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Advanced management of the Mexican rice borer (*Eoreuma loftini*) in sugarcane

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**ADVANCED MANAGEMENT OF THE
MEXICAN RICE BORER (*EOREUMA LOFTINI*)
IN SUGARCANE**

A Thesis

**Submitted to the Graduate Faculty of the
Louisiana State University and
Agricultural and Mechanical College
In partial fulfillment of the
requirements for the degree of
Master of Science**

in

The Department of Entomology

**by
Blake E. Wilson
B.S., Louisiana State University, 2009
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ABSTRACT

Insecticide, greenhouse and varietal resistance experiments were conducted in Texas to develop management tactics for the Mexican rice borer, *Eoreuma loftini* (Dyar), sugarcane. A 3-treatment, large plot aerial application study was set up in 5 commercial sugarcane fields (35-84 acres) to evaluate the utilization of pheromone traps to improve chemical control strategies for *E. loftini* during 2009 and 2010 growing seasons. A threshold of 20-25 moths/trap/wk was used as an indicator to initiate monitoring for *E. loftini* larval infestations. Larval infestations were directly related to the no. of moths/trap/wk ($R^2=0.71$). Reductions in borer injury and adult emergence (60% and 40% for novaluron and β -cyfluthrin, respectively) were detected when a threshold of 5% of stalks with treatable larvae was used for timing an insecticide application. Data revealed novaluron treatments increased sugar production by 14%. A greenhouse study assessed the establishment and behavior of *E. loftini* neonates on two phenological stages of stalkborer resistant (HoCP 85-845) and susceptible (HoCP 00-950) cultivars. Approximately half (55%) of neonates on HoCP 00-950 and 28% on HoCP 85-845 tunneled inside the leaf mid-ribs within 1d of eclosion. Duration of neonate exposure ranged from 3.5 - 6.4 d. This research shows a short window of vulnerability of *E. loftini* to insecticide applications, and demonstrates the potential to use pheromone traps and new chemistries for enhancing chemical control. A 5-replication field test evaluated stalkborer resistance in 25 sugarcane cultivars. Differences were detected between cultivars in *E. loftini* injury which ranged from 1.0-20.3% bored. The resistant standard HoCP 85-845 and a South African cultivar, N-21, were the most resistant. HoCP 96-540, which represents the majority of sugarcane acreage in Louisiana, was among the most susceptible. Assessment of stalkborer resistance in sugarcane cultivars is needed as host plant resistance will continue to be important to *E. loftini* IPM.

CHAPTER 1

INTRODUCTION AND REVIEW OF LITERATURE

Introduction

Management of invasive species is a growing concern in the United States. Currently there are approximately 50,000 invasive species in the U.S. responsible for \$137 billion in damages and control costs, annually (Pimentel *et al.* 2000). Of those species, approximately 1,000 are crop pests which account for \$14.4 billion annually in damages (Pimentel *et al.* 2000). One invasive insect that has become established as a major pest of sugarcane and rice in Texas is the Mexican rice borer, *Eoreuma loftini* (Dyar). This species was first reported as a pest of sugarcane in the U.S. in the Lower Rio Grande Valley (LRGV) of Texas in 1980 (Johnson and Van Leerdaam 1981), where it now accounts for >95% of the sugarcane stalkborer population (Legaspi *et al.* 1999a). It has since spread northeast through the Texas rice belt along the Gulf Coast (Reay-Jones *et al.* 2007a). Despite a 2005 quarantine designed to prevent movement of Texas sugarcane into Louisiana, *E. loftini* was discovered in Louisiana in December 2008 (Hummel *et al.* 2010). Based on its current 16.5 km/yr rate of expansion, *E. loftini* is predicted to infest the entire state by 2035 when it is projected to cause >\$260 million in annual revenue loss to Louisiana agriculture (Reay-Jones *et al.* 2008). As *E. loftini* threatens Louisiana, the need to develop effective control tactics is of growing importance to Louisiana agriculture.

Management of *E. loftini* in sugarcane relies on a balance of multiple control tactics. Reay-Jones *et al.* (2005) showed that a combination of irrigation, cultivar resistance, and insecticide applications greatly reduced *E. loftini* injury. However, *E. loftini* control is complicated by the tunneling behavior of larvae limiting exposure to insecticides, beneficial insects, and other control tactics (Van Leerdaam 1986, Meagher *et al.* 1994). Biological control has been largely unsuccessful against *E. loftini* in the LRGV (Legaspi *et al.* 1997, Meagher *et al.* 1998) despite releases of multiple parasitoids (Browning and Melton 1987). In addition, the efficacy of

insecticide applications is often inadequate to improve subsequent sugar yields and chemical control of *E. loftini* in sugarcane is not economical (Meagher *et al.* 1994, Legaspi *et al.* 1997, Reay-Jones *et al.* 2005). The lack of adequate chemical *E. loftini* control in sugarcane is most often attributed to the insufficient exposure of larvae (Johnson *et al.* 1985, Meagher *et al.* 1994). Plant characteristics which affect larval establishment such as rind hardness and nutritional factors vary between cultivars (Posey *et al.* 2006, Reay-Jones *et al.* 2005, 2007b, Showler and Castro 2009) and phenological stage (Atkinson and Nuss 1989, Reay-Jones *et al.* 2007b). A better understanding of *E. loftini* neonate behavior, improved timing of insecticide applications, and varietal resistance could lead to the development of management strategies which effectively control larvae before they become protected within the stalk. This research attempts to evaluate: (1) the use of pheromone traps to improve chemical control of *E. loftini* by increasing scouting efficiency and enhancing the timing of insecticide applications; (2) the effect of a single aerial application of novaluron on borer injury and sugarcane yield in a commercial setting; (3) neonate establishment, feeding behavior, and exposure to control agents on different sugarcane cultivars and phenological growth stages, and (4) cultivar resistance to *E. loftini* among 25 commercial and experimental sugarcane varieties.

Literature Review

Eoreuma loftini was first reported in the U.S. in Arizona (Dyar 1917), but did not become established in the LRGV of Texas until 1980 (Johnson and Van Leerdam 1981). It has since become the dominant pest of sugarcane in the LRGV where it now accounts for >95% of the sugarcane stalkborer population (Legaspi *et al.* 1999a). Stalkborers are the primary pest of sugarcane in the LRGV and are responsible for damaging ~20% of sugarcane internodes annually (Legaspi *et al.* 1999a). *Eoreuma loftini* is a devastating pest of sugarcane responsible

for >\$10 million in annual revenue losses (Legaspi *et al.* 1997) and yield losses of 50-65% have been attributed to *E. loftini* infestations in the LRGV (Johnson 1984). Shortly after establishment, sugarcane was the source of 97% of all *E. loftini* collected in the LRGV (Johnson 1984). However, recent evidence suggests other crops including rice, corn, and sorghum as well as certain weed hosts may also be important to *E. loftini* ecology (Beuzelin *et al.* 2010b). The range of *E. loftini* has since expanded north-easterly through the Texas rice belt along the Gulf Coast (Reay-Jones *et al.* 2007a). Despite a 2005 quarantine preventing the entry of Texas sugarcane into Louisiana, *E. loftini* was discovered in the state in December of 2008 (Hummel *et al.* 2010). The continued spread of *E. loftini* will have serious economic consequences to Louisiana agriculture which emphasizes the need for effective area-wide population management. Based on the current rate of expansion, 16.5 km/yr, *E. loftini* is predicted to infest the entire Louisiana sugarcane industry by 2035 (Reay-Jones *et al.* 2008). Its establishment as an economic pest in Louisiana sugarcane is predicted to cause \$220 million and \$48 million in annual revenue loss for the sugarcane and rice industries, respectively, by 2035 (Reay-Jones *et al.* 2008). The need to develop *E. loftini* IPM strategies which reduce crop injury as well as area-wide populations is of critical importance to the state of Louisiana.

Inability to provide effective pest control of *E. loftini* in sugarcane can be attributed to the pest's biology which limits exposure to control tactics. Female *E. loftini* oviposit in sheltered sites on dried sugarcane leaves located on the lower portion of the plant, i.e. between the soil surface and 80 cm height (Van Leerdam *et al.* 1984, 1986, Showler and Castro 2010). The globular cream-colored eggs are usually laid in groups of 5 to 100. Temperature is directly related to total fecundity, with an average of 259 eggs per female at 20°C and 406 eggs at 26°C. The oviposition rate varies from 29 eggs/d at 20°C to 64 eggs/d at 32°C (Van Leerdam 1986).

Oviposition peaks 2 d after female emergence. Increasing temperature appears to decrease the time between emergence and oviposition (Van Leerdam 1986). While oviposition is an important factor affecting *E. loftini* ecology, it is the larval characteristics which complicate pest management.

