m*/*z* = 257 (M<sup>+</sup>); Anal. Calcd for C<sub>17</sub>H<sub>21</sub>NO: C, 79.96; H, 8.29. Found: C, 79.41; H, 8.11.

For **28u**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.6-7.3 (m, 6H, Ar), 2.99 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.66 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.57 (br, 2H, CH<sub>2</sub>), 1.5-1.3 (br, 2H, CH<sub>2</sub>), 0.84 (s, 9H, C(CH<sub>3</sub>)<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.0 (CO), 137.2, 135.2, 133.4, 131.1, 127.7, 127.5, 127.1, 126.4, 125.6 and 125.2 (Ar), 45.6 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 38.7 (N(CH<sub>3</sub>)<sub>2</sub>), 34.4 (N(CH<sub>3</sub>)<sub>2</sub>), 29.1 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 28.3 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 28.4 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>); GC-MS  $m/z = 283 \text{ (M}^+\text{)}$ ; Anal. Calcd for C<sub>19</sub>H<sub>25</sub>NO: C, 80.52; H, 8.89. Found: C, 79.92; H, 8.63.

For **28v**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.6-7.4 (m, 6H, Ar), 3.09 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 3.05 (m, 1H, ArCH), 2.77 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.2-1.6 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.8 (CO), 140.9, 135.7, 133.7, 131.5, 127.7, 127.6, 126.6, 125.9, 125.1 and 125.0 (Ar), 42.5 (ArCH), 39.3 (N(CH<sub>3</sub>)<sub>2</sub>), 34.9 (CH<sub>2</sub>), 34.8 (N(CH<sub>3</sub>)<sub>2</sub>), 34.7 (CH<sub>2</sub>), 25.7 (CH<sub>3</sub>); GC-MS *m*/*z* = 267 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>21</sub>NO: C, 80.86; H, 7.92. Found: C, 79.90; H, 7.90.

For **28w**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.0 (m, 10H, Ar), 6.22 (t, *J* = 5.6 Hz, 1H, NH), 4.52 (d, *J* = 6.0 Hz, 2H, NHC*H*<sub>2</sub>Ar), 2.44 (d, *J* = 7.8 Hz, 2H, C*H*<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>), 1.89 (m, 1H, CH<sub>2</sub>C*H*(CH<sub>3</sub>)<sub>2</sub>), 0.84 (d, J = 6.4 Hz, 6H, CH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.9 (CO), 138.3, 138.0, 137.8, 128.7, 128.5, 128.1, 127.6 and 127.3 (Ar), 43.8 (*C*H<sub>2</sub>Ar), 42.4 (CH<sub>2</sub>CHCH<sub>2</sub>), 29.7 (CH<sub>2</sub>), 22.6 (CH<sub>3</sub>); GC-MS *m*/*z* = 267 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>21</sub>NO: C, 80.86; H, 7.92. Found: C, 80.72; H, 8.08.

For **28x**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 10H, Ar), 6.33 (t, *J* = 4.9 Hz, 1H, NH), 4.54 (d, *J* = 5.9 Hz, 2H, NHCH<sub>2</sub>Ar), 2.71 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 1.42 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 0.94 (s, 9H, CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.1 (CO), 141.7, 138.4, 136.4, 130.2, 129.9, 128.8, 128.0, 127.6, 126.9 and 125.7 (Ar), 46.6 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>) 43.9 (CH<sub>2</sub>Ar), 30.7 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 29.3 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), 28.7 (CH<sub>2</sub>CH<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>); GC-MS *m*/*z* = 295 (M<sup>+</sup>); Anal. Calcd for C<sub>20</sub>H<sub>25</sub>NO: C, 81.31; H, 8.53. Found: C, 81.09; H, 8.26.

For **28y**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.3-7.0 (m, 4H, Ar), 3.78 (m, 1H, NCH<sub>2</sub>CH<sub>2</sub>), 3.56 (m, 1H, NCH<sub>2</sub>CH<sub>2</sub>), 3.05 (m, 2H, CH<sub>2</sub>), 2.93 (m, 1H, CH<sub>2</sub>CHCH<sub>2</sub>), 2.0-1.7 (br, 4H,

CH<sub>2</sub>), 1.64 (m, 6H, CH<sub>2</sub>), 1.39 (m, 2H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.0 (CO), 142.9, 136.5, 128.8, 126.4, 125.7, and 125.3 (Ar), 48.2 (CH<sub>2</sub>CHCH<sub>2</sub>), 42.3 (N(*C*H<sub>2</sub>)<sub>2</sub>), 35.5 (CH<sub>2</sub>), 34.9 (CH<sub>2</sub>), 28.4 (CH<sub>2</sub>), 25.9 (CH<sub>2</sub>), 25.8 (CH<sub>2</sub>), 25.7 (CH<sub>2</sub>), 24.5 (CH<sub>2</sub>); GC-MS *m*/*z* = 257 (M<sup>+</sup>); Anal. Calcd for C<sub>17</sub>H<sub>23</sub>NO: C, 79.33; H, 9.01. Found: C, 78.99; H, 9.01.

For **28z**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.4-6.3 (m, 2H, Ar), 3.73 (m, 1H, CH<sub>2</sub>C*H*CH<sub>2</sub>), 3.58 (s, 3H, NCH<sub>3</sub>), 2.36 (s, 3H, COCH<sub>3</sub>), 1.9-1.6 (br, 8H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  194.4 (CO), 142.1, 121.4, 121.0 and 110.8 (Ar), 36.2 (NCH<sub>3</sub>), 35.1 (COCH<sub>3</sub>), 30.5 (CH<sub>2</sub>), 28.9 (CH), 26.7 (CH<sub>2</sub>); GC-MS *m*/*z* = 295 (M<sup>+</sup>); Anal. Calcd for C<sub>12</sub>H<sub>17</sub>NO: C, 75.35; H, 8.96. Found: C, 75.35; H, 8.87.

## For Chapter 3

Representative Procedure of the Catalytic Reaction. In a glove box, complex 26 (10.0 mg, 17.4  $\mu$ mol), carbonyl compound (0.58 mmol) and alkene (2.90 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2.0 mL) in a 25 mL Schlenk tube equipped with a Teflon stopcock and a magnetic stirring bar. The tube was brought out of the box, and was stirred for 2 h in an oil bath which was preset at 70 °C. Reaction tube was taken out of the oil bath, and was immediately cooled in a dry ice/acetone bath. After filtering through a small silica gel column (hexanes/EtOAc = 2:1), the solution was analyzed by GC. Analytically pure products were isolated after a simple column chromatography on silica gel (hexanes/EtOAc = from 20:1 to 4:1).

**Synthesis of 56.** In a glove box, tetrameric ruthenium complex,  $\{[(PCy_3)(CO)RuH]_4(\mu_4-O)(\mu_3-OH)(\mu_2-OH)\}$  (4) (200 mg, 0.09 mmol) and (*E*)-*N*,*N*-dimethyl-3-(naphthalene-2-yl)-2-propenamide (80 mg, 3.5 mmol) were dissolved in dichloromethane (5 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar.

The tube was brought out of the box, and HBF<sub>4</sub>·OEt<sub>2</sub> (48  $\mu$ L, 0.36 mmol) was added and then cyclopentene (250 mg, 3.6 mmol) and water (66 mg, 3.6 mmol) were added under N<sub>2</sub> stream. The color of the solution was changed from dark red to orange. After stirring for 24 h at room temperature, the solvent was removed under vacuum, and the residue was crashed by adding hexanes (10 mL). Filtering the resulting solid through a fritted funnel and recrystallization from CH<sub>2</sub>Cl<sub>2</sub>/hexanes yielded the product as a orange powder (260 mg, 82% yield). Single crystals of **56** suitable for X-ray crystallography were obtained from CH<sub>2</sub>Cl<sub>2</sub>/pentane solution.

For **56**: <sup>1</sup>H NMR (CD<sub>2</sub>Cl<sub>2</sub>, 400.0 MHz)  $\delta$  8.0-7.5 (m, 7H, Ar), 3.57 (d, *J* = 18.3 Hz, CH<sub>2</sub>), 3.25 (dd, *J* = 18.3, 4.7 Hz, CH<sub>2</sub>), 3.02 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 3.01 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.68 (m, CC*H*CH<sub>2</sub>), 2.54 (s, CCH<sub>2</sub>CO), 2.51 (s, H<sub>2</sub>O), 2.37 (br m, 3H, PCy<sub>3</sub>), 2.16 (dd, *J* = 16.0, 8.0 Hz, CH<sub>2</sub>), 2.1-1.9 (m, 8H, PCy<sub>3</sub>), 1.88 (dd, *J* = 16.0, 8.0 Hz, CHCH<sub>2</sub>), 1.7-1.3 (m, 21H, PCy<sub>3</sub>), 1.27 (br, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (CD<sub>2</sub>Cl<sub>2</sub>, 100.5 MHz),  $\delta$  207.1 (d, *J*<sub>CP</sub> = 15.6 Hz, Ru-CO), 185.4 (d, *J*<sub>CP</sub> = 7.8 Hz, CON(CH<sub>3</sub>)<sub>2</sub>), 143.5 (d, *J*<sub>CP</sub> = 2.7 Hz, Ar*C*(CH<sub>2</sub>)C), 133.9, 133.7, 128.8, 128.4, 128.0, 127.6, 127.5, 127.4, 126.9 and 126.5 (Ar) 115.6 (ArCCCH), 59.3 (ArCCCH), 41.2 and 41.1 (CH<sub>2</sub>), 40.0 (N(CH<sub>3</sub>)<sub>2</sub>), 37.5 (N(CH<sub>3</sub>)<sub>2</sub>), 36.2 and 36.1 (CH), 35.1, 33.8, 29.7, 29.4, 28.9, 28.8, 28.3, 28.2, 26.8 and 22.1 (PCy<sub>3</sub>); <sup>31</sup>P{<sup>1</sup>H} NMR (CD<sub>2</sub>Cl<sub>2</sub>, 161.8 MHz)  $\delta$  37.3 (s, PCy<sub>3</sub>); IR (CD<sub>2</sub>Cl<sub>2</sub>) v<sub>CO</sub> = 1931, 1603 cm<sup>-1</sup>; Anal. Calcd for C<sub>40</sub>H<sub>59</sub>BCl<sub>2</sub>F<sub>4</sub>NO<sub>3</sub>PRu: C, 53.88; H, 6.67. Found: C, 53.58; H, 6.60.

**Kinetic Profile Experiment.** In a glove box, **26** (2 mg, 3.5  $\mu$ mol) and (*E*)-*p*-CF<sub>3</sub>-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et (28 mg, 0.12 mmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> (0.4 mL) in a thick wall J-Young NMR tube with a Teflon screw cap. The tube was cooled in a liquid nitrogen bath, and excess propene (0.6 mmol) was condensed via a vacuum line transfer. The tube was gradually warmed to room temperature. The sample was inserted into the NMR probe which was preset at 60 °C. The appearance and disappearance of the proton

signals for 44 ( $\delta$  6.55, CH), 45 ( $\delta$  2.88, CH<sub>2</sub>), and 35g ( $\delta$  3.37 CH<sub>2</sub>) were monitored by 1H NMR at 60 °C in 5 min intervals. The rate of the product formation was determined by measuring the integration of the product peaks against the disappearance of 44. By using a nonlinear regression technique (Sigmaplot Version 10), the experimental data were globally fitted to the kinetic equation. The rate constants  $k_1 = 0.042 \text{ min}^{-1}$  and  $k_2 = 0.011 \text{ min}^{-1}$  were obtained from this analysis

Derivation of the Kinetic Equation of the Conversion of 44 to 35g



The derivation for two consecutive first-order reaction was followed the method described in: Moore, J. W.; Pearson, R. G. Kinetics and Mechanism, 3<sup>rd</sup> Ed.; Wiley: New York, 1981, pp290-296.

$$d[44]/dt = -k_1[44]$$
  

$$d[45]/dt = k_1[44] - k_2[45]$$
  

$$d[35g]/dt = k_2[45]$$
  

$$[44] = [44]_0e^{-k_1t}$$
  

$$d[45]/dt = k_1[44]_0e^{-k_1t} - k_2[44]$$
  

$$[45] = [44]_0k_1(e^{-k_1t} - e^{-k_2t})/(k_2 - k_1) - [45]_0e^{-k_2t}$$
  

$$d[44]/dt + d[45]/dt + d[35g]dt = 0$$
  
Since  

$$[44] + [45] + [35g] = [44]_0$$
  

$$[35g] = [44]_0 - [44] - [45]$$

$$= [\mathbf{44}]_0(1 + (k_1 e^{-k_2 t} - k_2 e^{-k_1 t})/(k_2 - k_1)) + [\mathbf{45}]_0(1 - e^{-k_2 t}) + [\mathbf{35g}]_0$$
  
d[35g]/dt = [\mathbf{44}]\_0 k\_1 k\_2 (e^{-k\_2 t} - e^{-k\_1 t})/(k\_1 - k\_2) - [\mathbf{45}]\_0 k\_2 e^{-k\_2 t}

The experimental data was globally fitted to the kinetic equation by using non-linear regression techniques (Sigmaplot V. 10). The rate constants  $k_1 = 0.042 \text{ min}^{-1}$  and  $k_2 = 0.011 \text{ min}^{-1}$  were obtained and from this analysis.

$$d[\mathbf{35g}]/dt = [\mathbf{44}]_0 \ge 0.042 \ge 0.011 \ge (e^{-0.011t} - e^{-0.042t})/(0.042 - 0.011) - [\mathbf{45}]_0 \ge 0.011 \ge e^{-0.011t}$$
$$= [\mathbf{44}]_0 \ge 0.015 \ge (e^{-0.011t} - e^{-0.042t}) - [\mathbf{45}]_0 \ge 0.011 \ge e^{-0.011t}$$

Hammett Study: Reaction in CD<sub>2</sub>Cl<sub>2</sub>. In a glove box, *para*-substituted *p*-X-C<sub>6</sub>H<sub>4</sub>CH=CHCO<sub>2</sub>Et (X = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CO<sub>2</sub>Me, CF<sub>3</sub>) (0.12 mmol) and complex **26** (2 mg, 3.5 µmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> (0.4 mL) in a thick wall J-Young NMR tube with a Teflon screw cap. The tube was cooled in a liquid nitrogen bath, and excess propene (0.6 mmol) was condensed via a vacuum line transfer. The tube was gradually warmed to room temperature. The sample was inserted into the NMR probe which was preset at 20 °C. The rate was measured by monitoring the <sup>1</sup>H NMR in 5 min intervals. The  $k_{obs}$  was estimated from a first-order plot of  $-ln([(E)-C_6H_5CH=CHCO_2Et]_t/[(E)-C_6H_5CH=CHCO_2Et]_o)$  vs time by measuring the <sup>1</sup>H integration of starting material ( $\delta$  1.24, OCH<sub>2</sub>CH<sub>3</sub>) and product ( $\delta$  1.12, OCH<sub>2</sub>CH<sub>3</sub>) signals, which was normalized against and internal standard (solvent resonance).

**Reaction in CH<sub>2</sub>Cl<sub>2</sub>.** In a glove box, complex **26** (20 mg, 35  $\mu$ mol), (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> (122 mg, 0.7 mmol) or (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (0.7 mmol) and *para*-substituted *p*-Y-C<sub>6</sub>H<sub>4</sub>CH=CH<sub>2</sub> (X = OCH<sub>3</sub>, CH<sub>3</sub>, H, F, Cl, CF<sub>3</sub>) (3.5 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (8.0 mL) in a 25 mL Schlenk tube equipped with a Teflon

screw cap stopcock and a magnetic stirring bar. After the solution was stirred at room temperature for 10 min, an equal amount of the solution (1.0 mL) was divided and placed in 5 different Schlenk tubes. The tubes were brought out of the box, and were stirred for 150 min in an oil bath set at 50 °C. Each reaction tube was taken out from the oil bath in 30 min intervals, and was immediately cooled in a dry ice/acetone bath. After filtering through a small silica gel column (hexanes/EtOAc = 2:1), the solution was analyzed by GC with internal standard. The  $k_{obs}$  was determined from a first-order plot of  $-ln([starting material]_t/[starting material]_o)$  vs time.

**Deuterium Isotope Effect Study: Reaction in CD<sub>2</sub>Cl<sub>2</sub>.** In a glove box, complex **26** (2 mg, 3.5  $\mu$ mol) and (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (20 mg, 0.12 mmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> (0.4 mL) in a thick wall J-Young NMR tube with a Teflon screw cap. The tube was cooled in a liquid nitrogen bath, and excess ethylene and ethylene-*d*<sub>4</sub> (0.6 mmol) was condensed via a vacuum line transfer. The tube was gradually warmed to room temperature. The sample was inserted into the NMR probe which was preset at 60 °C. The rate was measured by monitoring the <sup>1</sup>H integration of starting material ( $\delta$  1. 22 OCH<sub>2</sub>CH<sub>3</sub>) and product ( $\delta$  1.16, OCH<sub>2</sub>CH<sub>3</sub>) signals in 5 min intervals, and these were normalized against and internal standard (solvent resonance). The *k*<sub>obs</sub> was estimated from a first-order plot of  $-ln([(E)-C_6H_5CH=CHCO_2Et]]_t/[(E)-C_6H_5CH=CHCO_2Et]_o)$  vs time.

**Reaction in CH<sub>2</sub>Cl<sub>2</sub>.** In a glove box, complex **26** (20 mg, 35  $\mu$ mol), (*E*)-C<sub>6</sub>H<sub>5</sub>CH=C(CH<sub>3</sub>)CONHCH<sub>3</sub> (122 mg, 0.7 mmol) or C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHCH<sub>3</sub> (122 mg, 0.7 mmol) and styrene or styrene- $d_{10}$  (0.36 g, 35 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (8.0 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a

magnetic stirring bar. After the solution was stirred at room temperature for 10 min, an equal amount of the solution (1.0 mL) was divided and placed in 5 different Schlenk tubes. The tubes were brought out of the box, and were stirred for 150 min in an oil bath set at 50 °C. Each reaction tube was taken out from the oil bath in 30 min intervals, and was immediately cooled in a dry ice/acetone bath. After filtering through a small silica gel column (hexanes/EtOAc = 2:1), the solution was analyzed by GC with internal standard. The of determined from first-order plot kobs а was  $ln([cinnamide]_t/[cinnamide]_0)$  vs time.

**Carbon Isotope Effect Study.** In a glove box, complex **26** (164 mg, 0.28 mmol), carbonyl compound (5.7 mmol) and alkene (5~10 equiv) were dissolved in  $CH_2Cl_2$  (10 mL) in two or three separate 100 mL Schlenk tubes equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The tubes were brought out of the box, and stirred for 14 h, respectively, in an oil bath which was preset at 50 °C. Unreacted carbonyl compound was collected separately after filtering through a small silica gel column (hexanes/EtOAc = 2:1), and the solution was analyzed by GC (60-82% conversion).

The <sup>13</sup>C{<sup>1</sup>H} NMR analysis of the recovered and virgin samples of carbonyl compound was performed by following Singleton's NMR method (ref. 11 in the main text). The NMR sample of virgin and recovered carbonyl compound was prepared identically by dissolving carbonyl compound (100 mg) in CDCl<sub>3</sub> (0.5 mL) in a 5 mm high precision NMR tube. The <sup>13</sup>C{<sup>1</sup>H} NMR spectra were recorded with H-decoupling and 45 degree pulses. A 60 s delay between pulses was imposed to minimize *T*<sub>1</sub> variations (d1 = 60 s, at = 5.0 s, np = 245098, nt = 704).