Extensive laboratory examination of *E. loftini* development was conducted by Van Leerdam (1986). Newly hatched *E. loftini* larvae disperse from dry leaves where the eggs are deposited to green parts of the plant where they mine into leaf sheaths and stalks, and become protected from beneficial insects and insecticides. *Eoreuma loftini* completes the egg stage in 14 d at 20°C and 5 d at 32°C when reared at constant temperatures (Van Leerdam 1986). In appearance, the larvae have an orange/brown head capsule and four dark broken lines that run the length of the light-colored body. Laboratory examination revealed developmental polymorphism, with five to seven instars occurring before pupation. The number of stadia is affected by sex, with a higher average for females (six) than for males (five) (Van Leerdam 1986). Temperature is inversely related to this number, with a six stadia larval development at 23°C, and five stadia at 29°C. The mean duration of each stadium decreases rapidly as temperature increases, with an average of 78 d at 20°C and 21 d at 32°C for completion of all larval stages (Van Leerdam 1986). Van Leerdam (1986) found that *E. loftini* larvae typically feed in the leaf sheath until becoming third instars when they enter the stalk and begin to feed internally. In addition, the average age of third instars reared on artificial diet is roughly 11 d. However, these studies were conducted under artificial laboratory conditions and may not be representative of *E. loftini* behavior on live sugarcane stalks. Once inside the stalk *E. loftini* larvae tunnel both vertically and horizontally (Legaspi *et al.* 1997). Mature larvae create an emergence window in the stalk prior to pupation which takes place in frass-packed tunnels. This behavior creates a protected chamber for

development from stalk entry until adult emergence (Legaspi *et al.* 1997). The duration of the *E. loftini* pupal stage is also inversely related to temperature and lasts from 21 d (20°C) to 7 d at 32°C (Van Leerdam 1986). The developmental times of *E. loftini* on sugarcane stalk sections showed extended durations compared with those obtained from development on artificial diet, suggesting that sugarcane may be a less than optimal food source (Van Leerdam 1986). Variation in the duration of larval development between different cultivars (Kennedy and Kishaba 1976) may be related to nutritional content such as concentrations of free amino acids (FAA) (Reay-Jones *et al.* 2005) and other nutrients. Management in the LRGV is further complicated by overlapping generations which mean multiple life stages are present simultaneously. Studies have shown that *E. loftini* is active throughout the year in the LRGV, and active larval and adult stages have been observed throughout the winter (Johnson 1985, Van Leerdam *et al.* 1986, Beuzelin *et al.* 2010b).

The life stage of *E. loftini* targeted for chemical control is the neonate, which migrates from the oviposition site on dry leaves at the base of the plant to green parts of the plant (Meagher *et al.* 1994). Weekly applications of insecticides have reduced *E. loftini* injury, but the effect on sugarcane yield was rarely significant (Johnson 1985, Meagher *et al.* 1994, Legaspi *et al.* 1999b, Reay-Jones *et al.* 2005). Insecticides therefore are believed to have had such limited success in controlling *E. loftini* that most sugarcane growers in the LRGV have abandoned this control approach altogether (Legaspi *et al.* 1997). Similar to the sugarcane borer, *Diatraea saccharalis* F., *E. loftini* injury is most commonly expressed as a proportion of bored internodes. Key yield and quality parameters such as recoverable sugar, juice purity, and sucrose content are inversely related to percentage of bored internodes (White and Hensley 1987, Legaspi *et al.* 1999a). However, past failures to detect effects of insecticide treatments on yield despite a reduction in

percentage of bored internodes (Johnson 1985, Meagher *et al.* 1994, Legaspi *et al.* 1999b, and Reay-Jones *et al.* 2005) may be the result of high variability of sugarcane yield studies and differential responses to injury between cultivars (Hensley and Long 1969).

The narrow temporal window during which *E. loftini* larvae are potentially exposed to insecticides reduces the impact of chemical control. The difficulty of applying pesticides to foliage in the lower parts of sugarcane further reduces the efficiency. The potential to improve chemical control may lie in the use of insect growth regulators, as well as an effort to achieve better insecticide application timing. Tebufenozide has shown excellent efficacy against sugarcane stalkborers in both laboratory (Rodriguez *et al.* 2001) and field tests (Rodriguez *et al.* 1995, Reay-Jones *et al.* 2005). Laboratory studies revealed that tebufenozide is toxic to *E. loftini*, even though it is slower acting than traditionally used pyrethroid insecticides (Legaspi *et al.* 1999b). However, even when multiple applications of tebufenozide are made, the effect on sugar yield has not been significant (Reay-Jones *et al.* 2005). In addition, laboratory studies have shown there is potential for development of insecticide resistance to tebufenozide which highlighted the need for alternative chemistries (Akbar *et al.* 2008b). A recently labeled (“section 3” for sugarcane) insect growth regulator, novaluron, is a promising new chemistry which has also been shown to be effective in controlling stalkborers in sugarcane (Akbar *et al.* 2008a, 2009, Beuzelin *et al.* 2005, 2010). However, *E. loftini* control with novaluron using aerial application technology in a commercial setting has not yet been evaluated.

In addition to new chemistries, control of *E. loftini* could be improved by more precise timing of insecticide applications. In order to maximize efficiency, insecticide applications should be made when high densities of larvae are exposed and feeding on the external surfaces of stalks. Well timed insecticide applications may not only improve efficacy, but also reduce the

number of treatments needed. Insecticidal control of the sugarcane borer has been greatly improved by the utilization of scouting thresholds (Hensley 1971). Adapted from the sugarcane borer work in Louisiana, a treatment threshold of 5% of stalks with larvae present in the leaf sheaths (or exposed on the surface of the plant) has been used to trigger insecticide applications for control of *E. loftini* (Johnson *et al.* 1985, Meagher *et al.* 1994). However, scouting for *E. loftini* in sugarcane is labor intensive and requires a large number of stalks to be sampled to provide accurate estimations of larval infestations (Hall 1986, Meagher *et al.* 1996b). Previous studies (Meagher *et al.* 1996b) suggest that identification of relationships between adult population density estimations such as pheromone trap catches and larval injury might be used to improve early detection of pest population increases. In a small plot study (Reagan *et al.* 2001), the use of pheromone traps and treatment thresholds to direct insecticide applications showed potential to enhance chemical control of *E. loftini*.

Brown *et al.* (1988) were the first to provide evidence of the *E. loftini* female sex pheromone. The pheromone was subsequently synthesized (Shaver *et al.* 1988), and field experiments demonstrated the efficiency of pheromone-baited traps as a survey and monitoring tool (Shaver *et al.* 1990, 1991). Pheromone traps have been used to track the movement of *E. loftini* across the Texas rice production area (Reagan *et al.* 2005, Reay-Jones *et al.* 2007a) and provided the first detection of the invasive insect in Louisiana (Hummel *et al.* 2010). Pheromone traps are useful in monitoring population fluctuations and may provide improved early warning signs of pest outbreaks (Robacker and Landholt 2002). By monitoring adult population fluctuations, pheromone traps have potential to reduce scouting effort and improve insecticide application timing. The use of pheromone traps to assist scouting has yet to be evaluated in a commercial setting. However, even if chemical control of *E. loftini* is improved, management programs will

not be able to rely solely on insecticide applications. The use of multiple control tactics is likely the only method to achieve adequate control of this destructive pest (Reay-Jones *et al.* 2005).

The use of resistant cultivars may provide an important component of *E. loftini* IPM. In addition to reducing pest injury on an individual field basis, area-wide pest management aims to reduce population levels of the target organism over a large geographical area (Bessin *et al.* 1991).

Resistant cultivars have consistently shown reduced *E. loftini* injury when compared to susceptible cultivars (Pfannenstiel and Meagher 1991, Meagher *et al.* 1996a, Reay-Jones *et al.* 2003, 2005). Host plant resistance in *E. loftini* management can also aid in the reduction of area-wide populations. This led to the development of a moth production index based on adult emergence which compares the effects of treatments on area-wide populations (Bessin *et al.* 1990). Expansive acreage of susceptible varieties with elevated moth production increases endemic *E. loftini* populations and imposes additional pressure on the remaining acreage. Currently, the majority of sugarcane acreage in Louisiana is planted with *E. loftini* susceptible cultivars HoCP 96-540 and LCP 85-384 which had the greatest moth production in varietal resistance studies (Reay-Jones *et al.* 2003). In addition to reducing area-wide populations, resistant cultivars may improve the efficacy of other control tactics by impeding larval entry into the stalk. While host plant resistance has repeatedly been shown to be promising for *E. loftini* control, the mechanisms of resistance are not fully understood.

Host plant resistance to sugarcane stalkborers has been classified into four categories: (1) unattractiveness of plants to adults for oviposition, (2) plant characteristics unfavorable for larval establishment in the plant, (3) plant characteristics that inhibit or retard larval development, and (4) plant tolerance (Mathes and Charpentier 1969). While the importance of each component of resistance is continually being evaluated, the majority of work has been focused on *E. loftini*

oviposition. Oviposition preference may play a role in resistance because *E. loftini* prefers folds in senescing leaves as oviposition sites (Van Leerdaam 1986, Reay-Jones *et al.* 2007b, Showler and Castro 2010a,b). Plant vigor characteristics which minimize leaf senescence and attractiveness for egg laying may contribute to *E. loftini* resistance (Reay-Jones *et al.* 2005, 2007b, Showler and Castro 2010a,b). Showler and Castro (2010b) demonstrated *E. loftini* prefers folds in curled leaves as oviposition sites which further limit exposure of eggs and neonates. Oviposition preference may be linked to chemoreceptors which detect the presence/absence of primary or secondary compounds to assist females in accepting or rejecting a host plant (Ramaswamy 1988). FAA concentrations, which are elevated in drought stressed sugarcane (Reay-Jones *et al.* 2005, Showler and Castro 2010a), may be responsible for *E. loftini* oviposition preference. Reay-Jones *et al.* (2007b) demonstrated that a positive correlation exists between free essential amino acids concentrations and eggs per plant. *Eoreuma loftini* females have shown an oviposition preference for susceptible cultivars and drought stressed sugarcane containing elevated concentrations of essential FAAs (Reay-Jones *et al.* 2007b, Showler and Castro 2010a). Factors which may hinder larval establishment are physical characteristics such as rind hardness and leaf sheath appression, which impede stalk entry prolonging larval exposure on plant surfaces (Coburn and Hensley 1972). Premature rind hardness may also be a component of this relationship (Martin *et al.* 1975). This potential mechanism of resistance may provide a key component to *E. loftini* IPM because it has potential to enhance the efficacy of other management tactics such as chemical control by increasing the duration of larval exposure. In addition to physical factors, host plant concentrations of certain primary and secondary metabolites affect larval development. Differences in *E. loftini* larval weight and time to pupation may be linked to varying levels of allelochemicals among sugarcane cultivars (Meagher

et al. 1996a). Evidence suggests that drought stress tolerance may be a factor in *E. loftini* resistance. Infestations of *E. loftini* are enhanced by plant stress which leads to an increase in FAA concentrations in sugarcane under drought stress (Reay-Jones *et al.* 2005, Showler and Castro 2010a). This increase in FAAs is less pronounced in resistant varieties compared to susceptible varieties indicating that drought tolerance might be a component of *E. loftini* resistance (Reay-Jones *et al.* 2005, Showler and Castro 2010a), but further research is needed before the roles of primary metabolites as well as other metabolic components in host plant resistance is fully understood. In addition, resistance mechanisms which impede entry into the stalk and may enhance the effects of other control tactics by prolonging the duration of larval vulnerability should be evaluated.

As *E. loftini* threatens to become established in Louisiana where it is expected to cause substantial economic losses (Reay-Jones *et al.* 2008), the need for effective management strategies is becoming more important. Because of the limited vulnerability of *E. loftini* larvae, continued examination of control tactics which target exposed neonates will contribute to development of improved *E. loftini* IPM programs. Potential control strategies highlighted by this research include the use of pheromone traps to assist scouting and improve application timing, utilization of new chemistries which are more effective against neonates, and the incorporation of resistant sugarcane varieties into *E. loftini* IPM.

CHAPTER 2

CHEMICAL CONTROL TACTICS THAT TARGET EXPOSED *E. LOFTINI* LARVAE IN SUGARCANE

Introduction

The Mexican rice borer, *Eoreuma loftini* (Dyar), is an invasive species originating from Mexico which became established in the Lower Rio Grande Valley (LRGV) of Texas in 1980 (Johnson and Van Leerdam 1981) and has since become the dominant pest of sugarcane in that area representing >95% of the sugarcane stalkborer population (Legaspi *et al.* 1999a) and causing >\$10 million in annual revenue losses (Legaspi *et al.* 1997). The range of *E. loftini* has expanded across the rice production area in east Texas (Browning *et al.* 1989, Reay-Jones *et al.* 2007a), eventually reaching Louisiana in 2008 (Hummel *et al.* 2010). Based on its current 16.5 km/yr rate of expansion, *E. loftini* is predicted to infest the entire Louisiana sugarcane industry by 2035 and is projected to cause up to \$268 million in annual revenue losses in sugarcane (\$220 million) and rice (\$48 million) (Reay-Jones *et al.* 2008).

While insecticide applications have been shown to reduce *E. loftini* injury, chemical control has rarely affected sugarcane yield (Johnson 1985, Meagher *et al.* 1994, Reay-Jones *et al.* 2005), and most LRGV producers abandoned insecticides as a means of management (Legaspi *et al.* 1997). However, new insecticide chemistries and improved scouting methods might enhance approaches to control. A recently labeled insect growth regulator, novaluron, is a promising new compound that was demonstrated to suppress *E. loftini* in sugarcane (Akbar *et al.* 2009).

Chemical control of *E. loftini* is inhibited because the larvae tunnel and pack it with frass, protecting them from exposure to topically applied insecticides. Thus, insecticide applications target neonates (1st to 3rd instars) not yet within tunnels. Overlapping generations in the LRGV result in multiple life stages of the pest simultaneously (Johnson 1985, Van Leerdam 1986, and Meagher *et al.* 1994).

Timing insecticide applications against stalkborers is dependent on scouting for treatable larval infestations. Modeled after sugarcane borer, *Diatraea saccharalis* F., work (Hensley 1971), a treatment threshold of 5% of stalks with *E. loftini* larvae on plant surfaces indicates the need for an insecticide application (Johnson *et al.* 1982). Scouting for *E. loftini* in sugarcane is labor intensive and it has been suggested that identification of a relationship between adult population density and larval injury could improve early detection of population increases (Meagher *et al.* 1996). Brown *et al.* (1988) studied the presence of the *E. loftini* female sex pheromone by examining male response to ovipositor extracts. Pheromone traps are effective at trapping male *E. loftini* (Shaver *et al.* 1990, 1991) and have been used to assist scouting by carefully timing insecticide applications (Reagan *et al.* 2001). In addition, a more thorough understanding of *E. loftini* larval behavior could lead to the development of control strategies which target vulnerable neonates.

Chemical *E. loftini* control is hindered by limited exposure of larvae. *Eoreuma loftini* shows a preference for oviposition on folds of dry leaf material (Van Leerdam 1984, Reay-Jones *et al.* 2007b, Showler and Castro 2010b), areas which are difficult to access with insecticides. After eclosion, early instars disperse and begin feeding on the green tissue of leaves and leaf sheaths before they enter the stalk and begin to feed internally (Van Leerdam 1986). Van Leerdam (1986) found larvae typically enter the stalk when they are third instars stalk and the average age of third instars reared on artificial diet is roughly 11d. Plant characteristics which affect larval establishment such as rind hardness and nutritional factors vary between cultivars (Posey *et al.* 2006, Reay-Jones *et al.* 2005, 2007b, Showler and Castro 2010a) and phenological growth stage (Atkinson and Nuss 1989, Reay-Jones *et al.* 2007b). The duration of larval feeding in the leaf sheath has been shown to be directly related to plant age (Van Leerdam 1986) and internodes

<70 d old are most susceptible to *E. loftini* injury (Ring *et al.* 1991) attributable to changes in physiology such as increasing rind hardness as internodes mature. Determination of the duration of leaf sheath feeding could improve the efficacy of scouting and action thresholds in *E. loftini* chemical control.

The objectives of this study are (1) to evaluate the use of pheromone traps to improve the timing of chemical control of *E. loftini* and enhance scouting efficiency, (2) to evaluate the efficacy of a single aerial application of novaluron on borer injury and sugarcane yield in a commercial setting, and (3) to determine the duration of *E. loftini* neonate exposure to control agents and to assess the effect of cultivar and phenological stage on neonate establishment, behavior, and survival.