**Deuterium Labeling Study:** In a glove box, complex **26** (10.0 mg, 17.4 µmol) and (*E*)- $C_6D_5CD=CDCON(CH_3)_2$  (106 mg, 0.58 mmol) were dissolved in  $CH_2Cl_2$  (2.0 mL) in a 25 mL Schlenk tube equipped with a Teflon screw cap stopcock and a magnetic stirring bar. The tube was brought out of the box, and was stirred for 2 h in an oil bath set at 70 °C. The tube was immediately cooled and was open to air. After filtering through a small silica column (hexanes/EtOAc = 2:1), the conversion was determined by GC. Unreacted propylene- $d_n$  was collected separately via vacuum transfer. The product was separated by column chromatography on silica gel (hexanes/EtOAc), and each was analyzed by both <sup>1</sup>H and <sup>2</sup>H NMR.

In a glove box, complex **26** (4 mg, 6.0  $\mu$ mol), (*E*)-C<sub>6</sub>H<sub>5</sub>CD=C(CH<sub>3</sub>)CONHMe (20 mg, 0.12 mmol) and styrene (63 mg, 0.60 mmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> and CH<sub>2</sub>Cl<sub>2</sub> (0.4 mL) in a thick wall J-Young NMR tube with a Teflon screw cap. The thick wall J-Young NMR tubes were brought out of the box. The reaction mixture was monitored by both <sup>1</sup>H and <sup>2</sup>H NMR in 5 h intervals for 15 h, and these were normalized against and internal standard (solvent resonance). The deuterium incorporation of each substrate was estimated by both <sup>1</sup>H and <sup>2</sup>H NMR spectrum.

General Procedure for Empirical Rate Law Derivation. In a glove box, complex 26 (2 mg, 3.5  $\mu$ mol) and (*E*)-C<sub>6</sub>H<sub>5</sub>CH=CHCO<sub>2</sub>Et (20 mg, 0.12 mmol) were dissolved in CD<sub>2</sub>Cl<sub>2</sub> (0.4 mL) in a thick wall J-Young NMR tube with a Teflon screw cap. The tube was cooled in a liquid nitrogen bath, and excess propylene (0.6 mmol) was condensed via a vacuum line transfer. The tube was gradually warmed to room temperature. The sample

was inserted into the NMR probe which was preset at 20-40 °C. The rate was measured by monitoring the <sup>1</sup>H integration of starting material ( $\delta$  1.24, OCH<sub>2</sub>CH<sub>3</sub>) and product ( $\delta$ 1.16 OCH<sub>2</sub>CH<sub>3</sub>) signals in 5 min intervals, and these were normalized against and internal standard (solvent resonance). The  $k_{obs}$  was estimated from a first-order plot of –  $ln([(E)-C_6H_5CH=CHCO_2Et]_t/[(E)-C_6H_5CH=CHCO_2Et]_o)$  vs time.

## **Characterization Data of Organic Products**

For **35a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.1-7.0 (m, 2H, Ar), 6.9-6.8 (m, 2H, Ar), 4.07 (q, J = 7.2 Hz, OCH<sub>2</sub>CH<sub>3</sub>)), 3.98 (q, J = 7.0 Hz, ArOCH<sub>2</sub>CH<sub>3</sub>)), 3.34 (s, CH<sub>2</sub>), 1.82 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.62 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.38 (t, J = 7.0 Hz, ArOCH<sub>2</sub>CH<sub>3</sub>)), 1.16 (t, J = 7.2 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.9 (CO), 155.4 (Ar), 133.4 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 130.0 and 127.9 (Ar), 125.4 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 111.9 (Ar), 61.9 (ArOCH<sub>2</sub>CH<sub>3</sub>)), 58.4 (OCH<sub>2</sub>CH<sub>3</sub>), 38.3 (CH<sub>2</sub>), 20.3 and 18.7 (=C(CH<sub>3</sub>)<sub>2</sub>), 12.9 (ArOCH<sub>2</sub>CH<sub>3</sub>), 12.2 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 262 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>22</sub>O<sub>3</sub>: C, 73.25; H, 8.45. Found: C, 73.40; H, 8.33.

For **35b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.1-7.0 (m, 4H, Ar), 4.07 (q, J = 7.3 Hz, OCH<sub>2</sub>CH<sub>3</sub>)), 3.37 (s, CH<sub>2</sub>), 2.33 (s, ArCH<sub>3</sub>), 1.85 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.65 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.18 (t, J = 7.2 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.8 (CO), 140.2 (=C(CH<sub>3</sub>)<sub>2</sub>), 135.7, 132.1 and 128.7 (Ar), 127.5 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 60.4 (OCH<sub>2</sub>CH<sub>3</sub>), 40.3 (CH<sub>2</sub>), 22.3 (ArCH<sub>3</sub>), 21.1 and 20.7 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 232 (M<sup>+</sup>); The <sup>1</sup>H and <sup>13</sup>C NMR spectral data are in good agreement with the literature data.<sup>109</sup>
For **35c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 4H, Ar), 4.07 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.39 (s, 2H, CH<sub>2</sub>), 1.85 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.64 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.16 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.2 (CO), 143.2 (=C(CH<sub>3</sub>)<sub>2</sub>), 132.1, 128.8 and 127.9 (Ar), 127.8 (C=C(CH<sub>3</sub>)<sub>2</sub>), 126.2 (Ar), 60.3 (OCH<sub>2</sub>CH<sub>3</sub>), 40.1 (CH<sub>2</sub>), 22.2 and 20.6 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.1 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 218 (M<sup>+</sup>); The <sup>1</sup>H and <sup>13</sup>C NMR spectral data are in good agreement with the literature data.<sup>109</sup>

For **35d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 4.06 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>)), 3.33 (s, 2H, CH<sub>2</sub>), 1.81 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.58 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.14 (t, J = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  162.5 (CO), 160.1 (Ar), 139.0 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 132.8 and 130.4 (Ar), 126.8 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 114.9 (Ar), 60.4 (OCH<sub>2</sub>CH<sub>3</sub>), 40.1 (CH<sub>2</sub>), 22.1 and 20.6 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.1 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 252 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>17</sub>FO<sub>2</sub>: C, 71.16; H, 7.25. Found: C, 70.99; H, 7.21.

For **35e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 4.06 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.34 (s, 2H, CH<sub>2</sub>), 1.83 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.61 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.18 (t, J = 7.0 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (CO), 141.7 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 133.2, 132.1, 130.4 and 128.3 (Ar), 126.8 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 60.6 (OCH<sub>2</sub>CH<sub>3</sub>), 40.1 (CH<sub>2</sub>), 22.3 and 20.8 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 236 (M<sup>+</sup>); The <sup>1</sup>H and <sup>13</sup>C NMR spectral data are in good agreement with the literature data.<sup>109</sup>

For **35f**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.9-7.2 (m, 4H, Ar), 3.98 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>)), 3.30 (s, 2H, CH<sub>2</sub>), 1.77 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.54 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.10 (t, J = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.2 and 166.8 (CO), 148.1 (=C(CH<sub>3</sub>)<sub>2</sub>), 133.4, 129.3, 128.9 and 128.0 (Ar), 127.0 (C=C(CH<sub>3</sub>)<sub>2</sub>), 60.4 (OCH<sub>2</sub>CH<sub>3</sub>), 51.8 (OCH<sub>3</sub>), 39.7 (CH<sub>2</sub>), 22.1 and 20.7 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.0 (OCH<sub>2</sub>CH<sub>3</sub>); GC-

MS  $m/z = 276 \text{ (M}^+\text{)}$ ; Anal. Calcd for C<sub>16</sub>H<sub>20</sub>O<sub>4</sub>: C, 69.54; H, 7.30. Found: C, 70.01; H, 7.44.

For **35g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.6-7.5 (m, 2H, Ar), 7.3-7.2 (m, 2H, Ar), 4.07 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>)), 3.37 (s, 2H, CH<sub>2</sub>), 1.86 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.62 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.17 (t, J = 7.0 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (CO), 147.2 (q, J = 1.2 Hz, CCF<sub>3</sub>), 134.0 (=C(CH<sub>3</sub>)<sub>2</sub>), 129.5 (Ar), 126.9 (C=C(CH<sub>3</sub>)<sub>2</sub>), 125.2 (q, J = 3.7 Hz, CCF<sub>3</sub>), 60.7 (OCH<sub>2</sub>CH<sub>3</sub>), 40.1 (CH<sub>2</sub>), 22.4 and 20.9 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.3 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 286 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>17</sub>F<sub>3</sub>O<sub>2</sub>: C, 62.93; H, 5.99. Found: C, 63.23; H, 5.99.

For **35h**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.2 (m, 5H, Ar), 3.58 (s, 3H, OCH<sub>3</sub>), 3.41 (s, 2H, CH<sub>2</sub>), 1.85 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.64 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.9 (CO), 143.1 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 132.2, 128.7 and 127.9 (Ar), 127.6 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 126.2 (Ar), 51.4 (OCH<sub>3</sub>), 39.8 (CH<sub>2</sub>), 22.1 and 20.5 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS m/z = 204 (M<sup>+</sup>); The <sup>1</sup>H and <sup>13</sup>C NMR spectral data are in good agreement with the literature data.<sup>110</sup>

For **35i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 10H, Ar), 5.11 (s, 2H, ArC*H*<sub>2</sub>), 3.51 (s, CH<sub>2</sub>), 1.91 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.69 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 143.2 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 136.1, 132.7, 129.0, 128.6 and 128.2 (Ar), 127.7 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 126.4 (Ar), 66.4 (Ar*C*H<sub>2</sub>), 40.3 (CH<sub>2</sub>), 22.4 and 20.9 (=*C*(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 280 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>20</sub>O<sub>2</sub>: C, 81.40; H, 7.19. Found: C, 81.62; H, 7.26.

For **35j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 3.46 (s, 2H, CH<sub>2</sub>), 2.02 (s, 3H, C(O)CH<sub>3</sub>), 1.78 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.64 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  206.4 (CO), 143.2 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 132.4 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.7, 128.1 and

126.3 (Ar), 50.0 (CH<sub>2</sub>), 29.5 (C(O)*C*H<sub>3</sub>), 22.2 and 20.8 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS m/z = 188 (M<sup>+</sup>); The <sup>1</sup>H and <sup>13</sup>C NMR spectral data are in good agreement with the literature data.<sup>111</sup>

For **35k**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.9 (m, 2H, Ar), 7.5-7.2 (m, 8H, Ar), 4.12 (s, 2H, CH<sub>2</sub>), 1.87 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.74 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  197.3 (CO), 143.7 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 137.1 and 132.9 (Ar), 132.4 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.6, 128.6, 128.4 and 126.2 (Ar), 44.8 (CH<sub>2</sub>), 22.3 and 20.9 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 250 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>18</sub>O: C, 86.36; H, 7.25. Found: C, 86.22; H, 7.27.

For **351**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.1-7.0 (m, 4H, Ar), 3.48 (s, 2H, CH<sub>2</sub>), 2.32 (s, ArCH<sub>3</sub>), 2.05 (s, 3H, C(O)CH<sub>3</sub>), 1.81 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.67 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  206.9 (CO), 140.3 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 135.8, 132.3, 128.9 and 128.7 (Ar), 127.9 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 50.3 (CH<sub>2</sub>), 29.6 (C(O)CH<sub>3</sub>), 22.3 (=C(CH<sub>3</sub>)<sub>2</sub>), 21.2 (ArCH<sub>3</sub>), 20.9 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 202 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>18</sub>O: C, 83.12; H, 8.97. Found: C, 83.11; H, 8.93.

For (**Z**)-35m: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.2 (m, 4H, Ar), 6.01 (q, *J* = 7.0 Hz, 1H, =CHCH<sub>3</sub>), 4.09 (q, *J* = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>)), 3.46 (s, CH<sub>2</sub>), 1.82 (d, *J* = 6.9 Hz, =CHCH<sub>3</sub>), 1.17 (t, *J* = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>): NOESY  $\delta$  3.46  $\leftrightarrow$  1.82 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.1 (CO), 140.9 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 132.5 (Ar), 130.0 (=*C*HCH<sub>3</sub>), 129.1, 128.3 and 127.1 (Ar), 60.7 (OCH<sub>2</sub>CH<sub>3</sub>), 35.6 (CH<sub>2</sub>), 14.6 (OCH<sub>2</sub>CH<sub>3</sub>), 14.1 (=CHCH<sub>3</sub>); GC-MS *m*/*z* = 238 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>15</sub>ClO<sub>2</sub>: C, 65.41; H, 6.33. Found: C, 65.66; H, 6.57.

For (*E*)-35m: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.4-7.2 (m, 4H, Ar), 5.72 (q, *J* = 6.9 Hz, 1H, =C*H*CH<sub>3</sub>), 4.03 (q, *J* = 7.1 Hz, 2H, OC*H*<sub>2</sub>CH<sub>3</sub>)), 3.30 (s, 2H, CH<sub>2</sub>), 1.60 (d, *J* = 6.9 Hz,

3H, =CHC*H*<sub>3</sub>), 1.15 (t, *J* = 7.1 Hz, OCH<sub>2</sub>C*H*<sub>3</sub>; <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.4 (CO), 143.1 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 132.5 and 133.3 (Ar), 132.5 (=*C*HCH<sub>3</sub>), 128.2 and 127.2 (Ar), 60.6 (OCH<sub>2</sub>CH<sub>3</sub>), 44.3 (CH<sub>2</sub>), 14.9 (OCH<sub>2</sub>CH<sub>3</sub>), 14.1 (=CHCH<sub>3</sub>); GC-MS *m*/*z* = 238 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>15</sub>ClO<sub>2</sub>: C, 65.41; H, 6.33. Found: C, 65.66; H, 6.57.

For **35n**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 4H, Ar), 4.03 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.29 (s, CH<sub>2</sub>), 1.88 (q, J = 7.6 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 1.78 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 1.14 (t, J = 7.2 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 0.89 (t, J = 7.6 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (CO), 141.6 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 138.6, 130.1, 128.3 and 128.2 (Ar), 126.6 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 60.4 (OCH<sub>2</sub>CH<sub>3</sub>), 40.2 (CH<sub>2</sub>), 28.4 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 17.6 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>), 13.0 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 266 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>19</sub>ClO<sub>2</sub>: C, 67.54; H, 7.18. Found: C, 67.72; H, 6.78.

For **350**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 4.04 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.30 (s, 2H, CH<sub>2</sub>), 1.87 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.70 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.29 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.07 (t, J = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>), 0.65 (t, J = 7.2 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.4 (CO), 141.8 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 137.1, 132.1, 130.3 and 128.3 (Ar), 127.3 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 40.4 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 37.3 (CH<sub>2</sub>), 21.4 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 17.9 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>), 13.8 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 280 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>21</sub>ClO<sub>2</sub>: C, 68.44; H, 7.54. Found: C, 68.27; H, 7.38.

For **35p**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 4.00 (q, J = 7.2 Hz, OCH<sub>2</sub>CH<sub>3</sub>), 3.34 (s, CH<sub>2</sub>), 2.15 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.47 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.41 (m, , =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.05 (t, J = 7.2 Hz, 3H,

OCH<sub>2</sub>CH<sub>3</sub>), 0.94 (t, J = 7.2 Hz, 3H, CCH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ 171.6 (CO), 141.7 (=*C*(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 137.0, 132.0, 130.4 and 128.3 (Ar), 127.5 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 39.7 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 36.5 (CH<sub>2</sub>), 21.3 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 19.9 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>), 14.1 (=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 280 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>21</sub>ClO<sub>2</sub>: C, 68.44; H, 7.54. Found: C, 68.27; H, 7.38.

For **35q**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 5H, Ar), 3.92 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.83 (td, J = 10.2, 4.6 Hz, 1H, ArCH), 2.73 (dd, J = 15.0, 4.6 Hz, 1H ArCHCH<sub>2</sub>), 2.50 (dd J = 15.0, 10.5 Hz, 1H, ArCHCH<sub>2</sub>), 1.97 (m, 1H, ArCHCH), 1.90-1.14 (br, 8H, CH<sub>2</sub>), 1.04 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.2 (CO), 142.7, 131.9, 129.2 and 128.3 (Ar), 60.2 (OCH<sub>2</sub>CH<sub>3</sub>), 47.7 and 46.2 (CH), 40.7, 31. 5, 31.3, 25.3 and 24.9 (CH<sub>2</sub>), 14.1 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 280 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>21</sub>ClO<sub>2</sub>: C, 68.44; H, 7.54. Found: C, 68.50; H, 7.62.

For **35r**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 8H, Ar), 6.25 (AB,  $J_{AB} = 15.9$  Hz, 2H, CH=CHAr), 4.01 (q, J = 7.2 Hz, 2H,  $OCH_2CH_3$ ), 3.91 (q,  $J_X = 7.4$  Hz, 1H, ArCH(CH)CH<sub>2</sub>), 2.70 (t, J = 7.1 Hz, 2H, ArCH(CH)CH<sub>2</sub>), 1.11 (t, J = 7.2 Hz, 3H,  $OCH_2CH_3$ ); NOESY  $\delta$  6.25  $\leftrightarrow$  2.70 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (CO), 141.0 and 135.6 (Ar), 132.3 (CH=CHAr), 130.0 (Ar), 129.5 (CH=CHAr), 129.4, 129.2, 129.0, 128.9 and 127.7 (Ar), 60.8 (OCH<sub>2</sub>CH<sub>3</sub>), 44.6 (ArCHCH<sub>2</sub>), 40.7 (ArCHCH<sub>2</sub>), 14.4 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 349 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>18</sub>Cl<sub>2</sub>O<sub>2</sub>: C, 65.34; H, 5.19. Found: C, 65.54; H, 5.35.

For **35s**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.9-7.4 (m, 7H, Ar), 4.08 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>)), 3.51 (s, CH<sub>2</sub>), 1.93 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.71 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.18 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.9 (CO), 140.9

 $(=C(CH_3)_2)$ , 133.5 and 133.0 (Ar), 132.2 ( $C=C(CH_3)_2$ ), 128.0, 127.80, 127.77, 127.73, 127.70, 127.50, 126.0 and 125.6 (Ar), 60.6 ( $OCH_2CH_3$ ), 40.4 ( $CH_2$ ), 22.5 and 21.0 ( $=C(CH_3)_2$ ), 14.3 ( $OCH_2CH_3$ ); GC-MS m/z = 268 ( $M^+$ ); Anal. Calcd for  $C_{18}H_{20}O_2$ : C, 80.56; H, 7.51. Found: C, 80.62; H, 7.72.

For **35t**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.58 (dd, J = 2.6, 1.8 Hz, 1H, Ar), 6.10 (dd, J = 3.4, 2.6 Hz, 1H, Ar), 5.88 (dd, J = 3.4, 1.8 Hz, 1H, Ar), 4.09 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>)), 3.39 (s, 3H, NCH<sub>3</sub>), 3.35 (s, 2H, CH<sub>2</sub>), 1.87 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.60 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.22 (t, J = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.6 (CO), 137.4 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 134.4 (Ar), 120.7 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 119.5, 107.1 and 106.6 (Ar), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 40.3 (NCH<sub>3</sub>), 33.7 (CH<sub>2</sub>), 22.4 and 20.5 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 221 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>19</sub>NO<sub>3</sub>: C, 70.56; H, 8.65. Found: C, 70.85; H, 8.65.

For **35u**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.31 (q, J = 0.7 Hz, 1H, Ar), 6.35 (p, J = 1.7 Hz, 1H, Ar), 6.19 (d, J = 3.4 Hz, 1H, Ar), 4.09 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.44 (s, CH<sub>2</sub>), 1.98 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.87 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.19 (t, J = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 154.5 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 140.5 and 135.0 (Ar), 118.3 (*C*C(CH<sub>3</sub>)<sub>2</sub>), 110.8 and 108.2 (Ar), 60.4 (OCH<sub>2</sub>CH<sub>3</sub>), 36.8 (CH<sub>2</sub>), 23.0 and 22.1 (=C(CH<sub>3</sub>)<sub>2</sub>), 14.3 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS m/z = 208 (M<sup>+</sup>); Anal. Calcd for C<sub>12</sub>H<sub>16</sub>O<sub>3</sub>: C, 69.21; H, 7.74. Found: C, 68.98; H, 7.64.

For **35v**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 4.21 (m, 1H, CH), 3.92 (m, 2H, CHCH<sub>2</sub>CH<sub>2</sub>O), 2.29 (m, 1H, CHCH<sub>2</sub>), 2.03 (m, 1H, CHCH<sub>2</sub>), 1,91 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1,53 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  177.7 (CO), 140.3 (=C(CH<sub>3</sub>)<sub>2</sub>), 134.2 and 129.8 (Ar), 129.7 (C=C(CH<sub>3</sub>)<sub>2</sub>), 128.5 and 127.0 (Ar), 66.9

(CH<sub>2</sub>CO), 43.5 (CH), 27.5 (CH<sub>2</sub>), 22.8 and 20.5 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS m/z = 216 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>16</sub>O<sub>2</sub>: C, 77.75; H, 7.49. Found: C, 77.67; H, 7.53.

For **36a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 6.61 (br, 1H, NH<sub>2</sub>), 5.78 (br, 1H, NH<sub>2</sub>), 3.29 (s, 2H, CH<sub>2</sub>), 1.85 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.67 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.1 (CO), 142.7 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 133.6 (Ar), 129.0 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.7, 128.3 and 126.7 (Ar), 42.3 (CH<sub>2</sub>), 22.4 and 21.0 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 189 (M<sup>+</sup>); Anal. Calcd for C<sub>12</sub>H<sub>15</sub>NO: C, 76.16; H, 7.99. Found: C, 77.47; H, 7.78.

For **36b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 5.88 (br, 1H, N*H*CH<sub>3</sub>), 3.28 (s, 2H, CH<sub>2</sub>), 2.70 (d, *J* = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.82 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.64 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.3 (CO), 142.8 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 133.6 (Ar), 129.0 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.7, 128.3 and 126.5 (Ar), 42.6 (CH<sub>2</sub>), 26,4 (NHCH<sub>3</sub>), 22.5 and 21.0 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 203 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>17</sub>NO: C, 76.81; H, 8.43. Found: C, 76.38; H, 8.19.

For **36c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 3.37 (s, 2H, CH<sub>2</sub>), 2.79 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.76 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 1.81 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.57 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.8 (CO), 141.4 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 129.4 and 126.8 (Ar), 126.7 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 125.9 and 124.1 (Ar), 37.1 (CH<sub>2</sub>), 35.3 (N(CH<sub>3</sub>)<sub>2</sub>), 33.4 (N(CH<sub>3</sub>)<sub>2</sub>), 20.2 and 18.6 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 217 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>19</sub>NO: C, 77.38; H, 8.81. Found: C, 76.96; H, 8.76.

For **36d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 5H, Ar), 3.30 (s, 2H, CH<sub>2</sub>), 3.20 (q, *J* = 7.1 Hz, 2H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 3.03 (q, *J* = 7.1 Hz, 2H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 1.78 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.53 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 0.93 (t, *J* = 7.2 Hz, 2H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 0.87 (t, *J* = 7.1

Hz, 2H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.7 (CO), 143.0 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 130.9 (Ar), 129.0 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.7, 127.7 and 125.9 (Ar), 41.7 and 39.9 (N(*C*H<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 38.7 (CH<sub>2</sub>), 22.0 and 20.4 (=*C*(*C*H<sub>3</sub>)<sub>2</sub>), 14.0 and 12.6 (N(*C*H<sub>2</sub>*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 245 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>23</sub>NO: C, 78.32; H, 9.45. Found: C, 77.86; H, 9.56.

For **36e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 3.81 (p, *J* = 6.7 Hz, 1H, N(*CH*(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>), 3.35 (s, 2H, CH<sub>2</sub>), 3.31 (br, 1H, N(*CH*(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>), 1.84 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.58 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.27 (d, *J* = 6.8 Hz, 3H, N(CH(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>), 0.96 (d, *J* = 6.7 Hz, 3H, N(CH(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.9 (CO), 143.4 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 131.1 (Ar), 129.5 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 129.1, 128.0 and 126.2 (Ar), 48.9 (N(*C*H(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>), 45.8 (N(*C*H(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>)), 40.7 (CH<sub>2</sub>), 22.3 and 20.9 (=C(*C*H<sub>3</sub>)<sub>2</sub>), 20.8 (N(CH(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>), 20.7 (N(CH(CH<sub>3</sub>)<sub>2</sub>)<sub>2</sub>); GC-MS *m*/*z* = 273 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>27</sub>NO: C, 79.07; H, 9.95. Found: C, 79.04; H, 9.74.

For **36f**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 3.34 (s, 2H, CH<sub>2</sub>), 3.16 (dd, J = 7.6 Hz, 2H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 2.98 (dd, J = 7.7 Hz, 2H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 1.82 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.56 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.33 (m, 4H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 1.16 (m, 4H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 0.83 (t, J = 7.3, 3H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 0.81 (t, J = 7.3, 3H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 0.81 (t, J = 7.3, 3H, N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); 1<sup>3</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.3 (CO), 143.2 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 131.1 (Ar), 129.2 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.9, 127.9 and 126.1 (Ar), 47.7 and 45.8 (N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 40.7 (CH<sub>2</sub>), 31.1 and 29.7 (N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 22.1 and 20.7 (=C(CH<sub>3</sub>)<sub>2</sub>), 20.2 and 20.0 (N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 13.9 and 13.8 (N(CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 301 (M<sup>+</sup>); Anal. Calcd for C<sub>20</sub>H<sub>31</sub>NO: C, 79.68; H, 10.36. Found: C, 79.46; H, 10.11.

For **36g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.59 (br, 1H, NHAr), 7.4-6.9 (m, 9H, Ar), 3.48 (s, 2H, CH<sub>2</sub>), 2.30 (s, 3H, ArCH<sub>3</sub>), 1.94 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.75 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.9 (CO), 142.8 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 135.4 and 134.2 (Ar), 133.9 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 129.5, 128.8, 128.5, 126.8 and 120.0 (Ar), 43.7 (CH<sub>2</sub>), 22.5 (=C(CH<sub>3</sub>)<sub>2</sub>), 21.1 (ArCH<sub>3</sub>), 20.9 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 279 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO: C, 81.68; H, 7.58. Found: C, 81.23; H, 7.61.

For **36h**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.54 (br, 1H, NHPh), 7.4-7.0 (m, 10H, Ar), 3.51 (s, 2H, CH<sub>2</sub>), 1.94 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.75 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.0 (CO), 142.7 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 137.9, 134.5, 129.0 and 128.8 (Ar), 128.7 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.6, 126.9, 124.3 and 120.0 (Ar), 43.9 (CH<sub>2</sub>), 22.6 and 21.2 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 265 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO: C, 81.47; H, 7.22. Found: C, 81.70; H, 6.93.

For **36i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.69 (br, 1H, NHAr), 7.3-7.0 (m, 9H, Ar), 3.47 (s, 2H, CH<sub>2</sub>), 1.92 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.72 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.3 (CO), 142.6 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 136.5, 134.4, 129.0, 128.8 and 128.48 (Ar), 128.4 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 126.9 and 121.3 (Ar), 43.6 (CH<sub>2</sub>), 22.6 and 21.1 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 299 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>18</sub>ClNO: C, 72.11; H, 6.05. Found: C, 71.61; H, 5.95.

For **36j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 10H, Ar), 6.09 (br, 1H, NHBz), 4.38 (d, J = 5.8, 2H, NHC $H_2$ Ar), 3.39 (s, 2H, CH<sub>2</sub>), 1.88 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.67 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.1 (CO), 142.6 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 138.3, 133.8, 128.9, 128.7, 128.4 and 127.6 (Ar), 128.9 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 126.7 (Ar), 43.6 (NCH<sub>2</sub>), 42.7 (CH<sub>2</sub>), 22.5 and 21.2 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS m/z = 279 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO: C, 81.68; H, 7.58. Found: C, 81.76; H, 7.52.

For **36k**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-6.9 (m, 10H, Ar), 3.16 (s, 3H, N(CH<sub>3</sub>)Ph), 3.15 (s, 2H, CH<sub>2</sub>), 1.73 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.55 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.0 (CO), 144.1 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 143.4 and 131.3 (Ar), 129.6 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.9, 127.9, 127.3 and 126.0 (Ar), 39.6 (CH<sub>2</sub>), 37.3 (N(CH<sub>3</sub>)Ph), 22.5 and 20.6 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 279 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO: C, 81.68; H, 7.58. Found: C, 81.74; H, 7.49.

For **361**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-7.0 (m, 5H, Ar), 3.28 (t, J = 6.8 Hz, 2H, NCH<sub>2</sub>), 3.27 (s, 2H, CH<sub>2</sub>), 3.09 (t, J = 6.5 Hz, 2H, NCH<sub>2</sub>), 1.78 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.66 (m, 4H, CH<sub>2</sub>), 1.52 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.2 (CO), 143.3 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 131.2 and 128.6 (Ar), 128.4 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 127.7 and 125.9 (Ar), 46.3 (NCH<sub>2</sub>), 45.4 (NCH<sub>2</sub>), 40.4, 25.9 and 24.1 (CH<sub>2</sub>), 21.9 and 20.5 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 243 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>21</sub>NO: C, 78.97; H, 8.70. Found: C, 78.72; H, 8.58.

For **36m**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 3.42 (dd, J = 5.6 Hz, 2H, NCH<sub>2</sub>), 3.39 (s, 2H, CH<sub>2</sub>), 3.17 (dd, J = 5.6 Hz, 2H, NCH<sub>2</sub>), 1.84 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.58 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (m, 2H, CH<sub>2</sub>), 1.40 (m, 2H, CH<sub>2</sub>), 1.27 (m, 2H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.3 (CO), 143.3 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 131.4 and 129.0(Ar), 128.9 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 128.0 and 126.2 (Ar), 46.8 and 42.7 (NCH<sub>2</sub>), 38.9, 26.2, 25.6 and 24.5 (CH<sub>2</sub>), 22.3 and 20.7 (=C(CH<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 257 (M<sup>+</sup>); Anal. Calcd for C<sub>17</sub>H<sub>23</sub>NO: C, 79.33; H, 9.01. Found: C, 79.33; H, 9.14.

For (**Z**)-36n: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 6.11 (q, J = 7.0 Hz, 1H, =CHCH<sub>3</sub>), 6.09 (s, 1H, NHCH<sub>3</sub>), 3.39 (s, 2H, CH<sub>2</sub>), 2.67 (d, J = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.79 (d, J = 7.0 Hz, 3H, =CHCH<sub>3</sub>): NOESY  $\delta$  3.39  $\leftrightarrow$  1.79 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.7 (CO), 141.5 (Ar), 134.2 (*C*=CHCH<sub>3</sub>), 128.4 and 127.2 (Ar), 127.1 (C=*C*HCH<sub>3</sub>), 125.4 (Ar), 37.6 (CH<sub>2</sub>), 26.3 (NHCH<sub>3</sub>), 14.5 (C=CH*C*H<sub>3</sub>); GC-MS m/z = 189 (M<sup>+</sup>); Anal. Calcd for C<sub>12</sub>H<sub>15</sub>NO: C, 76.16; H, 7.99. Found: C, 75.99; H, 7.92.

For (*E*)-36n: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 5.88 (s, 1H, N*H*CH<sub>3</sub>), 5.75 (qt, *J* = 5.3, 0.5 Hz, 1H, =C*H*CH<sub>3</sub>), 3.25 (s, CH<sub>2</sub>), 2.71 (d, *J* = 4.9 Hz, NHCH<sub>3</sub>), 1.65 (dt, *J* = 6.9, 1.0 Hz, 3H, =CHCH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 139.4 (Ar), 135.5 (*C*=CHCH<sub>3</sub>), 128.5, 128.4 and 127.3 (Ar), 127.2 (C=CHCH<sub>3</sub>), 47.1 (CH<sub>2</sub>), 26.5 (NHCH<sub>3</sub>), 15.2 (C=CHCH<sub>3</sub>); GC-MS *m*/*z* = 189 (M<sup>+</sup>); Anal. Calcd for C<sub>12</sub>H<sub>15</sub>NO: C, 76.16; H, 7.99. Found: C, 75.99; H, 7.92.

For **360**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 5.82 (br, 1H, NHCH<sub>3</sub>), 3.26 (s, 2H, CH<sub>2</sub>), 2.71 (d, J = 3.8 Hz, 3H, NHCH<sub>3</sub>), 1.96 (q, J = 7.5 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 1.82 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 0.94 (t, J = 7.4 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>); NOESY  $\delta$  3.26  $\leftrightarrow$  1.81 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.2 (CO), 142.9 (Ar), 139.1 (C=C(CH<sub>3</sub>)C<sub>2</sub>H<sub>5</sub>), 128.5, 128.4 and 126.6 (Ar), 43.0 (CH<sub>2</sub>), 28.6 (NHCH<sub>3</sub>), 26.5 (CH<sub>2</sub>), 17.9 and 13.3 (CH<sub>3</sub>); GC-MS m/z = 217 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>19</sub>NO: C, 77.38; H, 8.81. Found: C, 76.91; H, 8.76.

For **36p**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 5.82 (br, 1H, N*H*CH<sub>3</sub>), 3.34 (s, 2H, CH<sub>2</sub>), 2.79 (d, J = 4.9 Hz, 3H, NHCH<sub>3</sub>), 2.04 (dd, J = 7.3 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.88 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.41 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.19 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 0.82 (t, J = 7.3 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>); NOESY  $\delta$  3.34  $\leftrightarrow$  1.87 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.3 (CO), 143.0 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 138.1 (Ar), 129.5 (=*C*(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 128.6, 128.4 and 126.4 (Ar), 43.1, 35.3 and

30.8 (CH<sub>2</sub>), 26.6 (NHCH<sub>3</sub>), 22.7 (CH<sub>2</sub>), 18.4 and 14.1 (CH<sub>3</sub>); GC-MS m/z = 245; Anal. Calcd for C<sub>16</sub>H<sub>23</sub>NO: C, 78.32; H, 9.45. Found: C, 78.30; H, 9.30.

For **36q**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 5.88 (br, 1H, NHCH<sub>3</sub>), 3.25 (s, 2H, CH<sub>2</sub>), 2.70 (d, J = 4.8 Hz, 3H, NHCH<sub>3</sub>), 1.95 (dd, J = 7.7 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 1.80 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 1.33 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 1.21 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 1.09 (m, 4H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 0.79 (t, J = 7.0 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>); NOESY  $\delta$  3.25  $\leftrightarrow$  1.80 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.2 (CO), 142.9 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 137.8 (Ar), 129.4 (=*C*(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>), 128.5, 128.3 and 126.5 (Ar), 42.9, 35.4, 31.6, 29.2 and 28.3 (CH<sub>2</sub>), 26.4 (NHCH<sub>3</sub>), 22.6 (CH<sub>2</sub>), 18.3 and 14.1 (CH<sub>3</sub>); GC-MS *m*/*z* = 273; Anal. Calcd for C<sub>16</sub>H<sub>23</sub>NO: C, 79.07; H, 9.95. Found: C, 78.37; H, 9.87.

For **36r**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 5.88 (br, 1H, N*H*CH3), 3.24 (s, 2H, CH<sub>2</sub>), 2.68 (d, *J* = 4.8 Hz, 3H, CH<sub>3</sub>), 1.87 (d, *J* = 7.3 Hz, 2H, CH<sub>2</sub>), 1.78 (s, 3H, CH<sub>3</sub>), 1.6-1.5 (br, 5H, CH<sub>2</sub>), 1.42 (m, 1H, CH), 1.10 (m, 2H, CH<sub>2</sub>), 1.07 (m, 1H, CH<sub>2</sub>), 0.62 (m, 2H, CH<sub>2</sub>); NOESY  $\delta$  3.24  $\leftrightarrow$  1.78 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.2 (CO), 143.1 (*C*=C(CH<sub>3</sub>)C<sub>7</sub>H<sub>13</sub>), 136.0 (Ar), 130.4 (=*C*(CH<sub>3</sub>)C<sub>7</sub>H<sub>13</sub>), 128.9, 128.2 and 126.4 (Ar), 43.1 and 42.8 (CH<sub>2</sub>), 36.1 (CH), 33.1 and 26.4 and 26.3 (CH<sub>2</sub>), 18.3 (CH<sub>3</sub>); GC-MS *m*/*z*: 285; Anal. Calcd for C<sub>19</sub>H<sub>27</sub>NO: C, 79.95; H, 9.53. Found: C, 79.80; H, 9.30.

For **36s**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-6.9 (m, 10H, Ar), 5.64 (br, 1H, NHCH<sub>3</sub>), 3.23 (s, 2H, CH<sub>2</sub>), 2.71 (t, J = 7.4 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 2.62 (d, J = 4.8 Hz, 3H, NHCH<sub>3</sub>), 2.36 (t, J = 7.4 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 1.87 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph); NOESY  $\delta$  3.23  $\leftrightarrow$  1.87 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H}

NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.8 (CO), 142.7 and 141.4 (Ar), 136.1 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 131.0 (=*C*(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 128.5, 128.3, 128.2, 128.1, 126.6 and 126.0 (Ar), 43.0, 36.5 and 34.0 (CH<sub>2</sub>), 26.4 (NHCH<sub>3</sub>), 18.0 (CH<sub>3</sub>); GC-MS *m/z*: 293; Anal. Calcd for C<sub>16</sub>H<sub>23</sub>NO: C, 81.87; H, 7.90. Found: C, 81.65; H, 7.70.

For **36t**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 6.71 (br s, N*H*CH<sub>3</sub>), 5.67 (m, CH), 5.27 (m, 1H, CH), 3.02 (m, 2H, CH<sub>2</sub>), 2.58 (dd, *J* = 13.9, 4.0 Hz, 1H, CHC*H*<sub>2</sub>), 2.46 (d, *J* = 5.1 Hz, 3H, NHC*H*<sub>3</sub>), 2.39 (dd, *J* = 14.0, 9.1 Hz, 1H, CHC*H*<sub>2</sub>), 2.17 (m, CH<sub>2</sub>), 2.09 (m, 1H, CH), 1.63 (m, 1H, CH); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.7 (CO), 143.6 (Ar), 133.0 and 131.7 (CH), 128.4, 128.0 and 126.5 (Ar), 51.5 (CH), 48.2 (CH), 42.1 (CHCH<sub>2</sub>), 32.3 (CH<sub>2</sub>), 26.2 (NHCH<sub>3</sub>); HRMS (*m*/*z*): calcd for C<sub>15</sub>H<sub>19</sub>NO (M+H)<sup>+</sup>, 230.1539; Found: 230.1504.

For **36u**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 5.4 (t, *J* = 0.6 Hz, 1H, CC(Ph)=CH<sub>2</sub>), 5.2 (br, 1H, NHCH<sub>3</sub>), 5.1 (dd, *J* = 0.7 Hz, *J* = 1.4 Hz, 1H, CC(Ph)=CH<sub>2</sub>), 4.4 (t, *J* = 7.8 Hz, ArCH), 2.7 (dd, *J* = 6.5 Hz, *J* = 14.1 Hz, 1H, ArCHCH<sub>2</sub>), 2.6 (d, *J* = 4.8 Hz, NHCH<sub>3</sub>), 2.5 (dd, *J* = 8.6 Hz, *J* = 14.1 Hz, 1H, ArCHCH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.8 (CO), 151.0 (CHC(Ph)=CH<sub>2</sub>), 143.1, 137.3, 128.9, 128.6, 127.8 and 126.4 (Ar), 113.4 (CHC(Ph)=CH<sub>2</sub>), 45.5 (ArCHCH<sub>2</sub>), 43.5 (ArCHCH<sub>2</sub>), 26.5 (NHCH<sub>3</sub>); GC-MS *m*/*z* = 265 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO: C, 81.47; H, 7.22. Found: C, 81.84; H, 7.42.