Materials and Methods

Aerial Insecticidal Control. A 2009/2010 study was conducted using a randomized complete block experimental design with five replications (fields) in the LRGV (Cameron and Hidalgo Counties, TX). Insecticide treatments were assigned randomly to 4-ha plots in 5 commercial sugarcane fields ranging from 14-33 ha of variety CP 72-1210. Treatments were a single application of either novaluron (Diamond[®] 0.83EC, Makeshim Agan of North America Inc, Raleigh, NC) applied at 80 g (AI)/ha, β -cyfluthrin (Baythroid[®] XL, Bayer CropScience LP, Research Triangle Park, NC) applied at 25 g (AI) /ha, and another 5 plots were left as non-treated controls. Adult *E. loftini* population densities were monitored throughout the growing season with pheromone traps. Bucket type traps (one/field in 2009, two/field in 2010) baited with a synthetic *E. loftini* female sex pheromone lure (Luresept; Hercon Environmental, Emigsville, PA) attached to metal poles at a height of \approx 1 m above the ground were placed on opposite edges of experimental sugarcane fields. An insecticidal strip (Vaportape II; Hercon

Environmental) was placed inside traps to kill all insects trapped. Pheromone lures were replaced every 2 wk and insecticidal strips were replaced every 4 wk.

Traps were checked weekly from 15 July to 14 October 2009 and from 1 June to 14 August 2010. The number of male *E. loftini* caught per trap per week was recorded. Trap catches of >20–25 moths/trap/week were used as a scouting threshold to initiate monitoring for larval infestations. This threshold was developed from pheromone trap catch numbers collected in a small plot insecticide trial (Reagan *et al.* 2001). Larval infestations exceeding the economic threshold of 5% of stalks with treatable larvae present on plant surfaces initiated insecticide applications in all fields made by fixed wing aircraft flying at 233 kph equipped with CP-03 nozzles at 96 L/ha with less than 8 kph wind the mornings of 21 August 2009 and 14 August 2010. In 2010 weekly larval scouting was conducted by careful examination of 10 stalks (1 June–6 July) or 20 (13 July–14 August) from two locations several rows in from trap sites in all fields. Prior to harvest (28 October, 2009 and 8 November, 2010) two 15-stalk samples were collected from each plot and the numbers of bored internodes and emergences holes were recorded. Treatments plots were harvested separately and each load was weighed to determine the total weight of cane from each plot. The number of loads taken from each plot was variable, and all plots were completely harvested. A core sample from each load was weighed and prepared for quality analysis. A hydraulic press was used to extract juice. Brix, percent soluble solids in juice, was recorded using a brix refractometer. Percent sucrose in juice (Pol) was measured with an automatic sucrolyser after clarifying the juice with acetate. The ratio of sucrose to all other dissolved solids (Brix) is referred to as juice purity and expressed as a percentage. Tons of cane per acre (TCA) was calculated by dividing the total weight of cane (tons) harvested from each plot by the plot size (acres). Commercially recoverable sugar (CRS)

was recorded for each core sample and extrapolated to one ton of cane which is expressed as lbs of sugar/ton of cane. Yield was further extrapolated to tons of sugar per acre (TSA) calculated by the following: $TSA = (\text{Mean CRS} * TCA) / 2000$. Data were analyzed with generalized linear mixed models (Proc GLIMMIX, SAS Institute 2006) with Gaussian distributions. Tukey's honestly significant difference (HSD) was used for mean separation for all analyses except for sugar/ha and cane/ha which were separated with Fisher's least significant difference (LSD) test due to low degrees of freedom. All reported means were converted to metric units after analysis.

The total number of internodes, bored internodes, and emergence holes from stalks were summed for each 15 stalk sample. Data from 2009 and 2010 were analyzed together with year, field, field X year, and field X yield X treatment as random effects (Appendix B). Injury data, proportion of bored internodes, and relative survival to adulthood were analyzed using a generalized linear mixed model (Proc GLIMMIX, SAS Institute 2006) with a binomial distribution. Numbers of adult emergence holes were analyzed using a generalized linear mixed model with a Poisson distribution (Poisson 1837). A relative index was used to estimate survival of larvae to adulthood (relative survival = no. emergence holes/no. bored internodes) (Bessin *et al.* 1990, Reay-Jones *et al.* 2003). For all models, the Kenward-Roger method (Kenward and Roger 1997) was used to compute denominator degrees of freedom for the test of fixed effects for all variables. In addition, a simple linear regression between the numbers of male *E. loftini*/trap/wk and the percentages of stalks infested with treatable larvae was performed.

Neonate Establishment and Behavior. A greenhouse study was conducted during the summer of 2010 at the USDA-ARS Kika de la Garza Subtropical Agricultural Research Center in Weslaco, TX to investigate *E. loftini* neonate establishment and feeding behavior on two phenological stages of a resistant sugarcane variety, HoCP 85-845, and a susceptible variety,

HoCP 00-950 (Reay-Jones *et al.* 2005, Reagan *et al.* 2007). Twenty-four sugarcane nodes of each cultivar were obtained from Certis U.S.A. sugarcane tissue cultures. Plants were arranged on greenhouse tables in a completely randomized design with a 2 x 2 factorial, cultivar x phenological stage, with each of the four treatments representing 12 stalks. All stalks pieces were planted in late spring in 7.6 L pots in Sunshine mix no.1 nursery potting soil (~75% sphagnum peat moss, perlite, dolomitic limestone, and gypsum; Sungro Horticulture, Bellevue, Washington). Plants were kept well watered throughout their growth and 200 ml of Peters Professional (Scotts-Sierra Horticulture Products Company, Marysville, Ohio) water-soluble general purpose fertilizer was applied to the soil once plants reached the 2-leaf stage.

The experiment was conducted on immature cane once stalks had produced 6 nodes, 14 June – 2 July and on mature cane from 30 July–17 August once stalks had 12 nodes. *Eoreuma loftini* eggs were manually placed on sugarcane stalks in locations consistent with normal oviposition activity 15–25 cm from the stalk on the underside of sugarcane leaves which showed early signs of senescence to simulate natural *E. loftini* oviposition (Reay-Jones *et al.* 2007b, Showler and Castro 2010a,b). Eggs were obtained from a laboratory colony reared from *E. loftini* larvae collected from commercial sugarcane fields in Hidalgo County, TX on artificial diet (Martinez *et al.* 1988) at 25°C, 65% RH, and a photo period of 14:10 (L:D). After mating, egg masses of 10-80 eggs were deposited by the *E. loftini* females on to ½-inch-wide paper strips. Eggs on each strip were counted prior to clipping strips to leaves with 1-inch paper clips.

Development of early instar larvae was examined by direct observation and stalk dissection. Egg strips were removed after hatching, 7 d after strips were clipped to leaves, and the numbers of unhatched eggs were counted under a microscope. The location of initial establishment was recorded as either sheath feeding or mid-rib entry. The number and position of mid-rib entry

holes was recorded. All leaves and leaf sheaths on each plant were examined daily for the presence of *E. loftini* neonates, and the location of feeding sites (mid-rib or sheath), dispersal distance from oviposition sites, and time to stalk entry were recorded. The percentage of larvae that became established on each stalk was calculated by dividing the number of larvae observed feeding in leaves or sheaths by the number of hatched eggs. Dispersal of neonates, expressed as the number of internodes moved from oviposition sites, was recorded for all established larvae. Neonates which became established feeding within the leaf sheath were monitored daily by carefully checking between the stalk and sheath for the presence of larvae. Daily examination of each sheath was conducted until entry holes were observed or larvae were recorded as dead or vanished. Survival to stalk entry and duration of leaf sheath feeding (time from eclosion to stalk boring) were recorded. After allowing 4 wk for development, stalks were destructively sampled and the numbers and locations of entry holes and live larvae and pupa recorded. Data was analyzed using generalized linear mixed models (Proc GLIMMIX, SAS Institute 2006) (Appendix C). The proportion of larvae which became established on the stalk, the proportion which entered into the midrib of the leaf, and the proportion of larvae which survived to stalk entry were analyzed with binomial distributions because data are expressed as proportions of eclosed or established neonates. A Gaussian distribution was used to analyze data on the duration of larval exposure, and dispersal distance. Data were not transformed because response variables were dependent on the number of eggs pinned on each plant.

Results

Aerial Insecticidal Control. Pheromone trap catches in both 2009 and 2010 peaked in late August. Live larval infestations were sampled from ten stalks per plot and ranged from 5% to 32% with a mean of 13.8 ± 1.8 [SE] % of stalks with treatable larvae present on plant surfaces in

various fields on 20 Aug 2009 just prior to insecticide applications and a steady decline in the mean number *E. loftini*/trap/wk followed (Fig. 2.1A). On 14 August 2010 larval infestations ranged from 5 to 22.5% with a mean of 11.25 ± 1.5 [SE] % of stalks with treatable larvae. Weekly monitoring of larval infestations in 2010 depicted the relationship between adult population density and larval infestation (Fig. 2.1B). Linear regression revealed a substantial correlation ($F = 280.7$; $df = 1,114$; $P < 0.0001$, $r^2=0.71$) between pheromone trap catches (x) and larval infestation (y) which can be summarized by the equation, $y = 0.213x - 0.03833$ where x is equal to the number of male *E. loftini*/trap/wk and y = percentage of stalks infested with treatable larvae on plant surfaces.