For **36v**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 6.3 (AB,  $J_{AB} = 16.0$  Hz, 2H, ArC*H*=C*H*Ph), 5.3 (br, 1H, N*H*CH<sub>3</sub>), 4.0 (q,  $J_X = 7.3$  Hz, ArC*H*CH<sub>2</sub>), 2.6 (dd, J = 7.7 Hz, J = 14.1 Hz, 1H, ArCHC*H*<sub>2</sub>), 2.6 (d, J = 4.8 Hz, NHC*H*<sub>3</sub>), 2.5 (dd, J = 7.5 Hz, J = 14.0 Hz, 1H, ArCHC*H*<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.1 (CO), 142.2, 141.8, 132.4, 130.4, 128.4 and 128.1 (Ar), 127.6 (ArCH=CHPh), 127.1 and 127.0 (Ar), 126.8

(ArCHCHPh) 46.7 (ArCHCH<sub>2</sub>), 43.0 (ArCHCH<sub>2</sub>), 26.5 (NHCH<sub>3</sub>); GC-MS m/z = 265 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>19</sub>NO: C, 81.47; H, 7.22. Found: C, 81.84; H, 7.42.

For **36w**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.2 (m, 5H, Ar), 6.4 (br, 1H, N*H*CH<sub>3</sub>), 6.2 (t, J = 7.7 Hz, C=C*H*CH<sub>2</sub>), 4.2 (q, J = 7.2 Hz, 2H, CO<sub>2</sub>C*H*<sub>2</sub>CH<sub>3</sub>), 3.5 (s, 2H, CH<sub>2</sub>), 3.3 (d, J = 7.7 Hz, 2H, C=CHC*H*<sub>2</sub>), 2.7 (d, J = 4.8 Hz, 3H, NHC*H*<sub>3</sub>), 1.3 (t, J = 7.2, 3H, CO<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.0 and 170.2 (CO), 140.8 (Ar), 137.6 (C=CHCH<sub>2</sub>), 128.8 (Ar), 128.0 (C=CHCH<sub>2</sub>), 126.1 and 123.6 (Ar), 61.5 (CO<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 38.6 (C=CHCH<sub>2</sub>), 34.6 (ArCCH<sub>2</sub>), 26.6 (NHCH<sub>3</sub>), 14.4 (CO<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 261 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>19</sub>NO<sub>3</sub>: C, 68.94; H, 7.33. Found: C, 68.85; H, 7.21.

For **36x**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.3 (m, 7H, Ar), 3.48 (s, 2H, CH<sub>2</sub>), 2.82 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 2.77 (s, 3H, N(CH<sub>3</sub>)<sub>2</sub>), 1.90 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.67 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  170.8 (CO), 140.9 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 133.3 and 132.0 (Ar), 128.6 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 127.8, 127.7, 127.5, 127.4, 127.2, 125,7 and 125.4 (Ar), 39.1 (CH<sub>2</sub>), 37.3 (N(CH<sub>3</sub>)<sub>2</sub>), 35.4 (N(CH<sub>3</sub>)<sub>2</sub>), 22.3 and 20.7 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 267 (M<sup>+</sup>); Anal. Calcd for C<sub>18</sub>H<sub>21</sub>NO: C, 80.86; H, 7.92. Found: C, 80.55; H, 7.86.

For **37a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.1-6.9 (m, 4H, Ar), 4.08 (m, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.80 (q, *J* = 7.1 Hz, 1H, CHCH<sub>3</sub>), 2.34 (s, 3H, ArCH<sub>3</sub>) 1.88 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.51 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.24 (t, *J* = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>) 1.16 (d, *J* = 7.1 Hz, 3H, CHCH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.8 (CO), 137.9 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 135.8 and 133.9 (Ar), 130.9 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 129.5 and 128.7 (Ar), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 42.8 (CH), 22.8 (ArCH<sub>3</sub>), 21.3 and 20.3 (=C(CH<sub>3</sub>)<sub>2</sub>), 15.9 (CH<sub>3</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 246 (M<sup>+</sup>); Anal. Calcd for C<sub>16</sub>H<sub>22</sub>O<sub>2</sub>: C, 78.01; H, 9.00. Found: C, 77.75; H, 8.94. For **37b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 4H, Ar), 4.07 (m, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.80 (q, *J* = 7.1 Hz, 1H, CHCH<sub>3</sub>), 1.87 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.20 (t, *J* = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>), 1.15 (d, *J* = 7.1 Hz, 3H, CHCH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.7 (CO), 141.0 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 134.0 (Ar), 131.0 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 129.7, 127.9 and 126.5 (Ar), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 42.8 (CH), 22.7 and 20.3 (=C(CH<sub>3</sub>)<sub>2</sub>), 15.9 (CH<sub>3</sub>), 14.1 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 232 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>20</sub>O<sub>2</sub>: C, 77.55; H, 8.68. Found: C, 77.09; H, 8.62.

For **37c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-6.9 (m, 4H, Ar), 4.05 (m, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.77 (q, *J* = 7.1 Hz, 1H, CHCH<sub>3</sub>), 1.84 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.45 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.18 (t, *J* = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>) 1.09 (d, *J* = 7.1 Hz, 3H, CHCH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.5 (CO), 139.3 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 132.8 (Ar), 132.4 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 131.9, 131.1 and 128.1 (Ar), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 42.5 (CH), 22.7 and 20.3 (=C(CH<sub>3</sub>)<sub>2</sub>), 15.9 (CH<sub>3</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 252 (M<sup>+</sup>); Anal. Calcd for C<sub>15</sub>H<sub>19</sub>ClO<sub>2</sub>: C, 67.54; H, 7.18. Found: C, 67.64; H, 7.20.

For **37d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-6.9 (m, 5H, Ar), 5.88 (br, 1H, N*H*CH<sub>3</sub>), 3.62 (q, *J* = 7.2 Hz, 1H, C*H*CH<sub>3</sub>), 2.66 (d, *J* = 4.8 Hz, 3H, NHCH<sub>3</sub>), 1.79 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.43 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.01 (d, *J* = 7.2 Hz, 3H, CHCH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.4 (CO), 140.5 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 135.1 (Ar), 131.2 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 129.2, 127.7 and 126.2 (Ar), 43.2 (*C*HCH<sub>3</sub>), 26,2 (NHCH<sub>3</sub>), 22.6 and 21.2 (=C(*C*H<sub>3</sub>)<sub>2</sub>), 15.4 (CH<sub>3</sub>); GC-MS *m*/*z* = 217 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>19</sub>NO: C, 76.38; H, 8.81. Found: C, 77.17; H, 8.73.

For **37e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-6.8 (m, 15H, Ar), 6.01 (t, J = 5.8 Hz, 1H, NH), 4.89 (s, 1H, CHPh), 4.13 (dd, J = 6.0, 5.2 Hz, 2H, NHCH<sub>2</sub>Ar), 1.71 (s, 3H,

=C(CH<sub>3</sub>)<sub>2</sub>), 1.43 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.1 (CO), 141.5 (=C(CH<sub>3</sub>)<sub>2</sub>), 138.3, 138.2 and 133.5 (Ar), 133.0 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 130.1, 129.6, 128.6, 128.4, 127.8, 127.7, 127.0 and 126.3 (Ar), 57.5 (*C*HPh), 43.7 (NCH<sub>2</sub>), 23.2 and 21.1 (=C(*C*H<sub>3</sub>)<sub>2</sub>); GC-MS *m*/*z* = 355 (M<sup>+</sup>); calcd for C<sub>25</sub>H<sub>25</sub>NO (M+H)<sup>+</sup>, 356.4851; Found: 356.4864.

For **37f**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.47 (br, 1H, NHPh), 7.4-6.9 (m, 10H, Ar), 3.79 (q, *J* = 7.0 Hz, 1H, CHCH<sub>3</sub>), 1.90 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.53 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.10 (d, *J* = 7.0 Hz, 3H, CHCH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  173.6 (CO), 140.5 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 138.5 and 135.0 (Ar), 131.7 (*C*=C(CH<sub>3</sub>)<sub>2</sub>), 129.4, 128.6, 127.9, 127.8, 127.3 and 126.4 (Ar), 43.6 (CHCH<sub>3</sub>), 22.7 and 20.4 (=C(CH<sub>3</sub>)<sub>2</sub>), 15.6 (CH<sub>3</sub>); GC-MS *m*/*z* = 279 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO: C, 81.68; H, 7.58. Found: C, 81.97; H, 7.71.

For **37g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) 7.2-6.8 (m, 10H, Ar), 6.15 (t, J = 5.5 Hz, 1H, NHBz), 4.27 (dd, J = 5.9, 2.6 Hz, 2H, NHC $H_2$ Ar), 3.62 (q, J = 7.2 Hz, 1H, CHCH<sub>3</sub>), 1.75 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.39 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.01 (d, J = 7.2 Hz, 3H, CHC $H_3$ ); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  173.7 (CO), 140.6 (=C(CH<sub>3</sub>)<sub>2</sub>), 138.6 and 135.1 (Ar), 132.0 (C=C(CH<sub>3</sub>)<sub>2</sub>), 129.4, 128.6, 127.9, 127.8, 127.3 and 126.4 (Ar), 43.6 (CHCH<sub>3</sub>), 22.7 and 20.4 (=C(CH<sub>3</sub>)<sub>2</sub>), 15.6 (CH<sub>3</sub>); GC-MS m/z = 279 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>21</sub>NO: C, 81.68; H, 7.58. Found: C, 81.97; H, 7.71.

For (**Z**)-37h: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 5H, Ar), 5.99 (s, 1H, NHCH<sub>3</sub>), 5.81 (q, *J* = 6.8 Hz, 1H, =CHCH<sub>3</sub>), 3.31 (q, *J* = 7.0 Hz, 1H, CHCH<sub>3</sub>), 2.71 (d, *J* = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.62 (d, *J* = 6.8 Hz, 3H, =CHCH<sub>3</sub>), 1.32 (d, *J* = 7.0 Hz, 3H, CHCH<sub>3</sub>): NOESY  $\delta$  3.31  $\leftrightarrow$  1.62 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.7 (CO), 141.7 (Ar), 139.6 (*C*=CHCH<sub>3</sub>), 128.7 and 127.3 (Ar), 126.9 (=CHCH<sub>3</sub>), 124.0 (Ar), 49.3 (CHCH<sub>3</sub>), 26.4 (NHCH<sub>3</sub>), 16.4 (=CHCH<sub>3</sub>), 15.0 (CHCH<sub>3</sub>); GC-MS m/z= 203 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>17</sub>NO: C, 76.81; H, 8.43. Found: C, 76.76; H, 8.27.

For (*E*)-37h: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 5H, Ar), 5.86 (s, 1H, NHCH<sub>3</sub>), 5.83 (q, *J* = 6.8 Hz, 1H, =CHCH<sub>3</sub>), 3.30 (q, *J* = 7.0 Hz, 1H, CHCH<sub>3</sub>), 2.78 (d, *J* = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.63 (d, *J* = 6.8 Hz, 3H, =CHCH<sub>3</sub>), 1.29 (d, *J* = 7.0 Hz, 3H, CHCH<sub>3</sub>): NOESY  $\delta$  5.83  $\leftrightarrow$  1.63 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.7 (CO), 141.8 (Ar), 139.6 (*C*=CHCH<sub>3</sub>), 128.8 and 127.3 (Ar), 127.0 (=CHCH<sub>3</sub>), 124.1 (Ar), 49.4 (CHCH<sub>3</sub>), 26.5 (NHCH<sub>3</sub>), 16.4 (=CHCH<sub>3</sub>), 15.0 (CHCH<sub>3</sub>); GC-MS *m*/*z* = 203 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>17</sub>NO: C, 76.81; H, 8.43. Found: C, 76.60; H, 8.27.

For **37i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-6.9 (m, 5H, Ar), 5.72 (br, 1H, NHCH<sub>3</sub>), 3.59 (q, J = 7.1 Hz, 1H, CHCH<sub>3</sub>), 2.71 (d, J = 4.8 Hz, 3H, NHCH<sub>3</sub>), 1.82 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 1.75 (q, J = 7.5 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 1.01 (d, J = 7.1 Hz, 3H, CHCH<sub>3</sub>), 0.84 (t, J = 7.5 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>); NOESY  $\delta$  3.59  $\leftrightarrow$  1.82 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.4 (CO), 140.4 (Ar), 137.3 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 137.3 (Ar), 135.3 (=*C*(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>3</sub>), 129.3, 128.2 and 126.6 (Ar), 43.0 (CHCH<sub>3</sub>), 29.2 (CH<sub>2</sub>), 26.6 (NHCH<sub>3</sub>), 17.6 (CHCH<sub>3</sub>), 15.5 and 13.3 (CH<sub>3</sub>); GC-MS m/z = 231 (M<sup>+</sup>); calcd for C<sub>25</sub>H<sub>25</sub>NO (M+H)<sup>+</sup>, 232.3460; Found: 232.3487.

For **37j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.0 (m, 5H, Ar), 5.78 (br, 1H, NHCH<sub>3</sub>), 3.66 (q, J = 7.2 Hz, 1H, CHCH<sub>3</sub>), 2.78 (d, J = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1. 86 (s, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.82 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.30 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.11 (m, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 1.06 (d, J = 7.2 Hz, 3H, CHCH<sub>3</sub>), 0.75 (t, J = 7.5 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>); NOESY  $\delta$  3.66  $\leftrightarrow$  1.86 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.4 (CO), 140.5 (C=C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 136.1 (Ar), 135.8 (=C(CH<sub>3</sub>)CH<sub>2</sub>(CH<sub>2</sub>)<sub>2</sub>CH<sub>3</sub>), 129.5, 128.4,

128.0 and 126.6 (Ar), 43.8 (CHCH<sub>3</sub>), 35.8 and 30.7 (CH<sub>2</sub>), 26.6 (NHCH<sub>3</sub>), 22.7 (CH<sub>2</sub>), 18.1 (CHCH<sub>3</sub>), 15.6 and 14.1 (CH<sub>3</sub>); GC-MS m/z = 259; Anal. Calcd for C<sub>17</sub>H<sub>25</sub>NO: C, 78.72; H, 9.71. Found: C, 78.84; H, 9.55.

For **37k**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.2-6.8 (m, 10H, Ar), 5.48 (br, 1H, NHCH<sub>3</sub>), 3.59 (q, *J* = 7.0 Hz, 2H, CHCH<sub>3</sub>), 2.67 (t, J = 7.4 Hz, 2H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 2.64 (d, *J* = 4.8 Hz, 3H, NHCH<sub>3</sub>), 2.33 (m, 1H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 2.16 (m, 1H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 1.89 (s, 3H, CH<sub>3</sub>), 1.01 (d, *J* = 7.0 Hz, 3H, =C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph); NOESY  $\delta$  3.23  $\leftrightarrow$  1.87 ( $\leftrightarrow$  denotes NOE correlation); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.1 (CO), 141.5 and 140.1 (Ar), 136.1 (*C*=C(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 131.0 (=*C*(CH<sub>3</sub>)CH<sub>2</sub>CH<sub>2</sub>Ph), 129.4, 128.6, 128.3, 128.0, 126.7 and 126.1 (Ar), 43.0 (*C*HCH<sub>3</sub>), 37.0 and 34.0 (CH<sub>2</sub>), 26.6 (NHCH<sub>3</sub>), 17.8 (CHCH<sub>3</sub>), 15.3 (CH<sub>3</sub>) ; GC-MS *m*/*z*: 307; Anal. Calcd for C<sub>21</sub>H<sub>25</sub>NO: C, 82.04; H, 8.20. Found: C, 81.65; H, 8.03.

For **371**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 6.61 (br, 1H, N*H*CH<sub>3</sub>), 5.71 (m, 1H, =CH), 5.69 (m, 1H, =CH), 3.14 (m, 1H, CH), 2.97 (dd, *J* = 9.9, 9.8 Hz, 1H, CH), 2.85 (d, *J* = 4.8 Hz, NHC*H*<sub>3</sub>), 2.72 (m, 1H, CH), 2.10 (m, 1H, CH<sub>2</sub>), 2.01 (m, 1H, CH<sub>2</sub>), 1.83 (m, 1H, CH<sub>2</sub>), 1.51 (m, 1H, CH<sub>2</sub>), 0.94 (d, *J* = 6.8 Hz, 3H, CHC*H*<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  177.1 (CO), 140.8 (Ar), 132.5 and 132.2 (CH), 129.6, 127.9 and 126.3 (Ar), 53.6 (CH), 48.8 (CHCH<sub>3</sub>), 45.2 (CH), 32.0 and 28.4 (CH<sub>2</sub>), 26.4 (NHCH<sub>3</sub>), 16.6 (CH*C*H<sub>3</sub>); HRMS (*m*/*z*): calcd for C<sub>16</sub>H<sub>21</sub>NO (M+H)<sup>+</sup>, 244.3570; Found: 244.3534.

For **37m**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.3-7.1 (m, 5H, Ar), 6.16 (br, 1H, N*H*CH<sub>3</sub>), 5.66 (m, 1H, =CH), 5.58 (m, 1H, =CH), 3.13 (m, 1H, CH), 3.05 (dd, *J* = 9.9, 4.8 Hz, 1H, CH), 2.79 (d, *J* = 4.8 Hz, NHC*H*<sub>3</sub>), 2.76 (m, 1H, CH), 2.10 (m, 1H, CH<sub>2</sub>), 2.01 (m, 1H, CH<sub>2</sub>), 1.83 (m, 1H, CH<sub>2</sub>), 1.51 (m, 1H, CH<sub>2</sub>), 0.96 (d, *J* = 7.2 Hz, 3H, CHC*H*<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  176.8 (CO), 141.8 (Ar), 130.1 and 130.0 (CH), 129.1, 128.2 and

126.4 (Ar), 53.5 (CH), 45.7 (*C*HCH<sub>3</sub>), 45.1 (CH), 37.9 and 37.0 (CH<sub>2</sub>), 31.7 (NHCH<sub>3</sub>), 15.0 (CH*C*H<sub>3</sub>); HRMS (*m*/*z*): calcd for C<sub>16</sub>H<sub>21</sub>NO (M+H)<sup>+</sup>, 244.3570; Found: 244.3534.

For **37n**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.30 (dd, *J* = 1.8, 1.1 Hz, 1H, Ar), 6.32 (dd, *J* = 3.2, 1.8 Hz, 1H, Ar), 6.10 (dd, *J* = 3.2, 0.8 Hz, 1H, Ar), 4.10 (m, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.74 (q, J = 7.1 Hz, 1H, CHCH<sub>3</sub>), 1.87 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.80 (s, 3H, =C(CH<sub>3</sub>)<sub>2</sub>), 1.22 (d, *J* = 7.1 Hz, 3H, CHCH<sub>3</sub>), 1.15 (t, *J* = 7.1 Hz, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  174.6 (CO), 152.9 (=*C*(CH<sub>3</sub>)<sub>2</sub>), 140.7 and 135.7 (Ar), 125.2 (*C*C(CH<sub>3</sub>)<sub>2</sub>), 110.3 and 108.5 (Ar), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 41.8 (CHCH<sub>3</sub>), 23.3 and 22.1 (=C(CH<sub>3</sub>)<sub>2</sub>), 15.7 (CHCH<sub>3</sub>), 14.73 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z* = 222 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>18</sub>O<sub>3</sub>: C, 70.24; H, 8.16. Found: C, 70.45; H, 7.97.

For **38e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 10H, Ar), 7.32 (d, *J* = 15.9 Hz, 1H, CH), 6.02 (d, *J* = 15.9 Hz, 1H, CH), 5.97 (br, 1H, NHCH<sub>3</sub>), 3.00 (d, *J* = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.79 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.2 (CO), 140.4 (*C*=C(CH<sub>3</sub>)CO), 138.2 and 137.2 (Ar), 133.7 (CH), 133.3 (C=*C*(CH<sub>3</sub>)CO), 129.5, 128.7 and 128.6 (Ar), 128.3 (CH), 127.9, 127.5 and 126.8 (Ar), 26.6 (NHCH<sub>3</sub>), 18.6 (CH<sub>3</sub>); GC-MS *m*/*z* = 277 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>19</sub>NO: C, 82.28; H, 6.90. Found: C, 81.98; H, 7.04.

For **38f**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 9H, Ar), 7.31 (d, *J* = 15.8 Hz, 1H, CH), 5.96 (d, *J* = 15.9 Hz, 1H, CH), 5.94 (br, 1H, NHCH<sub>3</sub>), 2.99 (d, *J* = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.78 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.9 (CO), 140.2 (*C*=C(CH<sub>3</sub>)CO), 137.9 and 135.6 (Ar), 133.5 (CH), 133.3 (C=*C*(CH<sub>3</sub>)CO), 129.3 (Ar), 128.8 (CH), 128.7, 128.6, 128.5, 127.8 and 127.5 (Ar), 26.6 (NHCH<sub>3</sub>), 18.5 (CH<sub>3</sub>); GC-MS *m*/*z* = 311 (M<sup>+</sup>); Anal. Calcd for C<sub>19</sub>H<sub>18</sub>ClNO: C, 73.19; H, 5.82. Found: C, 73.27; H, 6.00.

For **38g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-6.8(m, 9H, Ar), 7.18 (d, J = 15.9 Hz, 1H, CH), 6.09 (d, J = 15.9 Hz, 1H, CH), 5.96 (br, 1H, NHCH<sub>3</sub>), 3.75 (s, 3H, OCH<sub>3</sub>), 2.98 (d, J = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.77 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.3 (CO), 159.5 (Ar), 140.6 (*C*=C(CH<sub>3</sub>)CO), 138.4 (CH), 133.2, 132.1, 129.5, 128.5 and 128.0 (Ar), 127.4 (CH), 126.3 (C=C(CH<sub>3</sub>)CO), 114.1 (Ar), 55.4 (OCH<sub>3</sub>), 26.5 (NHCH<sub>3</sub>), 18.5 (CH<sub>3</sub>); GC-MS m/z = 307 (M<sup>+</sup>); Anal. Calcd for C<sub>20</sub>H<sub>21</sub>NO<sub>2</sub>: C, 78.15; H, 6.89. Found: C, 77.91; H, 7.02.

For **38h**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.0 (m, 9H, Ar), 7.29 (d, J = 15.9 Hz, 1H, CH), 6.02 (d, J = 15.9 Hz, 1H, CH), 5.92 (br, 1H, 3H, NHCH<sub>3</sub>), 3.00 (d, J = 4.9 Hz, NHCH<sub>3</sub>), 2.30 (s, 3H, PhCH<sub>3</sub>), 1.79 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.3 (CO), 140.6 (*C*=C(CH<sub>3</sub>)CO), 138.3 and 137.9 (Ar), 133.8 (CH), 132.7 (C=*C*(CH<sub>3</sub>)CO), 129.6, 129.5, 128.6 and 127.5 (Ar), 127.4 (CH), 126.7 (Ar), 26.6 (NHCH<sub>3</sub>), 21.4 (PhCH<sub>3</sub>), 18.6 (CH<sub>3</sub>); GC-MS m/z = 291 (M<sup>+</sup>); Anal. Calcd for C<sub>20</sub>H<sub>21</sub>NO: C, 82.44; H, 7.26. Found: C, 82.04; H, 7.29.

For **38i**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.1 (m, 14H, Ar), 7.31 (d, *J* = 15.9 Hz, 1H, CH), 5.98 (d, *J* = 15.9 Hz, 1H, CH), 5.80 (br, 1H, N*H*CH<sub>3</sub>), 2.93 (d, *J* = 4.9 Hz, NHC*H*<sub>3</sub>), 1.73 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.2 (CO), 140.7, 140.6 and 140.5 (Ar), 138.2 (*C*=C(CH<sub>3</sub>)CO), 136.3 (Ar), 133.73 (CH), 129.6, 129.1, 129.0 and 128.6 (Ar), 128.4 (C=*C*(CH<sub>3</sub>)CO), 127.6 (CH), 127.5, 127.3, 127.2 and 127.0 (Ar), 26.7 (NHCH<sub>3</sub>), 18.7 (CH<sub>3</sub>); GC-MS *m*/*z* = 353 (M<sup>+</sup>); Anal. Calcd for C<sub>25</sub>H<sub>23</sub>NO: C, 84.95; H, 6.56. Found: C, 84.59; H, 6.77.

For **38j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.7-7.2 (m, 12H, Ar), 7.39 (d, J = 15.9 Hz, 1H, CH), 6.20 (d, J = 15.9 Hz, 1H, CH), 6.09 (br, 1H, NHCH<sub>3</sub>), 3.01 (d, J = 4.9 Hz, 3H,

NHC*H*<sub>3</sub>), 1.82 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.2 (CO), 140.5 (*C*=C(CH<sub>3</sub>)CO), 138.2, 134.7, 133.8 and 133.6 (Ar), 133.3 (CH), 133.1, 129.5, 128.3, 128.1 and 127.7 (Ar), 127.5 (CH), 127.2 and 126.4 (Ar), 126.1 (C=*C*(CH<sub>3</sub>)CO), 123.5 (Ar), 26.6 (NHCH<sub>3</sub>), 18.7 (CH<sub>3</sub>); GC-MS *m*/*z* = 327 (M<sup>+</sup>); Anal. Calcd for C<sub>23</sub>H<sub>21</sub>NO: C, 84.37; H, 6.46. Found: C, 84.68; H, 6.50.

For **38k**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.6-7.0 (m, 11H, Ar), 7.41 (d, J = 15.9 Hz, 1H, CH), 6.18 (d, J = 15.9 Hz, 1H, CH), 5.92 (br, 1H, NHCH<sub>3</sub>), 3.98 (s, 3H, OCH<sub>3</sub>), 3.05 (d, J = 4.9 Hz, 3H, NHCH<sub>3</sub>), 1.82 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  172.3 (CO), 158.0 (Ar), 140.8 (*C*=C(CH<sub>3</sub>)CO), 138.4 and 134.4 (Ar), 134.1 (CH), 132.7, 132.6, 129.7 and 129.6 (Ar), 129.1 (C=*C*(CH<sub>3</sub>)CO), 128.6 (Ar), 127.7 (CH), 127.5, 127.3 and 127.1 (Ar), 55.5 (OCH<sub>3</sub>), 26.7 (NHCH<sub>3</sub>), 18.7 (CH<sub>3</sub>); GC-MS *m*/*z* = 357 (M<sup>+</sup>); Anal. Calcd for C<sub>24</sub>H<sub>23</sub>NO<sub>2</sub>: C, 80.64; H, 6.49. Found: C, 80.55; H, 6.59.

For **39a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.2 (m, 4H, Ar), 3.72 (s, 3H, OCH<sub>3</sub>), 3.60 (s, 2H, CH<sub>2</sub>), 3.39 (s, 2H, CH<sub>2</sub>), 2.18 (s, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 146.0 (C), 142.3 and 142.2 (Ar), 129.8 (C), 126.4, 124.1, 123.4 and 118.5 (Ar), 52.2 (OCH<sub>3</sub>), 42.9 and 31.5 (CH<sub>2</sub>), 14.3 (CH<sub>3</sub>); GC-MS *m*/*z* = 202 (M<sup>+</sup>); Anal. Calcd for C<sub>13</sub>H<sub>14</sub>O<sub>2</sub>: C, 77.20; H, 6.98. Found: C, 77.61; H, 6.37.

For **39b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.5-7.2 (m, 4H, Ar), 3.73 (s, 3H, OCH<sub>3</sub>), 3.60 (s, 2H, CH<sub>2</sub>), 3.42 (s, 2H, CH<sub>2</sub>), 2.60 (q, *J* = 7.6 Hz, 2H, CH<sub>2</sub>), 1.22 (t, *J* = 7.6 Hz, 3H, CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  171.7 (CO), 148.3 and 146.0 (Ar), 142.3 and 129.0 (C), 126.4, 124.2, 123.5 and 118.7 (Ar), 52.2 (OCH<sub>3</sub>), 40.1, 31.4 and 22.0 (CH<sub>2</sub>), 14.3 (CH<sub>3</sub>); GC-MS *m*/*z* = 216 (M<sup>+</sup>); Anal. Calcd for C<sub>14</sub>H<sub>16</sub>O<sub>2</sub>: C, 77.75; H, 7.46. Found: C, 77.61; H, 7.37. For **39c**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.59 (s, 1H, CH), 6.92 (s, 1H, CH), 4.34 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.91 (d, J = 7.2 Hz, 2H, CH<sub>2</sub>), 2.76 (m, 4H, CH<sub>2</sub>), 2.07 (m, 1H, CH), 1.80 (m, 4H, CH<sub>2</sub>), 1.65 (m, 4H, CH<sub>2</sub>), 1.53 (m, 4H, CH<sub>2</sub>), 1.38 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.4 (CO), 141.3, 140.8 and 138.5 (C), 132.2 and 131.0 (CH), 127. 1 (C), 60.7 (OCH<sub>2</sub>CH<sub>3</sub>), 42.2 (CH<sub>2</sub>), 39.7 (CH), 36.8, 36.3, 33.7, 28.5, 28.4 and 25.0 (CH<sub>2</sub>), 14.5 (OCH<sub>2</sub>CH<sub>3</sub>); HRMS (m/z): calcd for C<sub>20</sub>H<sub>28</sub>O<sub>2</sub> (M)<sup>+</sup>, 286.4146; Found: 286.4167.

For **39d**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.53 (s, 1H, CH), 6.882 (s, 1H, CH), 4.26 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.84 (d, J = 7.2 Hz, 2H, CH<sub>2</sub>), 2.70 (m, 4H, CH<sub>2</sub>), 1.96 (m, 1H, CH), 1.66 (m, 2H, CH<sub>2</sub>), 1.57 (m, 4H, CH<sub>2</sub>), 1.43 (m, 4H, CH<sub>2</sub>), 1.35 (m, 4H, CH<sub>2</sub>), 1.31 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.2 (CO), 141.6, 140.7, 134 (C), 132.2 and 131.4 (CH), 127. 3 (C), 60.7 (OCH<sub>2</sub>CH<sub>3</sub>), 42.1 (CH<sub>2</sub>), 39.8 (CH), 32.7, 29.6, 29.0, 25.0, 23.3 and 23.2 (CH<sub>2</sub>), 14.3 (OCH<sub>2</sub>CH<sub>3</sub>); HRMS (*m*/*z*): calcd for C<sub>20</sub>H<sub>28</sub>O<sub>2</sub> (M)<sup>+</sup>, 300.4415; Found: 300.4457.

For **39e**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.62 (s, 1H, CH), 6.96 (s, 1H, CH), 4.34 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.94 (d, J = 7.1 Hz, 2H, CH<sub>2</sub>), 2.75 (m, 4H, CH<sub>2</sub>), 2.06 (m, 1H, CH), 1.68 (m, 4H, CH<sub>2</sub>), 1.64 (m, 4H, CH<sub>2</sub>), 1.35 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>), 1.19 (m, 4H, CH<sub>2</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.5 (CO), 145.5, 141.9 and 135.6 (C), 132.4 and 131.3 (CH), 127. 7 (C), 60.7 (OCH<sub>2</sub>CH<sub>3</sub>), 42.2 (CH<sub>2</sub>), 39.9 (CH), 34.2, 32.7, 32.4, 32.3, 32.0, 26.1, 25.9 and 25.0 (CH<sub>2</sub>), 14.6 (OCH<sub>2</sub>CH<sub>3</sub>); HRMS (*m*/*z*): calcd for C<sub>21</sub>H<sub>30</sub>O<sub>2</sub> (M)<sup>+</sup>, 314.4684; Found: 314.4723.

For **39f**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.60 (s, 1H, CH), 7.4-7.3 (m, 5H, Ar), 7.08 (s, 1H, CH), 4.09 (q, *J* = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.83 (m, 4H, CH<sub>2</sub>), 1.85 (m, 4H, CH<sub>2</sub>), 1.00 (t, *J* = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.0 (CO), 141.9,

140.9, 140.9 and 136.4 (C), 131.5 (CH), 130.8, 128.6, 128.4 and 128.0 (Ar), 126.9 (CH), 60.8 (OCH<sub>2</sub>CH<sub>3</sub>), 29.5, 29.1, 23.3 and 23.2 (CH<sub>2</sub>), 13.8 (OCH<sub>2</sub>CH<sub>3</sub>); HRMS (*m*/*z*): calcd for C<sub>19</sub>H<sub>20</sub>O<sub>2</sub> (M)<sup>+</sup>, 280.3670; Found: 280.3562.

For **39g**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.50 (s, 1H, CH), 7.3-7.2 (m, 5H, Ar), 7.01 (s, 1H, CH), 3.98 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.77 (m, 4H, CH<sub>2</sub>), 1.76 (m, 2H, CH<sub>2</sub>), 1.58 (m, 4H, CH<sub>2</sub>), 0.88 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.0 (CO), 147.2, 142.6, 141.8 and 140.5 (C), 131.6 (CH), 130.5, 128.6, 128.5 and 128.4 (Ar), 127.0 (CH), 60.8 (OCH<sub>2</sub>CH<sub>3</sub>), 36.7, 36.3, 32.7, 28.3 and 28.2 (CH<sub>2</sub>), 13.8 (OCH<sub>2</sub>CH<sub>3</sub>); HRMS (m/z): calcd for C<sub>20</sub>H<sub>22</sub>O<sub>2</sub> (M)<sup>+</sup>, 294.3939; Found: 294.3948.

For **39h**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.51 (s, 1H, CH), 7.3-7.2 (m, 5H, Ar), 7.01 (s, 1H, CH), 3.98 (q, J = 7.2 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.70 (m, 4H, CH<sub>2</sub>), 1.61 (m, 4H, CH<sub>2</sub>), 1.29 (m, 4H, CH<sub>2</sub>), 0.88 (t, J = 7.2 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.1 (CO), 145.1, 140.5, 139.6 and 138.5 (C), 132.0 (Ar), 131.5 and 130.7 (CH), 127.9, 126.9 and 126.3 (Ar), 60.7 (OCH<sub>2</sub>CH<sub>3</sub>), 32.3, 32.2, 32.1, 31.9, 26.0 and 25.9 (CH<sub>2</sub>), 13.8 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z*: 308; Anal. Calcd for C<sub>21</sub>H<sub>24</sub>O<sub>2</sub>: C, 81.78; H, 7.84. Found: C, 81.78; H, 7.68.

For **60a**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.5-7.3 (m, 4H, Ar), 4.30 (q, J = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 3.77 (s, 2H, CH<sub>2</sub>), 3.05 (t, J = 7.6 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.70 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>) 1.05 (t, J = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>), 0.91 (t, J = 7.5 Hz, 3H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  165.9 (CO), 155.9 ((CH<sub>2</sub>)*C*=C(CO)), 127.5, 126.6, 124.2 and 122.9 (Ar), 121.4 ((CH<sub>2</sub>)C=C(CO)), 60.0 (OCH<sub>2</sub>CH<sub>3</sub>), 39.0, 28.7 and 22.7 (CH<sub>2</sub>), 14.2 (OCH<sub>2</sub>CH<sub>3</sub>); HRMS (m/z): calcd for C<sub>15</sub>H<sub>18</sub>O<sub>2</sub> (M)<sup>+</sup>, 230.3071; Found: 230.3062. For **60b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.4-7.2 (m, 4H, Ar), 4.19 (q, *J* = 7.1 Hz, 2H, OCH<sub>2</sub>CH<sub>3</sub>), 2.83 (dd, *J* = 7.5, 7.3 Hz, 2H, CH<sub>2</sub>), 2.71 (dd, *J* = 7.5, 7.3 Hz, 2H, CH<sub>2</sub>), 2.48 (t, *J* = 7.4 Hz, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 1.56 (m, 2H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>) 1.05 (t, *J* = 7.1 Hz, 3H, OCH<sub>2</sub>CH<sub>3</sub>), 0.89 (t, J = 7.5 Hz, 3H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  169.4 (CO), 145.0 ((CH<sub>2</sub>)C=C(CO)), 138.1, 134.8, 128.4, and 127.8 (Ar), 127.7 ((CH<sub>2</sub>)C=*C*(CO)), 125.0 (Ar), 60.5 (OCH<sub>2</sub>CH<sub>3</sub>), 31.3, 28.5, 25.3 and 23.2 (CH<sub>2</sub>), 14.5 (OCH<sub>2</sub>CH<sub>3</sub>); GC-MS *m*/*z*: 293; Anal. Calcd for C<sub>16</sub>H<sub>20</sub>NO: C, 78.65; H, 8.25. Found: C, 78.41; H, 7.99.

## For Chapter 4

General Procedure of the Catalytic Reaction. In air, the complex 80 (6 mg, 10  $\mu$ mol), an alkane substrate (1.0 mmol) and *t*-BuOOH (70 wt% in H<sub>2</sub>O, 0.43 mL, 3.0 mmol) were dissolved in water (3 mL) in a 25 mL Schlenk tube equipped with a magnetic stirring bar. The reaction mixture was stirred at 20 °C for 2-24 h. The reaction tube was opened to air and the solution was extracted with CH<sub>2</sub>Cl<sub>2</sub> (10 mL). The solution was filtered through a small pad of silica gel. An internal standard (C<sub>6</sub>Me<sub>6</sub>, 20 mg) was added to the solution, and the product yield was determined by GC. The ketone product was readily isolated by a column chromatography on silica gel (hexane/EtOAc).

Synthesis of 80. In a 25 mL Schlenk tube equipped with a Teflon stopcock and a magnetic stirring bar, the complex 82 (100 mg, 0.19 mmol), NaBF<sub>4</sub> (90 mg, 0.95 mmol) and *t*-BuOOH (5.5 M in decane, 0.42 mL, 1.9 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (5 mL). The reaction mixture was stirred for 5 h at room temperature. The solvent was removed under vacuum. The residue was recrystallized in CH<sub>2</sub>Cl<sub>2</sub>/*n*-hexanes to obtain the product

**80** (85 mg, 73% yield). Single crystals of complex **80** suitable for X-ray crystallographic analysis were obtained from CH<sub>2</sub>Cl<sub>2</sub>/*n*-hexanes solution. The Evans NMR method was used to measure the magnetic moment of the complex by following the experimental procedure described in: Girolami, G. S.; Rauchfuss, T. B.; Angelici, R. J. *Synthesis and Technique in Inorganic Chemistry: A Laboratory Manual*, University Science Books: Sausalito, CA, 1999, pp. 125-126.

For **80**: Anal. Calcd for C<sub>20</sub>H<sub>24</sub>BCl<sub>2</sub>F<sub>4</sub>N<sub>4</sub>O<sub>2</sub>Ru: C, 39.30; H, 3.96. Found C, 38.70; H, 3.77.  $\mu_{eff} = 1.55 \text{ B}_{\text{M}}$  at 293 K.

Synthesis of 81. In a glove box, 4,4-dimethyl-2-(2-pyridyl)oxazoline (0.21 g, 1.2 mmol) and  $[Ru(COD)Cl_2]_x$  (0.16 g, 0.5 mmol) were dissolved in ClCH<sub>2</sub>CH<sub>2</sub>Cl (15 mL) in a 25 mL Schlenk tube equipped with a magnetic stirring bar and Teflon stopcock. The reaction tube was brought out of the box, and was stirred in an oil bath at 50 °C for 24 h. After the reaction tube was cooled to room temperature, the volatiles were removed under vacuum, and the residue was recrystallized in CH<sub>2</sub>Cl<sub>2</sub>/*n*-hexanes to obtain a crude product mixture. The mixture was further purified by flash column chromatography (*n*-hexanes/EtOAc = 4:1) to afford analytically pure product **81** (0.15 g, 65% yield). Single crystals of **81** suitable for X-ray crystallographic analysis were obtained from slow evaporation of CH<sub>2</sub>Cl<sub>2</sub> solution.