Insecticide treatments significantly reduced the probability of occurrence of a bored internode by an average of 40.3% and 60.2% over both years for β -cyfluthrin and novaluron, respectively ($F = 11.41$; $df= 2,18.2$; $P = 0.0006$) (Fig. 2.2A). Insecticide applications reduced emergence holes per stalk by 37.4 and 58.4% over both years for β -cyfluthrin and novaluron, respectively ($F = 4.65$; $df = 2,17.2$; $P = 0.0244$) (Fig. 2.2B). Novaluron provided the best control in both years reducing injury (proportion of bored internodes) by 2.2-fold and 3.5-fold, and moth emergence by 1.7-fold and 4.3-fold for 2009 and 2010, respectively. β -cyfluthrin reduced the proportion of bored internodes by only 1.4-fold and 2.4-fold and adult emergence by 1.7-fold and 1.9-fold in 2009 and 2010, respectively. Mean relative survival to adulthood ranged from 0.237-0.260, however, differences between treatments were not detected. Insecticide applications reduced *E. loftini* injury and moth production to a greater degree in 2010 than in 2009. The probability of occurrence of a bored internode was 1.2-fold greater in 2010 (0.140) than in 2009 (0.124).

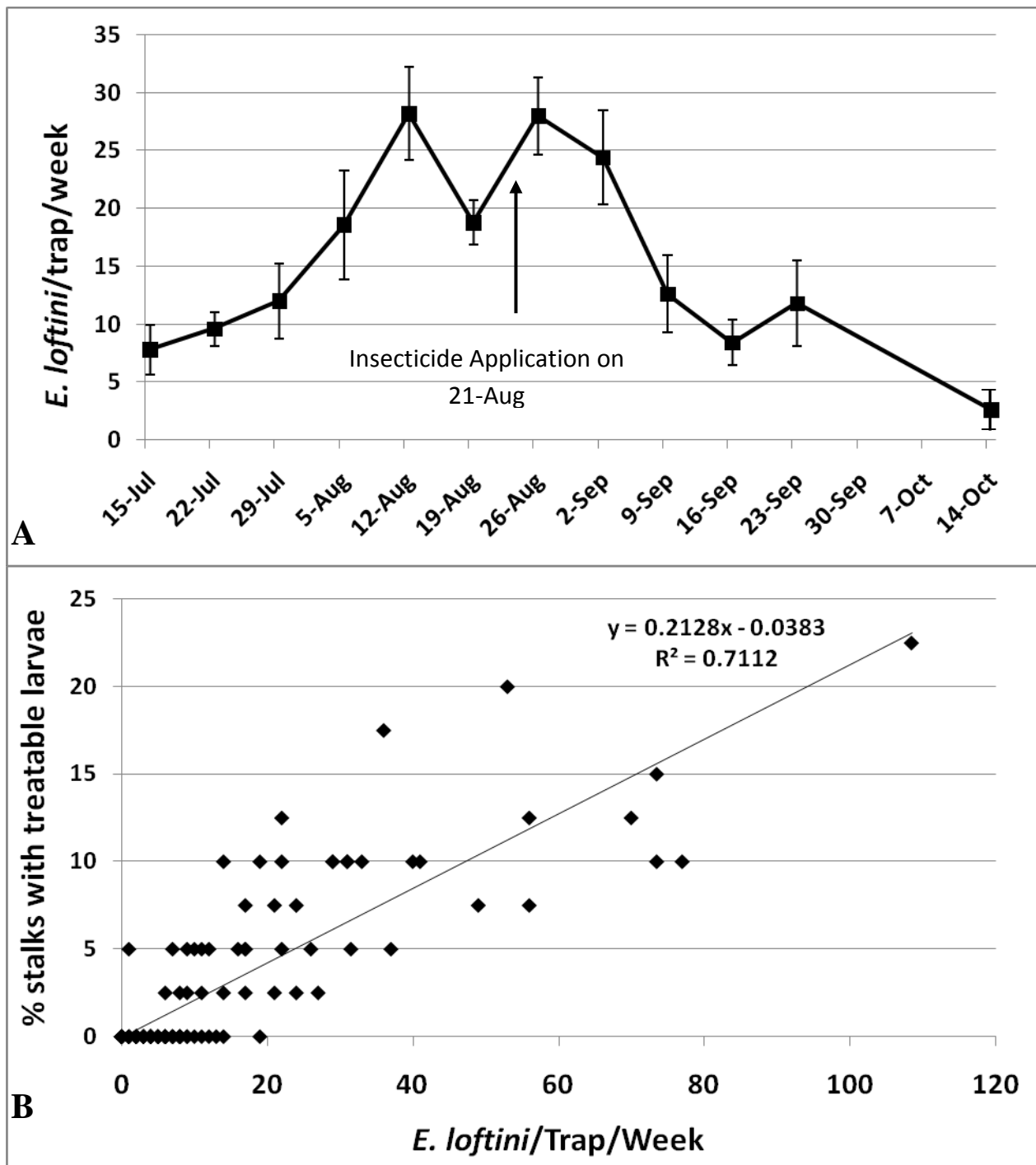


Fig. 2.1: Pheromone Trap Catch Results LRGV 2009 and 2010. (A) Average no. of male *E. loftini*/trap/wk through out the growing season. (B) The relationship between adult population densities (no. of male *E. loftini*/trap/wk) and larval infestation (percent of stalks infested with treatable larvae feeding in leaf sheaths), 2010.

Due to a late season crop killing freeze in the winter of 2009-2010, yield was not collected in 2 replications and 2009 sugar yield and quality data were not included in the analyses. Data from 2010 indicate that insecticide treatments improved juice purity, percentage sucrose, brix, and sugar/metric ton cane, metric tons of cane per acre, and recoverable sugar (tons of sugar/acre) (Table 2.1). A single application of novaluron increased sugar yield by 14% (7.29 metric tons sugar/ha) over untreated controls (6.39). β -cyfluthrin treated plots were only significantly different from untreated controls in terms of sugar yield per metric ton of cane.

Table 2.1: Sugar Yield and Quality as Affected by Insecticide Treatments
Cameron and Hildalgo Counties, TX. 2010

	Purity	POL (% Sucrose)	Brix	Sugar (Kg/ tonne of cane)	Cane (tonnes/ha)	Sugar (tonnes/ha)
Novaluron	85.3 (± 0.4)A	14.5 (± 0.17)A	17.0 (± 0.2)A	104.07 (± 1.85)A	70.1 (± 4.2) A	7.29 (± 0.48) A
Baythroid	85.0 (± 0.4) AB	14.2 (± 0.18)B	16.7 (± 0.2)B	101.47 (± 1.85)B	58.7 (± 4.2) B	5.97 ($\pm .48$) B
Control	84.4 (± 0.4)B	14.0 (± 0.17)B	16.5 (± 0.2)B	98.87 (± 1.85)C	64.4 (± 4.2) AB	6.39 ($\pm .48$) B
F	4.15 ^a	13.94 ^a	7.47 ^a	16.03 ^a	5.60 ^b	6.78 ^b
P > F	0.018	<0.0001	0.0009	<0.0001	0.03	0.019

*Means which share the same letter are not significantly different

^a df= 2, 124; Means were separated with Tukey's HSD ($\alpha=0.05$)