For **81**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.05 (ddd, J = 5.3, 1.0, 0.7 Hz, py-6-H), 7.86-7.96 (m, 2H, py-3 and 4-H), 7.50 (ddd, J = 12.6, 5.3, 2.0 Hz, py-5-H), 5.03 (t, J = 2.5 Hz, =CH), 4.52 (t, J = 2.6 Hz, =CH), 4.46 (s, 2H, OCH<sub>2</sub>), 2.61-2.78 (m, 4H, CH<sub>2</sub>), 2.03-2.21 (m, 4H, CH<sub>2</sub>), 1.52 (s, 6H, CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  166.6 (N=CO), 150.0,

148.0, 138.2, 128.0 and 126.2 (py), 89.5 and 89.2 (=CH), 82.5 (OCH<sub>2</sub>), 70.3 (*C*CH<sub>3</sub>), 30.5 and 29.0 (CH<sub>2</sub>), 27.5 (*CC*H<sub>3</sub>); Anal. Calcd for C<sub>18</sub>H<sub>24</sub>Cl<sub>2</sub>N<sub>2</sub>ORu: C, 47.37; H, 5.30. Found: C, 47.13; H, 5.22.

Synthesis of 82. In a glove box,  $[Ru(COD)Cl_2]_x (0.20 \text{ g}, 0.44 \text{ mmol})$  and 4,4-dimethyl-2-(2-pyridyl)oxazoline (0.34 g, 1.94 mmol) were dissolved in ClCH<sub>2</sub>CH<sub>2</sub>Cl (15 mL) in a 25 mL Schlenk tube equipped with a Teflon stopcock and a magnetic stirring bar. The reaction mixture was stirred in an oil bath at 100 °C for 24 h. After cooling to room temperature, the solvent was removed under vacuum. The residue was recrystallized in CH<sub>2</sub>Cl<sub>2</sub>/*n*-hexanes to obtain a crude product mixture. The product mixture was further purified by flash chromatography (*n*-hexanes/EtOAc = 4:1) to afford analytically pure product **82** (0.30 g, 65% yield). Alternatively, complex **81** (0.20 g, 0.44 mmol) and pymox-Me<sub>2</sub> (0.34 g, 1.94 mmol) were dissolved in ClCH<sub>2</sub>CH<sub>2</sub>Cl (15 mL) in a 25 mL Schlenk tube. The reaction mixture was removed under vacuum. The residue was purified by flash column chromatography (*n*-hexanes/EtOAc = 4:1) to afford pure product **82** (0.25 g, 55% yield). Single crystals of **82** suitable for X-ray crystallographic analysis were obtained from CH<sub>2</sub>Cl<sub>2</sub>/*n*-hexanes solution.

For **82**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 10.01 and 10.12 (s, py-6-H), 7.83-7.88 and 7.90-7.95 (m, 2H, py-3 and 4-H), 7.53-7.63 and 7.25-7.39 (m, py-5-H), 4.62 and 4.53 (s, OCH<sub>2</sub>), 1.15 and 0.65 (s, CCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 166.8 and 166.5 (N=CO), 156.2, 155.2, 151.6, 151.01, 133.1, 132.9, 132.8, 132.6, 126.1 and 124.9 (py), 83.4 and 82.7 (OCH<sub>2</sub>), 70.6 and 70.3 (CCH<sub>3</sub>), 28.2 and 27.1 (CCH<sub>3</sub>); Anal. Calcd for C<sub>20</sub>H<sub>24</sub>Cl<sub>2</sub>N<sub>4</sub>O<sub>2</sub>Ru: C, 45.81; H, 4.61. Found C, 44.92; H, 4.54.

Catalyst Recycling Experiment. The complex 80 (6 mg, 10  $\mu$ mol) was charged with ethylbenzene (0.12 mL, 1.0 mmol), *t*-BuOOH (70 wt% in H<sub>2</sub>O, 0.43 mL, 3.0 mmol) and H<sub>2</sub>O (2.5 mL) in a thick-walled 25 mL Schlenk tube equipped with a magnetic stirring bar. The reaction mixture was stirred for 16 h at 20 °C. After the reaction was completed, the reaction tube was opened to air and the solution was extracted with CH<sub>2</sub>Cl<sub>2</sub> (10 mL). The extracted solution was filtered through a small pad of silica gel and analyzed by GC. The second and third runs were repeated by using the same aqueous solution. The product yield as determined by GC.

**Hammett Study.** In five separate tubes, an equal amount of the complex **80** (6 mg, 10  $\mu$ mol), *p*-X-C<sub>6</sub>H<sub>4</sub>CH<sub>2</sub>CH<sub>3</sub> (X = OMe, Me, H, F, Cl) (0.12 mL, 1.0 mmol) and *t*-BuOOH (70 wt% in H<sub>2</sub>O, 0.43 mL, 3.0 mmol) were dissolved in H<sub>2</sub>O (1.5 mL) and *n*-hexanes (1.0 mL) in a 25 mL Schlenk tube equipped with a magnetic stirring bar in air. The reaction tubes were stirred at 20 °C. A small portion of the aliquot was drawn periodically from the organic layer, and the conversion was determined by GC. The *k*<sub>obs</sub> was estimated from a first-order plot of  $-ln([ArCH<sub>2</sub>CH<sub>3</sub>]_t/[ArCH<sub>2</sub>CH<sub>3</sub>]_0)$  vs time.

**Isotope Effect Study.** In two separate tubes, complex **80** (6 mg, 10  $\mu$ mol) was charged with ethylbenzene and ethylbenzene- $d_{10}$  (0.12 mL, 1.0 mmol), *t*-BuOOH (70 wt% in H<sub>2</sub>O, 0.43 mL, 3.0 mmol), H<sub>2</sub>O (1.5 mL) and *n*-hexanes (1.0 mL) in a 25 mL Schlenk tube equipped with a magnetic stirring bar in air. The reaction tube was stirred at 20 °C.

A small portion of the aliquot was drawn periodically from the organic layer, and the product conversion was determined by GC. The  $k_{obs}$  was obtained from a first-order plot of  $-ln([ethylbenzene]_t/[ethylbenzene]_0)$  vs time.

**Cyclic Voltammetry of 80.** In a volumetric flask, the sample solution was prepared by dissolving complex **80** (10 mg, 1.6 mM) and an electrolyte (0.25 M of  $Bu_4NPF_6$ ) in 10 mL of CH<sub>2</sub>Cl<sub>2</sub>. Electrochemical measurements were collected at a scan rate of 200 mV/s from a three three-electrode cell composed of a Ag/AgCl electrolyte, a platinum working electrode, and a glassy carbon counter electrode.

Catalytic Oxidation Reaction of Ethylbenzene with TEMPO. In air, complex 80 (6 mg, 10  $\mu$ mol) was charged with ethylbenzene (0.12 mL, 1.0 mmol), *t*-BuOOH (70 wt% in H<sub>2</sub>O, 0.43 mL, 3.0 mmol), TEMPO (16 mg, 0.1 mmol), H<sub>2</sub>O (1.5 mL) and *n*-hexanes (1.0 mL) in a thick-walled 25 mL Schlenk tube equipped with a magnetic stirring bar. The reaction mixture was stirred for 24 h at 20 °C. After the reaction was completed, the reaction tube was opened to air. The solution was extracted with CH<sub>2</sub>Cl<sub>2</sub> (10 mL) and organic solution was filtered through a small pad of silica gel. The product yield as determined by GC was 34% (without TEMPO, 40% conversion). It should be noted that *n*-hexanes was added to dissolve TEMPO, and under these biphasic conditions, the reaction rate was considerably lower than in pure water.

Table 19. Crystal Data and Structure Refinement for 30a.

Empirical formula	$C_{38}H_{55}BCl_2F_4NO_2PRu$	
Formula weight	847.58	
Temperature	100(2) K	
Wavelength	0.7107 Å	
Crystal system	Triclinic	
Space group	P-1	
Unit cell dimensions	$a = 10.23664(13) \text{ Å}  \alpha = 100.1596(12)^{\circ}$	
	b = 12.0713(2) Å $\beta$ = 102.3207(11)°	
	$c = 16.7295(2) \text{ Å} \qquad \gamma = 103.8371(12)^{\circ}$	
Volume	1903.84(5) Å <sup>3</sup>	
Z	2	
Density (calculated)	1.479 Mg/m <sup>3</sup>	
Absorption coefficient	$0.649 \text{ mm}^{-1}$	
F(000)	880	
Crystal size	0.4281 x 0.2686 x 0.1765 mm <sup>3</sup>	
Theta range for data collection	3.38 to 37.83°	
Index ranges	$-17 \le h \le 17, -20 \le k \le 20, -28 \le l \le 28$	
Reflections collected	95037	
Independent reflections	19744 [R(int) = 0.0279]	
Absorption correction	Numerical	
Max. and min. transmission	0.915 and 0.811	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	19744 / 0 / 487	
Goodness-of-fit on F <sup>2</sup>	1.004	
Final R indices [I>2sigma(I)]	$R_1 = 0.0214, \ wR_2 = 0.0609$	
R indices (all data)	$R_1 = 0.0270, wR_2 = 0.0625$	
Largest diff. peak and hole	0.692 and -0.496 e.Å <sup>-3</sup>	

Table 20. Crystal Data and Structure Refinement for 56.

Empirical formula	$C_{40}H_{59}BCl_2F_4NO_3PRu$	
Formula weight	891.63	
Temperature	100(2) K	
Wavelength	0.7107 Å	
Crystal system	Monoclinic	
Space group	C 2	
Unit cell dimensions	a = 25.7127(4) Å	$\alpha = 90.00^{\circ}$
	b = 10.6683(2) Å	$\beta = 94.9531(14)^{\circ}$
	c = 15.1324(2) Å	$\gamma = 90.00^{\circ}$
Volume	4135.48 Å <sup>3</sup>	
Z	4	
Density (calculated)	$1.432 \text{ Mg/m}^3$	
Absorption coefficient	$0.603 \text{ mm}^{-1}$	
F(000)	1856	
Crystal size	0.3531 x 0.2366 x 0.2	$2077 \text{ mm}^3$
Theta range for data collection	3.35 to 32.83°	
Index ranges	-38≤ <i>h</i> ≤38, -160≤ <i>k</i> ≤15, -22≤ <i>l</i> ≤23	
Reflections collected	34927	
Independent reflections	13807 [R(int) = 0.0323]	
Absorption correction	Numerical	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	13807 / 145 / 406	
Goodness-of-fit on F <sup>2</sup>	1.075	
Final R indices [I>2sigma(I)]	$R_1 = 0.0474, wR_2 = 0$	.1135
R indices (all data)	$R_1 = 0.0598, wR_2 = 0.1197$	
Largest diff. peak and hole	0.955 and -0.994 e.Å	3

Empirical formula	$C_{21}H_{26}BCl_4F_4N_4O_2Ru$	
Formula weight	696.14	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P 21/c	
Unit cell dimensions	a = 8.35260(10) Å	$\alpha = 90^{\circ}$
	b = 28.1943(4) Å	$\beta = 108.2210(10)^{\circ}$
	c = 12.6986(2) Å	$\gamma=90^\circ$
Volume	2840.52(7) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.628 Mg/m <sup>3</sup>	
Absorption coefficient	8.407 mm <sup>-1</sup>	
F(000)	1396	
Crystal size	0.55 x 0.41 x 0.05 mm <sup>3</sup>	
$\theta$ range for data collection	3.99 to 67.75°	
Index ranges	$-9 \le h \le 9,  0 \le k \le 33,  0 \le l \le 15$	
Reflections collected	23226	
Independent reflections	5024 [R(int) = 0.0211]	
Completeness to $\theta = 67.75^{\circ}$	97.6 %	
Absorption correction	Numerical	
Max. and min. transmission	0.6786 and 0.0905	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	5024 / 9 / 370	
Goodness-of-fit on F <sup>2</sup>	0.969	
Final R indices [I>2 $\sigma$ (I)]	$R_1 = 0.0478, wR_2 = 0.1237$	
R indices (all data)	$R_1 = 0.0489, wR_2 = 0.1245$	
Largest diff. peak and hole	1.507 and -1.065 e.Å <sup>-3</sup>	

Table 21. Crystal Data and Structure Refinement for 80.

Empirical formula	$C_{18.5}H_{25}C_{13}N_2ORu$	
Formula weight	498.83	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P21/c	
Unit cell dimensions	a = 16.4410(4) Å	$\alpha = 90^{\circ}$
	b = 9.8816(3) Å	$\beta = 110.3760(10)^{\circ}$
	c = 12.8838(3) Å	$\gamma=90^\circ$
Volume	1962.17(9) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.689 Mg/m <sup>3</sup>	
Absorption coefficient	10.305 mm <sup>-1</sup>	
F(000)	1012	
Crystal size	0.50 x 0.44 x 0.32 mm <sup>3</sup>	
$\theta$ range for data collection	5.32 to 67.53°	
Index ranges	$-19 \le h \le 17, 0 \le k \le 11, 0 \le l \le 15$	
Reflections collected	16178	
Independent reflections	3383 [R(int) = 0.0305]	
Completeness to $\theta = 67.53^{\circ}$	95.6 %	
Absorption correction	Numerical	
Max. and min. transmission	0.1371 and 0.0791	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	3383 / 0 / 332	
Goodness-of-fit on F <sup>2</sup>	1.115	
Final R indices [I>2 $\sigma$ (I)]	$R_1 = 0.0211$ , $wR_2 = 0.0516$	
R indices (all data)	$R_1 = 0.0212, wR_2 = 0.0517$	
Extinction coefficient	0.00074(5)	
Largest diff. peak and hole	0.639 and -0.452 e.Å <sup>-3</sup>	

Table 22. Crystal Data and Structure Refinement for 81.

Empirical formula	$C_{20}H_{24}C_{12}N_4O_{2.14}Ru$	
Formula weight	526.68	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P 21/c	
Unit cell dimensions	a = 10.72140(10) Å	$\alpha = 90^{\circ}$
	b = 14.1607(2) Å	$\beta = 97.3240(10)^{\circ}$
	c = 14.6702(2) Å	$\gamma=90^\circ$
Volume	2209.09(5) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.584 Mg/m <sup>3</sup>	
Absorption coefficient	8.173 mm <sup>-1</sup>	
F(000)	1069	
Crystal size	0.29 x 0.15 x 0.10 mm <sup>3</sup>	
$\theta$ range for data collection	4.16 to 68.00°	
Index ranges	$-12 \le h \le 12, 0 \le k \le 16, 0 \le l \le 17$	
Reflections collected	18294	
Independent reflections	3921 [R(int) = 0.0163]	
Completeness to $\theta = 68.00^{\circ}$	97.6 %	
Absorption correction	Numerical	
Max. and min. transmission	0.4954 and 0.2003	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	3921 / 0 / 272	
Goodness-of-fit on F <sup>2</sup>	0.981	
Final R indices [I>2 $\sigma$ (I)]	$R_1 = 0.0194, wR_2 = 0.0513$	
R indices (all data)	$R_1 = 0.0197, wR_2 = 0.0515$	
Extinction coefficient	0.00020(3)	
Largest diff. peak and hole	0.405 and -0.295 e.Å <sup>-3</sup>	

*Table 23*. Crystal Data and Structure Refinement for 82.

## References

Recent reviews: (a) Ritleng, V.; Sirlin, C.; Pfeffer, M. *Chem, Rev.* 2002, *102*, 1731. (b)
 Kakiuchi, F.; Murai, S. In *Activation of Unreactive Bonds and Organic Synthesis*; Murai,
 S. Ed.; Springer: New York, 1999. (c) Chen, X.; Engle, K. M.; Wang, D.-H.; Yu, J.-Q.
 *Angew. Chem., Int. Ed.* 2009, *48*, 5094–5115. (d) Colby, D. A.; Bergman, R. G.; Ellman,
 J. A. *Chem. Rev.* 2010, *110*, 624–655. (e) Lyons, T. W.; Sanford, M. S. *Chem. Rev.* 2010, *110*, 1147–1169.

(2) (a) Heck, R. F. In *Comprehensive Organic Synthesis*; Trost, B. M., Fleming, I., Eds.;
Pergamon Press: Oxford, 1991; Vol. 4. (b) Beletskaya, I. P.; Cheprakov, A. V. *Chem. Rev.* **2000**, *100*, 3009–3066.

(3) Murai, S.; Kakiuchi, F.; Sekine, S.; Tanaka, Y.; Kamatani, A.; Sonoda, M.; Chatani, N. *Nature* 1993, *366*, 529.

(4) (a) Murai, S.; Kakiuchi, F.; Sekine, S.; Tanaka, Y.; Kamatani.A.; Sonoda, M.; Chatani, N. *Pure Appl. Chem.* **1994**, *66*, 1527. (b) Trost, B. M.; Imi, K.; Davies, I. W. *J. Am. Chem. Soc.* **1995**, *117*, 5371. (c) Kakiuchi, F.; Sekine, S.; Tanaka, Y.; Kamatani, A.; Sonda, M.; Chatani, N.; Murai, S. *Bull. Chem. Soc. Jpn.* **1995**, *68*, 62. (d) Sonda, M.; Kakiuchi, F.; Chatani, N.; Murai, S. *J. Orgnomet. Chem.* **1995**, *504*,151. (e) Kakiuchi, F.; Tanaka, Y.; Sato, T.; Chatani, N.; Murai, S. *Chem. Lett.* **1995**, 679. (f) Sonda, M.; Kakiuchi, F.; Kamatani, A.; Chatani, N.; Murai, S. *Chem. Lett.* **1996**, 109. (g) Kakiuchi, F.; Yamauchi, M.; Chatani, N.; Murai, S. *Chem. Lett.* **1996**, 111

(5) (a) Guo, H.; Huang, D.; Weber, W. P. Poym. Prep. 1999, 40, 70. (b) Londergan, T.

M.; Teng, C. J.; Weber, W. P. Polym. Prep. 1999, 40, 154. (c) Gupta, S.; Londergan, T.

- M.; Paulasaari, J. K.; Sargent, J. R.; Weber, W. P. Polym. Mater. Sci. Eng. 1999, 80, 445.
- (6) (a) Harris, P. W. R.; Woodgate, P. D. J. Organomet. Chem. 1996, 506, 339. (b) Harris,
- P. W. R.; Woodgate, P. D. J. Organomet. Chem. 1997, 530, 211. (c) Harris, P. W. R.;

Richard, C. E. F.; Woodgate, P. D. J. Organomet. Chem. 1999, 589, 168.

(7) (a) Lim, Y.-G.; Kim. Y. H.; Kang, J.-B. J. Chem. Soc., Chem. Commun. 1994, 2267.
(b) Lim, Y.-G.; Kang, J.-B.; Kim, Y. H. J. Chem. Soc., Perkin Trans. 1 1996, 2201. (c) Lim, Y.-G.; Kang, J.-B.; Kim, Y. H. Chem. Commun. 1996, 585. (d) Lim, Y.-G.; Kang, J.-B.; Kim, Y. H. J. Chem. Soc., Perkin Trans. 1 1998, 699. (e) Lim, Y.-G.; Kang, J.-B.; Koo, B. T. Tetrahedron Lett. 1999, 40, 7691.

(8) Lenges, C. P.; Brookhart, M. J. Am. Chem. Soc. 1999, 121, 6616.

(9) Guari, Y.; Sabo-Etienne, S.; Chaudret, B. J. Am. Chem. Soc. 1998, 120, 4228.

(10) (a) Busch, S.; Leitner, W. Chem. Commun. 1999, 2305. (b) Busch, S.; Leitner, W. Adv. Synth. Catal. 2001, 343, 192.

(11) (a) Halbritter, G.; Knoch, F.; Wolski, A.; Kisch, H. Angew. Chem., Int. Ed. Engl.
1994, 33, 1603. (b) Halbritter, G.; Konch, F.; Kisch, H. J. Organomet. Chem. 1995, 492,
87. (c) Aulwurm, U. R.; Melchinger, J. U.; Kisch, H. Oranometallics 1995, 14, 3385. (d)
Durr, U.; Henemann, F. W.; Kisch, H. J. Organomet. Chem. 1997, 541, 307.

(12) Jun, C.-H.; Hong, J.-B.; Kim, Y.-H.; Chung, K.-W. Angew. Chem., Int. Ed. 2000, 39, 3440.

- (13) Tan, K. L.; Bergman, R. G.; Ellman, J. A. J. Am. Chem. Soc. 2001, 123, 2685.
- (14) Chen, H.; Schlecht, S.; Semple, T. C.; Hartwig, J. F. Science 2000, 287, 1995.
- (15) (a) Wasa, M.; Engle, K. M.; Yu, J.-Q. J. Am. Chem. Soc. 2010, 132, 3680. (b) Yoo,

E. J.; Wasa, M.; Yu, J.-Q. J. Am. Chem. Soc. 2010, 132, 17378.

(16) (a) Tan, K. L.; Bergman, R. G.; Ellman, J. A. J. Am. Chem. Soc. 2001, 123, 2685. (b)

Thalji, R. K.; Ahrendt, K. A.; Bergman, R. G.; Ellman, J. A. J. Am. Chem. Soc. 2002. 124, 13964.

(17) Jones, W. D.; Feher, F. J. Organometallics 1983, 2, 562.

(18) Liu, F.; Pak, E. B.; Singh, B.; Jensen, C. M.; Goldman, A. S. J. Am. Chem. Soc.1999, 121. 4086.

- (19) Renkema, K. B.; Kissin, Y. V.; Goldman, A. S. J. Am. Chem. Soc. 2003, 126. 7770.
- (20) Yi, C. S.; Lee, D. W. Organometallics 2009, 28, 4266.

- (21) Li, Z.; Li, C. J. Am. Chem. Soc. 2002, 126, 11810.
- (22) Davies, H.; Hansen, T.; Churchill, M. R. J. Am. Chem. Soc. 2000, 122, 3063.
- (23) Oi, S.; Ogino, Y.; Fukita, S.; Inoue, Y. Org. Lett. 2002, 4, 1783.
- (24) Martinez, R.; Simon, M.-O.; Chevalier, R.; Pautigny, C.; Genet, J.-P.; Darses, S. J. Am. Chem. Soc. 2009, 131, 7887.
- (25) Patureau, F. W.; Glorius, F. J. Am. Chem. Soc. 2010, 132, 9982.
- (26) Patureau, F. W.; Besset, T.; Glorius, F. Angew. Chem. Int. Ed. 2011, 50, 1064.
- (27) Wang, X.; Lu, Y.; Dai, H.-X.; Yu, J.-Q. J. Am. Chem. Soc. 2010, 132, 12203.
- (28) Kalyani, D.; Sanford, M. S. Org. Lett. 2005, 7, 4149.
- (29) Jia, C.; Piao, D.; Oyamada, J.; Lu, W.; Kitamura, T.; Fujiwara, Y. *Science* **2000**, 287, 1992.
- (30) Weissman, H.; Song, X.; Milstein, D. J. Am. Chem. Soc. 2001, 123, 337.
- (31) Cho, J.-Y.; Tse, M. K.; Holmes, D.; Malecazka Jr., R. E.; Smith, M. R. Science **2002**, 295, 305.
- (32) Tsukada, N.; Hartwig, J. F. J. Am. Chem. Soc. 2005, 127, 5022.
- (33) Jun, C.-H.; Moon, C. W.; Kim, Y.-M.; Lee, H.; Lee, J. H. *Tetrahedron Lett.* **2002**, *43*, 5604.
- (34) Colby, D. A.; Bergman, R. G.; Ellman, J. A. J. Am. Chem. Soc. 2006. 128, 5604.
- (35) DeBoef, B.; Pastine, S. J.; Sames, D. J. Am. Chem. Soc. 2004, 126, 6556.
- (36) Pastine, S. J.; Gribkov, D. V.; Sames, D. J. Am. Chem. Soc. 2006, 128, 14220.
- (37) Chatani, N.; Asaumi, T.; Yorimitsu, S.; Ikeda, T.; Kakiuchi, F.; Murai, S. J. Am. Chem. Soc. 2001, 123, 10935.
- (38) Yi, C. S.; Yun, S. Y. Organometallics 2004, 23, 5392.
- (39) Liu, C.; Han, X.; Wang, X.; Widenhoefer, R. A. J. Am. Chem. Soc. 2004, 126, 3700.
- (40) Liu, C.; Widenhoefer, R. A. J. Am. Chem. Soc. 2004, 126, 10250.
- (41) Wang, X.; Lane, B. S.; Sames, D. J. J. Am. Chem. Soc. 2005, 127, 4996.
- (42) Li, Z.; Li, C.-J. J. Am. Chem. Soc. 2005, 127, 6968.
(43) Moore, E. J.; Pretzer, W. R.; O'Connell, T. J.; Harris, J.; LaBounty, L.; Chou, L.; Grimmer, S. S. J. Am. Chem. Soc. **1992**, *114*, 5888.

(44) Murakami, M.; Hori, S. J. Am. Chem. Soc. 2003, 125, 4720.

- (45) Matsuura, Y.; Tamura, M.; Kochi, T.; Sato, M.; Chatani, N.; Kakiuchi, F. J. Am. Chem. Soc. 2007, 129, 9858.
- (46) Cho, S. H.; Hwang, S. J.; Chang, S. J. Am. Chem. Soc. 2008, 130, 9254.
- (47) Gao, K.; Yoshikai, N. J. Am. Chem. Soc. 2011, 133, 400.
- (48) Tsujita, H.; Ura, Y.; Matsuki, S.; Wada K.; Mitsudo, T.; Kondo, T. Angew. Chem. Int. Ed. 2007, 46, 5160.
- (49) Inoue, S.; Shiota, H.; Fukumoto, Y.; Chatani, N. J. Am. Chem. Soc. 2009, 131, 6898.
- (50) Tsuchikama, K.; Kasagawa, M.; Endo, K.; Shibata, T. Org. Lett. 2009, 11, 1821.
- (51) Shibata, Y.; Otake, Y.; Hirano, M. Tanaka, K. Org. Lett. 2009, 11, 689.
- (52) Tsai, A. S.; Brasse, M.; Bergman, R. G.; Ellman, J. A. Org. Lett. 2011, 13, 540.
- (53) Wang, F.; Song, G.; Li, X. Org. Lett. 2010, 12, 5430.
- (54) Rakshit, S.; Patureau, F. W.; Glorius, F. J. Am. Chem. Soc. 2010, 132, 9585.
- (55) Jia, C.; Kitamura, T.; Fujiwara, Y. Acc. Chem. Res. 2001, 34, 633
- (56) (a) Dick, A. R.; Hull, K. L.; Sanford, M. S. J. Am. Chem. Soc. 2004, 126, 2300. (b)
- Wan, X.; Ma, Z.; Li, B.; Zhang, K.; Cao, S.; Zhang, S.; Shi, Z. J. Am. Chem. Soc. 2006,
- 128, 7416. (c) Thu, H.-Y.; Yu, W.-Y.; Che, C.-M. J. Am. Chem. Soc. 2006, 128, 9048.
- (57) (a) Stuart, D. R.; Fagnou, K. Science 2007, 316, 1172. (b) Stuart, D. R.; Villemure,
  E.; Fagnou, K. J. Am. Chem. Soc. 2007, 129, 12072. (c) Campeau, L.-C.; Schipper, D. J.;
- Fagnou, K. J. Am. Chem. Soc. 2008, 130, 3266.
- (58)(a) Boele, D. K. M.; van Strijdonck, G. P. F.; de Vries, A. H. M.; Kamer, P. C. J.; de Vries, J. G.; van Leeuwen, P. W. N. M. J. Am. Chem. Soc. 2002, 124, 1586. (b) Beck, E. M.; Grimster, N. P.; Hatley, R.; Gaunt, M. J. J. Am. Chem. Soc. 2006, 128, 2528. (c) Matsuura, Y.; Tamura, M.; Kochi, T.; Sato, M.; Chatani, N.; Kakiuchi, F. Am. Chem. Soc. 2007, 129, 9858. (d) Cai, G.; Fu, Y.; Wan, X.; Shi, Z. Am. Chem. Soc. 2007, 129,

7666. (e) Cho, S. H.; Hwang, S. J.; Chang, S. J. Am. Chem. Soc. 2008, 130, 9254. (f)
Tsuchikama, K.; Kasagawa, M.; Endo, K.; Shibata, T. Org. Lett. 2009, 11, 1821. (g) Wu,
J.; Cui, X.; Chen, L.; Jiang, G.; Wu, Y. J. Am. Chem. Soc. 2009, 131, 13888. (h)
Patureau, F. W.; Glorius, F. J. Am. Chem. Soc. 2010, 132, 9982.

- (59) Wang, D.-H.; Engle, K. M.; Shi, B.-F.; Yu, J.-Q. Science 2010, 327, 315.
- (60) (a) Yi, C. S.; Lee, D. W. Organometallics 2009, 28, 4266. (b) Yi, C. S.; Lee, D. W. Organometallics 2010, 29, 1883. (c) Yi, C. S.; Zeczycki, T. N.; Guzei, I. A. Organometallics 2006, 25, 1047.
- (61) (a) Yi, C. S.; Lee, D. W.; Chen, Y. Organometallics 1999, 18, 2043. (b) Yi, C. S.;
- He, Z.; Lee, D. W.; Rheingold, A. L.; Lam, K.-C. *Organometallics* **2000**, *19*, 2036. (c) Yi, C. S.; He, Z.; Lee, D. W. *Organometallics* **2001**, *20*, 802.
- (62) (a) Singleton, D. A.; Thomas, A. A. J. Am. Chem. Soc. 1995, 117, 9357. (b) Frantz,
  D. E.; Singleton, D. A.; Snyder, J. P. J. Am. Chem. Soc. 1997, 119, 3383.
- (63) Melander, L.; Saunders, W. H., Jr. *Reaction Rates of Isotopic Molecules*; Wiley: New York, 1980, pp 95-102.
- (64) (a) Jazzar, R. F. R.; Mahon, M. F.; Whittlesey, M. K. Organometallics 2001, 20, 3745. (b) Drouin, S. D.; Amoroso, D.; Yap, G. P. A.; Fogg, D. E. Organometallics 2002, 21, 1042.
- (65) (a) Ueno, S.; Mizushima, E.; Chatani, N.; Kakiuchi, F. J. Am. Chem. Soc. 2006, 128,
- 16516. (b) Giri, R.; Lam, J. K.; Yu, J.-Q. J. Am. Chem. Soc. 2010, 132, 686.
- (66) (a) Lu, P.; Paulasaari, J.; Jin, K.; Bau, R.; Weber, W. P. Organometallics 1998, 17,
- 584. (b) Lenges, C. P.; Brookhart, M. J. Am. Chem. Soc. 1999, 121, 6616.
- (67) (a) Stoutland, P. O.; Bergman, R. G. J. Am. Chem. Soc. 1985, 107, 4581. (b)
- Renkema, K. B.; Kissin, Y. V.; Goldman, A. S. J. Am. Chem. Soc. 2003, 125, 7770.
- (68) Recent review: (a) Flynn, A. B.; Ogilvie, W. W. Chem. Rev. 2007, 107, 4698. (b)
- Arai, T.; Suemitsu, Y. Pure Appl. Chem. 2010, 82, 1458.

(69) (a) Negishi, E.; Huang, Z.; Wang, G.; Mohan, S.; Wang, C.; Hattori, H. Acc. Chem. *Res.* 2008, 41, 1474. (b) Itami, K.; Kamei, T.; Yoshida, J. J. Am. Chem. Soc. 2003, 125, 14670.

(70) (a) Hojo, D.; Noguchi, K.; Tanaka, K. Angew. Chem., Int. Ed. 2009, 48, 8129. (b) Ni,
Y.; Kassab, R. M.; Chevliakov, M. V.; Montgomery, J. Am. Chem. Soc. 2009, 131, 17714.
(71) (a) Stüdemann, T.; Knochel, P. Angew. Chem. 1997, 109, 132. (b) Itami, K.; Kamei,
T.; Yoshida, J.-I. J. Am. Chem. Soc. 2003, 125, 14670. (c) Lemay, A. B.; Vulic, K. S.;

Ogilvie, W. W. J. Org. Chem. 2006, 71, 3615.

(72) (a) Zhou, C.; Larock, R. C. J. Org. Chem. 2005, 70, 3765. (b) Zhou, C.; Larock, R.
C. J. Org. Chem. 2006, 71, 3184. (c) McKinley, N. F.; O'Shea, D. F. J. Org. Chem. 2006, 71, 9552.

(73) (a) Berlin, J. M.; Campbell, K.; Ritter, T.; Funk, T. W.; Chlenov, A.; Grubbs, R. H. *Org. Lett.* 2007, *9*, 1339. (b) Stenne, B.; Timperio, J.; Savoie, J.; Dudding T.; Collins, S.
K. *Org. Lett.* 2010, *12*, 2032. (c) Gansäuer, A.; Pierobon, M.; Bluhm, H. *Angew. Chem. Int. Ed.* 2002, *114*, 3341.

(74) (a) Berlin, J. M.; Campbell, K.; Ritter, T.; Funk, T. W.; Chlenov, A.; Grubbs, R. H. Org, Lett. 2007, 9, 1339. (b) Vorfalt, T.; Leuthäuβer, S.; Plenio, H. Angew. Chem. Int. Ed. 2009, 48, 5191. (c) Stenne, B.; Timperio, J.; Savoie, J.; Dudding, T.; Collins, S. K. Org. Lett. 2010, 12, 2032. (d) Donohoe, T. J.; Race, N. J.; Bower, J. F.; Callens, C. K. Org. Lett. 2010, 12, 4094.

(75) (a) Ho, C.-Y.; Ohmiya, H.; Jamison, T. F. *Angew. Chem. Int. Ed.* 2008, 47, 1893. (b)
Matsubara, R.; Jamison, T. F. *J. Am. Chem. Soc.* 2010, *132*, 6880. (c) Ogoshi, S.; Hada,

T.; Ohashi, M. J. Am. Chem. Soc. 2009, 131, 10350. (d) Li, W.; Chen, N.; Montgomery, J. Angew. Chem. Int. Ed. 2010, 49, 8712.

(76) Recent review: (a) Derien, S.; Monnier, F.; Dixneuf. P. H. In *Ruthenium Catalysts and Fine Chemistry*; Bruneau, C.; Dixneuf, P. H. Ed.; Springer: Berlin, 2004. (b) Yamamoto, Y.; Itoh, K. In *Ruthenium in Organic Sythesis*; Murahashi, S.-I. Ed.; Wiley-VCH: Weinheim, 2004. (c) Kakiuchi, F.; Murai, S. In *Activation of Unreactive Bonds and Organic Sythesis*; Murai, S. Ed.; Springer: New York, 1999. (d) Colby, D. A.; Bergman, R. G.; Ellman, J. A. *Chem. Rev.* 2010, *110*, 624-655. (e) Beccalli, E. M.; Broggini, G.; Martinelli, M.; Sottocornola, S. *Chem. Rev.* 2007, *107*, 5318-5365. (f) Ritleng, V.; Sirlin, C.; Pfeffer, M. *Chem. Rev.* 2002, *102*, 1731-1770. (g) Trost, B. M.; Toste, D.; Pinkerton, A. B. *Chem. Rev.* 2001, *101*, 2067-2098. (h) Jang, H.-Y.; Krische, M. J. Acc. Chem. Res. 2004, *37*, 653-601.

(77) (a) Yi, C. S.; Lee, D. W. Organometallics 2009, 28, 4266-4268. (b) Yi, C. S.; Lee, D.
W. Organometallics 2010, 29, 1883-1885. (c) Kwon, K.-H.; Lee, D. W.; Yi, C. S.
Organometallics 2010, 29, 5748-5750. (d) Kwon, K.-H.; Lee, D. W.; Yi, C. S. Angew.
Chem. Int. Ed. 2011, 50, 1692-1695.

(78) (a) Sheldon, R. A.; Kochi, J. K. *Metal-Catalyzed Oxidations of Organic Compounds*;
Academic Press: New York, 1981; (b) Shilov, A. E. *Activation of Saturated Hydrocarbons by Transition Metal Complexes*; Reidel: Dordrecht, The Netherlands, 1984.
(c) Hill, C. L.; *Activation and Functionaliztion of Alkanes*; Wily: New York, 1989. (d) Shilov, A. F.; Shulpin, G. A. *Chem. Rev.* **1997**, *97*, 2879.

- (79) (a) Hill, C. L. Activation and Functionalization of Alkanes; Wiley: New York, 1989.
  (b) Haines, A. H. Methods for the Oxidation of Organic Compounds, Alkanes, Alkenes Alkynes and Arenes; Academic Press: London, 1985, Chapter 2.
- (80) Murahashi, S. I.; Komiya, N.; Oda, Y.; Kuwabara, T.; Natoa, T. J. Org. Chem. 2000, 65, 9186.
- (81) Blake, R. C., II; Coon, M. J. J. Biol. Chem. 1981, 256, 12127.
- (82) Khanna, R. K.; Pauling, T. M.; Vajpayee, D. Tetrahedron Lett. 1991, 32, 3759.
- (83) (a) Sakurai, H.; Hosomi, A. J. Am. Chem. Soc. 1967, 89, 458. (b) Walling, C.;
- McGuinness, J. A. J. Am. Chem. Soc. 1969, 91, 2053.
- (84) Russell, G. A. *Free Radicals*; Kochi, J. K., Ed.; Wiley: New York, 1973; Vol. 1, pp 275-331.
- (85) Hjelmeland, L. M.; Aronow, L.; Trudell, J. R. Biochem. Biophys. Res. Commun.1977, 76, 541.
- (86) Kim, J.; Harrison, R. G.; Kim, C.; Que, L., Jr. J. Am. Chem. Soc. 1996, 118, 4373.
  (87) Che, C. M.; Cheng, K. W.; Chan, M. C. W.; Lau, T. C.; Mak, C. K. J. Org. Chem.
  2000, 65, 7996.
- (88) Goldstein, A. S.; Beer, R. H.; Drago, R. S. J. Am. Chem. Soc. 1994, 116, 2424.
- (89) (a) Doyle, M. P.; Catino, A. J.; Forslund, R. J. J. Am. Chem. Soc. 2004, 126, 13622.
- (90) (a) Bartlett, P. D.; Gunther, P. J. Am. Chem. Soc. 1966, 88, 3288. (b) Bartlett, P. D.;
  Guaraldi, G. J. Am. Chem. Soc. 1967, 89, 4799.
- (91) (a) Kochi, J. K. *Tetrahedron* 1962, *18*, 483. (b) Kochi, J. K.; Mains, H. E. *J. Org. Chem.* 1965, *30*, 1862. (c) Minisci, F. *Acc. Chem. Res.* 1975, *8*, 165. (d) Araneo, S.;
  Fontana, F.; Minisci, F.; Recupero, F.; Serri, A. *Chem. Commun.* 1995, 1399. (e) Minisci,

F.; Fontana, F.; Araneo, S.; Recupero, F.; Banfi, S.; Quici, S. J. Am. Chem. Soc. 1995, 117, 226.