^b df= 2,8 Means were separated with Fisher's LSD test

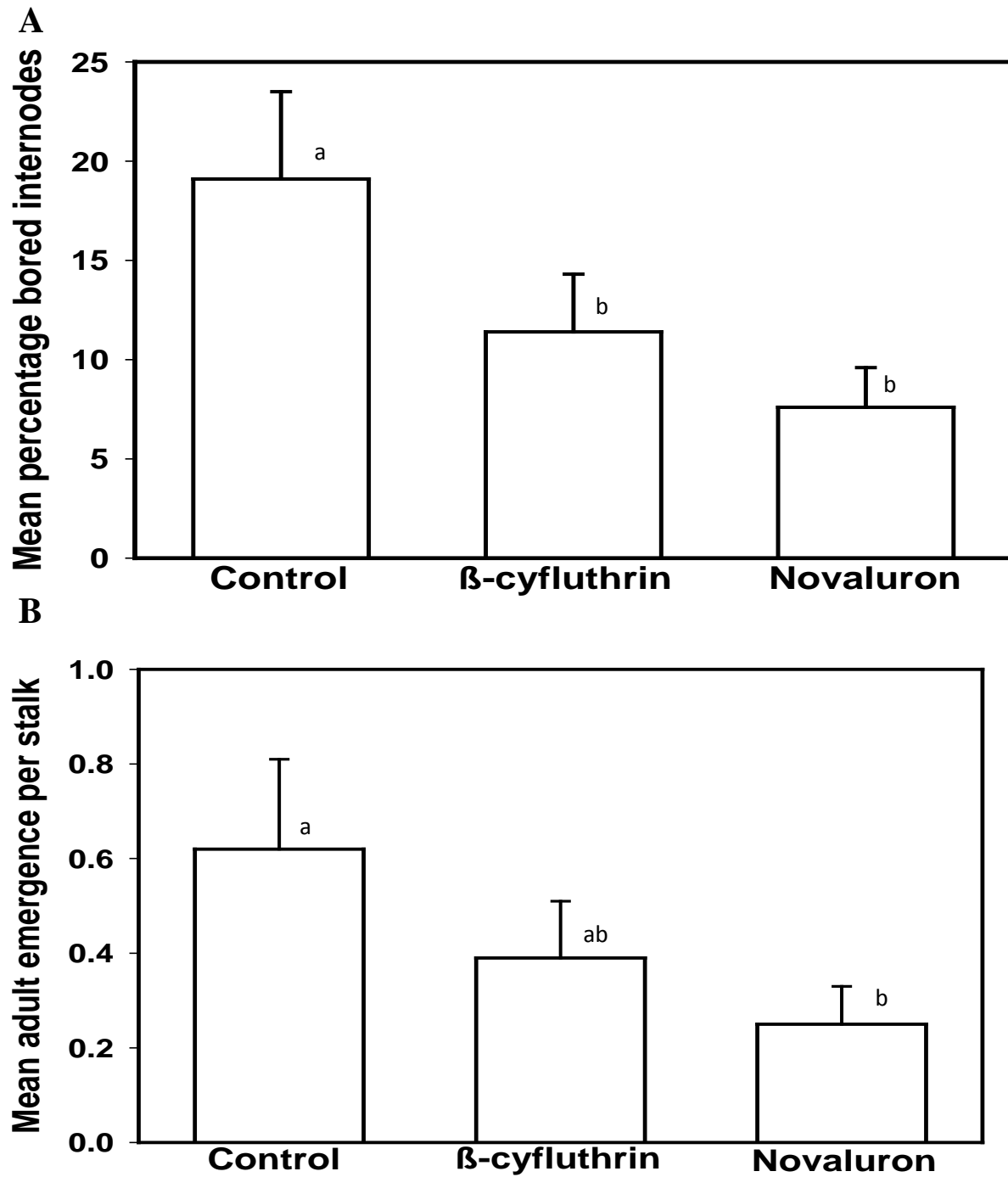


Fig. 2.2: *E. loftini* injury and emergence sugarcane aerial insecticide application experiment in Cameron and Hidalgo Counties, TX. (A) LS mean (\pm SE) probability of an *E. loftini* bored internode (equivalent to proportion of bored internodes) and (B) LS mean (\pm SE) number of moth emergence holes per stalk. Bars within each chart followed by the same letter are not significantly different ($P > 0.05$, Tukey's HSD).

Neonate Establishment and Behavior. Over all treatments and replications, establishment and behavior of a total of 277 larvae was monitored. On the first day after egg hatch, numerous entry holes in the mid-ribs of sugarcane leaves were observed indicating neonates had bored into leaves within one day of hatching. The percentage of larvae to enter the mid-rib within one day after eclosion ranged from 24.1–67.5%. The percentage of larvae surviving to stalk entry ranged from 27.4–72.4%, and mean duration of exposure ranged from 3.5–6.4 d. When all established larvae were considered, differences in dispersal distance were not detected between treatments; however, there was a trend for greater dispersal on HoCP 85-845 (resistant) compared to HoCP 00-950 (susceptible) and immature compared to mature cane (Table 2.2).

Over both phenological stages, the percentage of eclosed neonates (hatched eggs) which became established feeding on the stalk was 40% greater on susceptible cultivar HoCP 00-950 than on resistant HoCP 85-845 (Table 2.2). The proportion of established larvae which bored into the leaf mid-rib within one day was twice as high for HoCP 00-950 than HoCP 85-845 (Table 2.2). Average dispersal distance (no. of leaves/internodes from oviposition sites) was 19% greater on HoCP 85-845 than HoCP 00-950, but differences were not statistically significant. The duration of exposure of all established larvae was 40% longer on HoCP 85-845 than HoCP 00-950. When neonates that entered the mid-rib were excluded the mean duration of exposure over all treatments rose 34% and was 14.7% longer in HoCP 85-845 compared to HoCP 00-950 (Table 2.2).

Differences were detected between immature (6 nodes) and mature (12 nodes) sugarcane in the percentage of eclosed larvae which became established feeding on stalks, the proportion of established larvae to successfully enter the stalk, and duration of exposure. The percentage of eclosed larvae that established feeding was 60% greater on mature than immature stalks. The

percentage of established larvae surviving to stalk entry was 90% greater on immature than mature stalks. Average dispersal distance was greater on immature (1.458 internodes from oviposition sites) than on mature sugarcane (1.153). All dispersal on immature cane was towards the top of the stalk while 21% of larvae moved down from oviposition sites on mature sugarcane. Duration of exposure was 20% greater on mature stalks than immature. Mean duration of exposure of all established larvae was 4.67 and 5.90 d compared to 6.37 and 7.79 d for sheath feeding larvae on immature and mature sugarcane, respectively. A significant interaction effect was detected between cultivar and phenological stage for the percentage of larvae entering the mid-rib within one day and the percentage of established larvae surviving to stalk entry. Larval vulnerability was least on immature HoCP 00-950 which had the greatest percentage of larvae entering the mid-rib within 1 d (67.5%) and a mean duration of exposure of only 3.5 d. Larval exposure was maximized on mature HoCP 85-845 having a mean exposure duration of 6.4 d.

Discussion

The use of pheromone traps to assist scouting for stalkborers in sugarcane demonstrated potential to reduce scouting effort and improve chemical control of *E. loftini*. Scouting for stalkborers in sugarcane is both time consuming and labor intensive because of the high number of stalks that must be examined in order to accurately determine larval infestations (Meagher *et al.* 1996). The use of an action threshold based on *E. loftini* pheromone trap catches could enhance scouting efficiency by focusing scouting efforts at more appropriate times when adult population densities are high. When directed by a moth threshold of 20-25 *E. loftini*/trap/week, only one incident of larval scouting was necessary to indicate the need for treatment in 2009.

Table 2.2: *E. loftini* neonate establishment and behavior on two phenological growth stages of sugarcane cultivars HoCP 84-845 and HoCP 00-950, Weslaco, TX, 2010

	Percent of eclosed larvae established feeding	Percent of established larvae which entered midrib within 1 day	Percent of established larvae surviving to stalk entry	Mean dispersal distance (nodes from oviposition site)	Mean duration of exposure (days) all larvae	Mean duration of exposure (days) sheath feeding larvae
Growth Stage						
Immature	16.02	44.84	64.17	1.46	4.676	6.367
Mature	26.15	37.58	33.89	1.15	5.895	7.786
<i>F</i>	15.43 ^a	0.91 ^a	16.77 ^a	1.81 ^b	4.23 ^c	21.03 ^d
<i>P > F</i>	0.0003	0.3447	0.0002	0.1815	0.0417	<0.0001
Cultivar						
HoCP 85-845	17.63	28.30	49.88	1.419	6.181	7.559
HoCP 00-950	23.99	55.36	47.98	1.193	4.391	6.593
<i>F</i>	5.08 ^a	13.27 ^a	0.06 ^a	0.99 ^b	9.13 ^c	9.73 ^d
<i>P > F</i>	0.0176	0.0007	.8047	0.3214	0.0038	0.0025
Growth Stage X Cultivar						
Immature						
HoCP 85-845	14.08	24.14	72.41	1.756	5.952	7.118
HoCP 00-950	18.18	67.50	55.00	1.161	3.400	5.615
Mature						
HoCP 85-845	21.86	32.88	27.40	1.082	6.409	8.000
HoCP 00-950	30.95	42.54	41.04	1.225	5.381	7.571
<i>F</i>	0.28 ^a	5.42 ^a	5.08 ^a	2.65 ^b	1.65 ^c	3.01 ^d
<i>P > F</i>	0.5990	0.0246	0.0293	0.1073	0.2006	0.0864

^a df = 1,44

^b df= 1, 159; considers all larvae

^c df = 1,127; considers all larvae which survived until stalk entry

^d df = 1,85; considers sheath feeding larvae which survived until stalk entry

Weekly larval scouting from June to mid-August in 2010 further revealed a strong positive relationship between the number of *E. loftini*/trap/wk and the percentage of stalks infested with treatable larvae on plant surfaces. Linear regression analysis indicated that a trap catch of 23.6 *E. loftini*/trap/wk corresponds to the treatment threshold of 5% of stalks infested with treatable larvae. This suggests that an action threshold of 20-25 *E. loftini*/trap/week is appropriate to initiate scouting and verify larval infestations. Therefore, pheromone trap assisted scouting could potentially be further evaluated by consultants and utilized on a commercial scale to increase efficiency of consultant monitoring. The Louisiana sugarcane industry is heavily dependent on consultant scouting for the sugarcane borer and the infrastructure is in place to employ pheromone trap assisted scouting when *E. loftini* becomes established as a major economic pest in Louisiana. Furthermore, this approach also seems feasible to assist LRGV growers in their efforts to achieve more efficient *E. loftini* control. Growers should be aware that the scouting threshold may not always be representative of larval infestations due to variation in pheromone trap catches.