(92) White, M, C.; Chen, M. C. Science. 2007, 318, 783.

(93) Crabtree, R. H. *The Oragnometallic Chemistry of Transition Metals*; 3<sup>nd</sup> Ed,; Wiley International, New York, 2005.

(94) Cornil, B.l; Herrmannn, W. A. *Catalysis from A to Z*; Schlogl, R., Ed,: John Wiley & Sons, 2008.

(95) Li, C. J. Acc. Chem. Res., 2002, 35, 533.

(96) Herrerias, C. I.; Yao, X.; Li, Z.; Li, C. J. Chem. Rev. 2007, 107, 2546.

(97) (a) Lersch, M.; Tilset, M. Chem. Rev. 2005, 105, 2471. (b) Activation of Small

Molecules; William B. Tolman., Eds.; Wiley-VCH: Weinheim, 2006. (c)Periana, R. A.;

Taube, D. J.; Gamble, S.; Taube, H.; Satoh, T.; Fujii, H. Science 1998, 280, 560.

(98) Le Bras, J.; Muzart, J. Tetrahedron Lett. 2002, 43, 431.

(99) Sun, W.; Wang, H.; Xia, C.; Li, j.; Zhao, P. Angew. Chem. Int. Ed. 2003, 42, 1042.

(100) Wei, C.; Li, C.-J. J. Am. Chem. Soc. 2002, 124, 5638.

(101) Hirai, Y.; Kojima, T.; Mizutani, Y.; Shiota, Y.; Yoshizawa, K.; Fukuzumi, S. Angew. Chem. Int. Ed. 2008, 47, 1.

(102) Doyle, M. P.; Choi, H. Org. Lett. 2007, 9, 5349.

(103) Mchaughlin, E. C.; Doyle, M. P. J. Org. Chem. 2008, 73, 4317.

(104) Che, C. M.; Jiang, G.; Chen, J.; Thu, H. Y.; Huang, J. S.; Zhu, N. Angw. Chem. Int. Ed. 2008, 47, 1.

(105) Cornil, B.I; Herrmannn, W. A. *Catalysis from A to Z*; Schlogl, R., Ed,: John Wiley & Sons, 2008.

(106) Brunner, H.; Obermann, U.; Wimmer, P. J. Organomet. Chem. 1986, 316, C1.

(107) McManus, H.; Guiry, P. J. Chem. Rev. 2004, 104, 4151.

(108) For examples of Ru(III)-porphyrin complexes in alkane oxidation reactions, see:

Ohtake, H.; Higuchi, T.; and Hirobe, M. J. Am. Chem. Soc. 1992, 114, 10660-10662. (b)

Groves, J. T.; Bonchio, M.; Carofiglio, T.; Shalyaev, K. J. Am. Chem. Soc. 1996, 118, 8961-8962.

- (109) Kher, S. M.; Kulkami, G. H. Indian J. Chem. Section B 1989, 28B, 675.
- (110) Oku, A.; Abe, M.; Iwamoto, M. J. Org. Chem. 1994, 59, 7445.
- (111) Van, D. W.; Anton J. A.; Cerfontain, H. Tetrahedron 1981, 37, 2121.





Figure A1. Molecular Structure of 90.



Figure A2. Molecular Structure of 91.



Figure A3. Molecular Structure of 92.



Figure A4. Molecular Structure of 93.



Figure A5. Molecular Structure of 94.



Figure A6. Molecular Structure of 95.



*Figure A7*. Molecular Structure of **96**.



*Figure A8*. Molecular Structure of **97**.



Figure A9. Molecular Structure of 98.



Figure A10. Molecular Structure of 99.

## Appendix B

Table B1. Crystal Data and Structure Refinement for 90.

Identification code	yiha	
Empirical formula	$C_{56}H_{79}F_6NO_3P_2Ru$	
Formula weight	1091.21	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Triclinic	
Space group	P -1	
Unit cell dimensions	a = 10.9483(2) Å	$\alpha = 84.2760(10)^{\circ}$
	b = 11.7453(2) Å	$\beta = 75.6230(10)^{\circ}$
	c = 21.8972(4) Å	$\gamma = 81.6590(10)^{\circ}$
Volume	2692.78(8) Å <sup>3</sup>	
Z	2	
Density (calculated)	1.346 Mg/m <sup>3</sup>	
Absorption coefficient	3.448 mm <sup>-1</sup>	
F(000)	1148	
Crystal size	0.30 x 0.19 x 0.12 mm <sup>3</sup>	
Theta range for data collection	2.09 to 67.19°.	
Index ranges	$-12 \le h \le 13, -13 \le k \le 13, 0 \le l \le 25$	
Reflections collected	22577	
Independent reflections	8864 [R(int) = 0.0172]	
Completeness to theta = $67.19^{\circ}$	92.0 %	
Absorption correction	Numerical	
Max. and min. transmission	0.6824 and 0.4244	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	8864 / 0 / 630	
Goodness-of-fit on F <sup>2</sup>	1.148	
Final R indices [I>2sigma(I)]	$R_1 = 0.0305, wR_2 = 0.073$	5
R indices (all data)	$R_1 = 0.0310, wR_2 = 0.073$	8
Largest diff. peak and hole	0.866 and -0.356 e.Å <sup>-3</sup>	

Identification code	yika	
Empirical formula	$C_{98}H_{82}BCl_3F_4N_2O_3P_4Ru_2\\$	
Formula weight	1853.72	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Triclinic	
Space group	P -1	
Unit cell dimensions	a = 15.1928(3) Å	$\alpha = 67.2570(10)^{\circ}$
	b = 16.5925(4) Å	$\beta = 87.3490(10)^{\circ}$
	c = 20.8247(4)  Å	$\gamma = 63.8770(10)^{\circ}$
Volume	4297.92(14) Å <sup>3</sup>	
Z	2	
Density (calculated)	$1.432 \text{ Mg/m}^3$	
Absorption coefficient	4.902 mm <sup>-1</sup>	
F(000)	1895	
Crystal size	0.25 x 0.22 x 0.17 mm <sup>3</sup>	
Theta range for data collection	3.13 to 67.01°.	
Index ranges	$-17 {\leq} h {\leq} {17},  0 {\leq} k {\leq} {19},  0 {\leq} l {\leq} {24}$	
Reflections collected	35625	
Independent reflections	14081 [R(int) = $0.0253$ ]	
Completeness to theta = $67.01^{\circ}$	97.8 %	
Absorption correction	Numerical	
Max. and min. transmission	0.4895 and 0.3737	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	14081 / 10 / 1100	
Goodness-of-fit on F <sup>2</sup>	0.995	
Final R indices [I>2sigma(I)]	$R_1 = 0.0384, wR_2 = 0.100$	1
R indices (all data)	$R_1 = 0.0433,  wR_2 = 0.1027$	
Largest diff. peak and hole	0.532 and -0.589 e.Å <sup>-3</sup>	

Table B2. Crystal Data and Structure Refinement for 91.

Identification code	yika	
Empirical formula	$C_{52}H_{44}B_2F_8N_4P_2Ru$	
Formula weight	1061.54	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P 21/n	
Unit cell dimensions	a = 15.4945(4) Å	$\alpha = 90^{\circ}$
	b = 14.6409(4) Å	$\beta = 108.2400(10)^{\circ}$
	c = 21.9804(5) Å	$\gamma=90^\circ$
Volume	4735.8(2) Å <sup>3</sup>	
Z	4	
Density (calculated)	$1.489 \text{ Mg/m}^3$	
Absorption coefficient	3.946 mm <sup>-1</sup>	
F(000)	2160	
Crystal size	0.15 x 0.11 x 0.05 mm <sup>3</sup>	
Theta range for data collection	3.09 to 67.26°.	
Index ranges	$-18 \le h \le 17,  0 \le k \le 17,  0 \le l \le 26$	
Reflections collected	39472	
Independent reflections	8231 [R(int) = 0.0663]	
Completeness to theta = $67.26^{\circ}$	96.9 %	
Absorption correction	Numerical	
Max. and min. transmission	0.8271 and 0.5890	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	8231 / 0 / 626	
Goodness-of-fit on F <sup>2</sup>	1.045	
Final R indices [I>2sigma(I)]	$R_1 = 0.0417, wR_2 = 0.0926$	
R indices (all data)	$R_1 = 0.0583, wR_2 = 0.0984$	
Largest diff. peak and hole	0.458 and -0.557 e.Å <sup>-3</sup>	

*Table B3*. Crystal Data and Structure Refinement for 92.

Identification code	yima	
Empirical formula	$C_{18}H_{24}B_2F_8N_6ORu$	
Formula weight	615.12	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P 21/n	
Unit cell dimensions	a = 7.5577(2)  Å	$\alpha = 90^{\circ}$
	b = 9.8463(2) Å	$\beta = 90.0130(10)^{\circ}$
	c = 34.0339(9) Å	$\gamma=90^\circ$
Volume	2532.65(11) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.613 Mg/m <sup>3</sup>	
Absorption coefficient	$5.772 \text{ mm}^{-1}$	
F(000)	1232	
Crystal size	0.59 x 0.14 x 0.12 mm <sup>3</sup>	
Theta range for data collection	3.90 to 67.18°.	
Index ranges	$-8 \le h \le 8,  0 \le k \le 11,  0 \le l \le 39$	
Reflections collected	21068	
Independent reflections	4394 [R(int) = 0.0299]	
Completeness to theta = $67.18^{\circ}$	97.1 %	
Absorption correction	Numerical	
Max. and min. transmission	0.5443 and 0.1319	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	4394 / 10 / 346	
Goodness-of-fit on F <sup>2</sup>	1.072	
Final R indices [I>2sigma(I)]	$R_1 = 0.0201, wR_2 = 0.0521$	
R indices (all data)	$R_1 = 0.0201, wR_2 = 0.0521$	
Largest diff. peak and hole	0.438 and -0.350 e.Å <sup>-3</sup>	

Table B4. Crystal Data and Structure Refinement for 93.

Identification code	yin5	
Empirical formula	$C_{45}H_{60}Cl_4N_4O_2Ru_2$	
Formula weight	1032.91	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	C 2	
Unit cell dimensions	a = 30.0297(13)  Å	$\alpha = 90^{\circ}$
	b = 9.1898(4) Å	$\beta = 95.619(2)^{\circ}$
	c = 15.8658(7) Å	$\gamma=90^\circ$
Volume	4357.4(3) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.575 Mg/m <sup>3</sup>	
Absorption coefficient	8.202 mm <sup>-1</sup>	
F(000)	2120	
Crystal size	0.40 x 0.35 x 0.12 mm <sup>3</sup>	
Theta range for data collection	2.80 to 67.90°.	
Index ranges	$-35 \le h \le 35, -10 \le k \le 10, -18 \le l \le 18$	
Reflections collected	17156	
Independent reflections	7134 [ $\mathbf{R}(int) = 0.0674$ ]	
Completeness to theta = $67.90^{\circ}$	96.1 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.4394 and 0.1379	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	7134 / 1 / 516	
Goodness-of-fit on F <sup>2</sup>	1.037	
Final R indices [I>2sigma(I)]	$R_1 = 0.0492, wR_2 = 0.1305$	
R indices (all data)	$R_1 = 0.0493, wR_2 = 0.1308$	
Absolute structure parameter	0.017(11)	
Largest diff. peak and hole	1.565 and -0.830 e.Å <sup>-3</sup>	

*Table B5*. Crystal Data and Structure Refinement for 94.

Identification code	yioa	
Empirical formula	$C_{22}H_{24}Cl_2N_2ORu$	
Formula weight	504.40	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P 21	
Unit cell dimensions	a = 7.0471(6)  Å	$\alpha = 90^{\circ}$
	b = 16.9054(15) Å	$\beta = 99.8740(10)^{\circ}$
	c = 8.4172(8) Å	$\gamma=90^\circ$
Volume	987.92(15) Å <sup>3</sup>	
Z	2	
Density (calculated)	1.696 Mg/m <sup>3</sup>	
Absorption coefficient	1.080 mm <sup>-1-1</sup>	
F(000)	512	
Crystal size	0.18 x 0.14 x 0.10 mm <sup>3</sup>	
Theta range for data collection	2.41 to 32.12°.	
Index ranges	$-10 \le h \le 10,  -25 \le k \le 25,  0 \le l \le 12$	
Reflections collected	16325	
Independent reflections	6407 [R(int) = 0.0182]	
Completeness to theta = $32.12^{\circ}$	95.1 %	
Absorption correction	Numerical	
Max. and min. transmission	0.8997 and 0.8293	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	6407 / 1 / 349	
Goodness-of-fit on F <sup>2</sup>	1.032	
Final R indices [I>2sigma(I)]	$R_1 = 0.0179, wR_2 = 0.0446$	
R indices (all data)	$R_1 = 0.0183, wR_2 = 0.0448$	
Absolute structure parameter	-0.015(14)	
Largest diff. peak and hole	1.004 and -0.294 e.Å <sup>-3</sup>	

*Table B6*. Crystal Data and Structure Refinement for **95**.

Identification code	yipa	
Empirical formula	$C_{18}H_{24}Cl_2N_2ORu$	
Formula weight	456.36	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	C 2	
Unit cell dimensions	a = 30.3970(13) Å	$\alpha = 90^{\circ}$
	b = 8.7145(4) Å	$\beta = 96.9560(10)^{\circ}$
	c = 15.8881(7)  Å	$\gamma=90^\circ$
Volume	4177.7(3) Å <sup>3</sup>	
Z	8	
Density (calculated)	1.451 Mg/m <sup>3</sup>	
Absorption coefficient	1.013 mm <sup>-1</sup>	
F(000)	1856	
Crystal size	0.44 x 0.38 x 0.28 mm <sup>3</sup>	
Theta range for data collection	1.29 to 32.05°.	
Index ranges	$-45 \le h \le 43, -12 \le k \le 12, 0 \le l \le 23$	
Reflections collected	34398	
Independent reflections	13508 [R(int) = $0.0161$ ]	
Completeness to theta = $32.05^{\circ}$	95.1 %	
Absorption correction	Numerical	
Max. and min. transmission	0.7646 and 0.6642	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	13508 / 1 / 497	
Goodness-of-fit on F <sup>2</sup>	1.007	
Final R indices [I>2sigma(I)]	$R_1 = 0.0198, wR_2 = 0.0520$	
R indices (all data)	$R_1 = 0.0208, wR_2 = 0.0527$	
Absolute structure parameter	-0.028(14)	
Largest diff. peak and hole	0.920 and -0.372 e.Å <sup>-3</sup>	

*Table B7*. Crystal Data and Structure Refinement for 96.

Identification code	yita		
Empirical formula	$C_{26}H_{34}BCl_8F_4N_4O_{2.37}R_1$	u	
Formula weight	911.89		
Temperature	100(2) K		
Wavelength	1.54178 Å		
Crystal system	Orthorhombic		
Space group	P 21 21 21		
Unit cell dimensions	a = 11.19480(10) Å	$\alpha = 90^{\circ}$	
	b = 13.9654(2) Å	$\beta = 90^{\circ}$	
	c = 23.7007(3)  Å	$\gamma=90^\circ$	
Volume	3705.36(8) Å <sup>3</sup>		
Z	4		
Density (calculated)	1.635 $Mg/m^3$		
Absorption coefficient	9.200 mm <sup>-1</sup>		
F(000)	1832		
Crystal size	$0.36 \ge 0.20 \ge 0.10 \text{ mm}^3$	1	
Theta range for data collection	3.67 to 68.08°.		
Index ranges	$-13 \le h \le 13, 0 \le k \le 16$	5, $0 \le l \le 28$	
Reflections collected	31221		
Independent reflections	6686 [R(int) = 0.0294]		
Completeness to theta = $68.08^{\circ}$	99.1 %		
Absorption correction	Numerical		
Max. and min. transmission	0.4598 and 0.1364		
Refinement method	Full-matrix least-square	es on F <sup>2</sup>	
Data / restraints / parameters	6686 / 0 / 432		
Goodness-of-fit on F <sup>2</sup>	1.002		
Final R indices [I>2sigma(I)]	$R_1 = 0.0271, wR_2 = 0.0$	663	
R indices (all data)	$R_1 = 0.0279, wR_2 = 0.0$	$R_1 = 0.0279, wR_2 = 0.0667$	
Absolute structure parameter	-0.019(7)		
Extinction coefficient	0.00015(3)		
Largest diff. peak and hole	1.269 and -0.799 e.Å <sup>-3</sup>		

*Table B8*. Crystal Data and Structure Refinement for 97.

Identification code	yiwa	
Empirical formula	$C_{22} H_{26} Cl_8 Fe N_4 O_2$	
Formula weight	717.92	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P 21/c	
Unit cell dimensions	a = 15.6717(2)  Å	$\alpha = 90^{\circ}$
	b = 10.50400(10) Å	$\beta = 107.3650(10)^{\circ}$
	c = 19.0499(2) Å	$\gamma=90^\circ$
Volume	2992.98(6) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.593 Mg/m <sup>3</sup>	
Absorption coefficient	10.854 mm <sup>-1</sup>	
F(000)	1456	
Crystal size	0.33 x 0.27 x 0.22 mm <sup>3</sup>	
Theta range for data collection	4.86 to 67.95°.	
Index ranges	$-18 \le h \le 16, 0 \le k \le 12, 0 \le l \le 22$	
Reflections collected	25014	
Independent reflections	5242 [R(int) = 0.0319]	
Completeness to theta = $67.95^{\circ}$	96.3 %	
Absorption correction	Numerical	
Max. and min. transmission	0.1987 and 0.1241	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	5242 / 0 / 339	
Goodness-of-fit on F <sup>2</sup>	1.014	
Final R indices [I>2sigma(I)]	$R_1 = 0.0271, wR_2 = 0.0659$	
R indices (all data)	$R_1 = 0.0314, wR_2 = 0.0675$	
Extinction coefficient	0.00011(3)	
Largest diff. peak and hole	0.259 and -0.207 e.Å <sup>-3</sup>	

*Table B9*. Crystal Data and Structure Refinement for 98.

Identification code	yi1ca	
Empirical formula	$C_{43}H_{54}B_2Cl_8F_8Fe_2N_8O_4\\$	
Formula weight	1315.86	
Temperature	100(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	P 21	
Unit cell dimensions	a = 12.3494(4) Å	$\alpha = 90^{\circ}$
	b = 18.2325(6) Å	$\beta=92.442(2)^\circ$
	c = 12.6916(4) Å	$\gamma = 90^{\circ}$
Volume	2855.05(16) Å <sup>3</sup>	
Z	2	
Density (calculated)	1.531 Mg/m <sup>3</sup>	
Absorption coefficient	8.160 mm <sup>-1</sup>	
F(000)	1340	
Crystal size	0.57 x 0.18 x 0.15 mm <sup>3</sup>	
Theta range for data collection	3.49 to 67.90°.	
Index ranges	$\text{-}14 \leq h \leq 14,  \text{-}21 \leq k \leq 21,  0 \leq l \leq 15$	
Reflections collected	23677	
Independent reflections	9781 [R(int) = 0.0396]	
Completeness to theta = $67.90^{\circ}$	97.9 %	
Absorption correction	Numerical	
Max. and min. transmission	0.3801 and 0.0898	
Refinement method	Full-matrix least-squares on F <sup>2</sup>	
Data / restraints / parameters	9781 / 46 / 735	
Goodness-of-fit on F <sup>2</sup>	1.028	
Final R indices [I>2sigma(I)]	$R_1 = 0.0622, wR_2 = 0.152$	21
R indices (all data)	$R_1 = 0.0811, wR_2 = 0.1652$	
Absolute structure parameter	0.001(6)	
Largest diff. peak and hole	0.886 and -0.452 e.Å $^{\text{-3}}$	

Table B10. Crystal Data and Structure Refinement for 99.