When appropriately timed, a single insecticide application reduced *E. loftini* injury and adult emergence in both 2009 and 2010. Control tactics which reduce adult emergence could aid in reducing area-wide populations and slow the expansion of this invasive pest. The superior control of novaluron is likely the result of both longer residual activity and reduced mortality of beneficial insects (Beuzelin *et al.* 2010). Novaluron has been shown to have good residual activity, remaining effective for 10-30 d depending on environmental conditions (Ishauya *et al.* 2002). β -cyfluthrin has demonstrated longer residual activity relative to other pyrethroids (Athanassiou *et al.* 2003), but its residual toxicity is negatively correlated with temperature and is greatly reduced at temperatures exceeding 25°C (Arthur 1999) with less than 50% mortality

one week after application. The negative relationship between pyrethroid residual activity and temperature (Toth and Sparks 1990) may be an important factor limiting pyrethroid efficacy in the LRGV where average summer temperatures are ~ 35°C (National Weather Service).

Novaluron and other insect growth regulators have been shown to be less toxic to non-target arthropods than pyrethroid insecticides and better preserve natural pest suppression (Reagan and Posey 2001, Beuzelin *et al.* 2010). Novaluron, a benzoylphenyl urea insect growth regulator, is selective for larval stages because it inhibits chitin synthesis. Reduced predation in β -cyfluthrin treated plots likely also contributed to the superior control provided by novaluron.

Previous studies have shown that chemical control of *E. loftini* is inadequate to improve sugarcane yield even when multiple insecticide applications are made (Meagher *et al.* 1996, Reay-Jones *et al.* 2005). Because of the high input cost required with indefinite return, chemical control of *E. loftini* has often not been economical, and the use of insecticides against this pest has largely been abandoned in the LRGV (Meagher *et al.* 1994). However, our studies indicate that much of the difficulty may have been due to less efficient timing of the insecticidal control approach. The economics of *E. loftini* management could be greatly improved by reduction of input costs if effective control can be achieved with a single insecticide application. The large treatment plots (3.94 ha/plot) used in this study provided an accurate assessment of insecticide application effects on sugar yield and quality in a commercial setting. The two years of bored internode (insect damage) data helps to further substantiate the yield effects. The unavailability of 2009 yield and quality data from milling because of the hard freezes in December 2009 and Jan 2010 was due to the major rush for harvesting to prevent further deterioration of sugarcane yields across the LRGV. Insecticide treatments significantly reduced injury in 2009 and 2010 based on percent bored internodes.

Key yield and quality parameters such as sugar per acre, juice purity, and sucrose content have been documented (Legaspi *et al.* 1999) as being inversely related to percentage of bored internodes. We feel that some of the past failures to detect significant effects of insecticide treatments on yield despite a reduction in percentages of bored internodes (Johnson 1985, Meagher *et al.* 1994, Legaspi *et al.* 1999b, and Reay-Jones *et al.* 2005) may be the result of high variability of sugarcane yield studies, particularly as related to small plot studies. This study is the first to adequately replicate such large acreage (>20ha/treatment). A single application of novaluron enhanced sugar yield compared to untreated controls in 2010. Novaluron treated plots had substantially higher juice purity, percent sucrose, brix, sugar/ton cane, tons cane/ha, tons of sugar/ha compared to untreated plots. The 14% increase in tons of sugar /ha is consistent with the reduction in percent bored internodes as predicted by Legaspi *et al.* (1999a). Based on the current price of raw sugar of \$706.80/ton, the novaluron treatment would be expected to increase revenue by \$707.39 per ha. This provides the first clear evidence of insecticidal control of *E. loftini* resulting in a measurable increase in subsequent sugar yield and quality. Precisely timed applications of improved insecticide chemistries have potential to improve the economics of chemical control of *E. loftini*. In addition, the scouting methods and economic thresholds used to direct insecticide applications in this study are also expected to be useful in development of improved management practices of *E. loftini* as well as other stalk boring pests.

The importance of application timing and development of management tactics which target neonates is further supported by the greenhouse study which suggests the duration of larval feeding in the leaf sheaths is shorter than previously estimated (Van Leerdam 1986, Ring *et al.* 1999). Because larvae become protected once they bore into the stalk, the period of exposure while feeding on leaves and sheaths is the only time larvae are vulnerable to insecticides and

biological control. In our study, the duration of larval exposure over all cultivars and phenological stages ranged from 3.5-6.4 d. In addition, results indicate as many as 67.5% (immature HoCP 00-950) of *E. loftini* neonates bore into the mid-rib within one day where they become protected from control agents such as insecticides and natural enemies. Prior to this research, larval entry into the mid-rib of sugarcane leaves and potential applications to *E. loftini* IPM had not been documented.

Differences in larval behavior between cultivars suggest resistant varieties which impede larval establishment have potential to improve efficacy of other control tactics. Total larval establishment (percent of eclosed larvae feeding on the plant) was greater on susceptible HoCP 00-950 than on resistant HoCP 85-845 which we believe is due to the greater percentage of larvae which borer into the mid-rib becoming protected within one day in HoCP 00-950. Mean duration of larval exposure was nearly 2 d longer on HoCP 85-845 compared to HoCP 00-950. When all established larvae were considered, mean dispersal was greater on the HoCP 85-845 than on HoCP 00-950. The increased duration of exposure on HoCP 84-845 could be due to the greater dispersal distance on the resistant cultivar as larvae search for more suitable feeding sites with higher nutrient quality.

When both cultivars were considered, a greater percentage of eclosed larvae became established on mature sugarcane than immature. This may be related to increased expression of phenolic compounds in immature sugarcane (Atkinson and Nuss 1989). Another possible explanation is the increased space available for larval establishment on larger mature sugarcane as crowded conditions may have limited establishment on young cane. However, once established, larval survival to stalk entry was nearly twice as great on immature than mature cane indicating young internodes are more susceptible to borer entry. Although more larvae became

established feeding on the leaves and sheaths of immature cane, proportionately fewer successfully entered the stalk. In addition, mean duration of exposure (time to stalk entry) was greater on mature than immature cane. This suggests that physiological factors such as increased rind hardness of mature cane impede larval entry into the stalk. This is consistent with previous research (Van Leerdam 1986, Ring *et al.* 1990) which indicates that young internodes are more vulnerable to *E. loftini* injury.

Host plant characteristics which are unfavorable for larval establishment are a key component of host plant resistance to stalkborers (Mathes and Charpentier 1969). Resistance mechanisms which prolong larval exposure outside the stalk may enhance the efficacy of other control tactics such as insecticide applications or biological control. Improved efficacy of chemical control of stalkborers has been documented when insecticide applications are used in conjunction with host plant resistance (Posey *et al.* 2006).

Rapid neonate entry into the mid-rib and the short duration of sheath feeding strongly suggest that *E. loftini* larvae are protected from foliar applied contact insecticides. Thus, residual activity, ovicidal activity and sublethal effects of insecticides will likely contribute to improved chemical control. The residual activity of novaluron may be in part responsible for the superior control observed in field studies. Chemical control might also be improved by enhancing this residual activity through the use of surfactants. Other new insecticides which have demonstrated substantial residual activity should also be evaluated for *E. loftini* management in sugarcane. Novaluron has been shown to have significant ovicidal contact toxicity against the European corn borer, another Crambid pest (Boiteau and Noronha 2007). Sub-lethal effects of novaluron also include reduced egg viability following adult exposure (Lopez *et al.* 2008) and increased occurrence of morphological abnormalities such as malformations of the wing in emerging adults

(Cetin *et al.* 2006). Insecticides with translaminar properties may also better control larvae protected in leaf mid-ribs. Continued evaluation of new chemistries and application methods for control of *E. loftini* are necessary to improve chemical control of this devastating pest. In addition, future assessment of varietal resistance should emphasize identification of mechanisms which impede larval entry into the stalk. Resistant sugarcane varieties which prolong larval vulnerability would greatly improve the success of *E. loftini* IPM programs.

As *E. loftini* threatens to become established as an economic pest in Louisiana where it is expected to cause substantial revenue losses (Reay-Jones *et al.* 2008), the need for effective management strategies is becoming more important. Because of the limited vulnerability of *E. loftini* larvae which rapidly become protected within sugarcane leaves and stalks, continued examination of control tactics which target exposed neonates will contribute to development of improved *E. loftini* IPM programs. Potential control strategies highlighted by this research include the use of pheromone traps to assist scouting and substantially improved application timing, increased residual activity of insecticides, and resistant cultivars which impede larval entry into the stalk.

CHAPTER 3

FIELD EVALUATION OF 25 COMMERCIAL AND EXPERIMENTAL SUGARCANE CULTIVARS FOR *E. LOFTINI* RESISTANCE

Introduction

Host plant resistance is a vital component of sugarcane stalkborer IPM worldwide. Resistant cultivars have consistently shown reduced *E. loftini* injury when compared to susceptible cultivars (Pfannenstiel and Meagher 1991, Meagher *et al.* 1996a, Reay-Jones *et al.* 2005). The use of host plant resistance in stalkborer management can also aid in the reduction of area-wide populations (Bessin *et al.* 1991). Expansive hectarage of susceptible varieties with elevated moth production increases endemic stalkborer populations and imposes additional pressure on the remaining hectarage. Because the sugarcane borer, *D. saccharalis*, is adequately controlled through the use of insecticides, emphasis on development of stalkborer resistant sugarcane cultivars in Louisiana has declined. However, establishment of *E. loftini* in Louisiana will require multiple management tactics to reduce revenue losses (Reay-Jones *et al.* 2005). The importance of incorporating resistant varieties into *E. loftini* management programs is amplified by the insufficiency of insecticidal control. While host plant resistance has repeatedly been shown to be promising for *E. loftini* control, the mechanisms of resistance are not fully understood. Continued evaluation of commercial and experimental sugarcane cultivars is critical to the incorporation of resistant varieties into *E. loftini* IPM programs (Reay-Jones *et al.* 2003).

Unattractiveness for oviposition, impediment of larval establishment, hindrance of larval development and plant tolerance have all been cited as categories of stalkborer resistance (Mathes and Charpentier 1969, Reay-Jones *et al.* 2007b, Showler and Castro 2010a). However, the importance of each component of resistance is not well understood and is continually being evaluated. Oviposition preference may play a role in resistance because *E. loftini* is most attracted to folds in dry leaves as oviposition sites (Van Leerdam 1986, Reay-Jones *et al.* 2007b, Showler and Castro 2010b). Oviposition preference may be linked to chemoreceptors which

detect the presence or absence of primary or secondary compounds which assist females in the behavioral and physiological responses necessary for accepting or rejecting a host plant in the insect-plant interaction process (Kogan 1994). Recent research has demonstrated a positive correlation exists between free essential amino acid concentrations and eggs laid per plant (Reay-Jones *et al.* 2007b, Showler and Castro 2010a,b). *Eoreuma loftini* females have shown oviposition preference for young sugarcane (5 nodes), susceptible cultivars, and drought stressed sugarcane which all contain heightened concentrations of free essential amino acids (Reay-Jones *et al.* 2007b, Showler and Castro 2010a,b). While reduced oviposition preference contributes to resistance, other studies (Meagher *et al.* 1996a) suggest plant characteristics which affect larvae may also be important to this relationship.

Of particular importance are factors which may hinder larval establishment including physical characteristics which impede boring such as fiber content, rind hardness and leaf sheath appression. As suggested in Chapter 2, these factors might enhance the efficacy of other management tactics by prolonging larval exposure to control agents such as insecticides or beneficial insects. Some varieties currently in development by the USDA with potential to impede stalk entry are the high-fiber cultivars US 93-25, US 01-40, and Ho 06-9610. Resistance characteristics which retard larval development may be critical to reducing areawide populations. Host plant concentrations of certain primary metabolites affect larval development in addition to influencing oviposition preference. Differences in *E. loftini* larval weight and time to pupation are thought to be linked to varying levels of allelochemicals between sugarcane cultivars (Meagher *et al.* 1996a). Reduced expression of essential nutrients may lead to decreased *E. loftini* survival to adulthood and subsequent moth production (Reay-Jones *et al.* 2005, Showler and Castro 2010a).

Assessment of cultivar resistance must not only examine resistance mechanisms, but also implications of potential widespread use of resistant cultivars. Bessin *et al.* (1990) developed a sugarcane resistance rating criteria for *D. saccharalis* which assesses borer injury as well as the ability of a cultivar to enhance or reduce area-wide populations. A relative survival index for stalkborers was developed which incorporates emergence hole counts as well as percentage bored internodes to provide a season-long record of resistance (Reay-Jones *et al.* 2003). Relative survival is a key factor in assessing the effects of varieties on area-wide population densities. Variety tests are critical to the incorporation of cultivar resistance into stalkborer IPM; hence, assessment of varietal resistance to sugarcane stalkborers must be continually conducted as new high yielding varieties emerge.

One commercial cultivar which has consistently demonstrated a high level of *E. loftini* resistance is HoCP 85-845, while susceptible cultivars include L 03-371, LCP 85-384, and HoCP 05-961. Several cultivars have been previously evaluated for *E. loftini* resistance: HoCP 00-950 and HoCP 05-902 are susceptible, L 01-299, HoCP 04-838, and HoCP 96-540 have intermediate levels of *E. loftini* and *D. saccharalis* resistance (Reagan *et al.*, 2003, 2007, 2008). HoCP 96-540 demonstrated resistance against *D. saccharalis*, but was among the more susceptible cultivars when tested for *E. loftini* resistance (Reagan *et al.* 2002). L 03-371 has been shown to be resistant under conditions of heavy rainfall, but was the most susceptible under drought conditions (Reagan *et al.* 2008). Experimental varieties which may have stalkborer resistance include high-fiber varieties, clones from recurrent selection for sugarcane borer resistance, and South African cultivars (Pfannenstiel and Meagher 1991, Ring *et al.* 1991, Conlong *et al.* 2004, Posey *et al.* 2006).

Research being conducted in South Africa on host plant resistance to other sugarcane stalkborer species has revealed promising new cultivars. These cultivars, developed by the South African Sugar Research Institute in KwaZulu-Natal (N-cultivars), have potential resistance to *E. loftini* because they have demonstrated varying levels of resistance to African stalk borers, *Eldana saccharina* Walker and *Chilo sacchariphagus* Bojer (Nuss *et al.* 1991, Conlong *et al.* 2004) which share many characteristics with *E. loftini*. Drought tolerance is thought to be an important resistance mechanism to *E. saccharina* which, like the Mexican rice borer, prefers drought stressed conditions (Conlong *et al.* 2004, Showler and Castro 2010a). Of the South African cultivars, the most resistant is N-21 and the most susceptible is N-26 (Nuss *et al.* 1991, Conlong *et al.* 2004), but the levels of resistance to *E. loftini* have not been assessed.

Continued evaluation of commercial and experimental sugarcane cultivars for stalkborer resistance is critical to the area-wide pest management of *E. loftini*. In addition to reducing *E. loftini* injury on an individual field basis, area-wide pest management may help slow the spread of this invasive pest by reducing population levels across a large geographical area. The objectives of this research were to evaluate *E. loftini* resistance under natural field conditions among 25 commercial and experimental sugarcane cultivars based on plant injury as well as suppression of adult production. Cultivars in this experiment may be incorporated into sugarcane breeding programs or considered for commercial releases in Texas and Louisiana.

Materials and Methods

A field study was conducted at the Texas A&M AgriLife Research and Extension Center at Beaumont, Texas to assess cultivar resistance to sugarcane stalkborers, *E. loftini* and *D. saccharalis*, among 25 commercial and experimental sugarcane cultivars. The varieties evaluated include five in commercial use (HoCP 85-845, HoCP 96-540, HoCP 00-950, L 01-

299, and L 03-371), eleven experimental clones (HoCP 05-902, HoCP 05-961, HoCP 04-838, Ho 06-563, Ho 07-613, Ho 07-604, Ho 07-617, Ho 07-612, Ho 06-537, L 07-68, and L 07-57), three clones bred for high fiber content (Ho 06-9610, US 93-15, and US 01-40), two clones from recurrent selection for *D. saccharalis* resistance (US 08-9001 and US 08-9003), and four South African cultivars (N-17, N-21, N-24, N-27).

A randomized complete block design five replicates was used. Block replicates had one-row plots (3.66 m long, 1.60 m row spacing and 1.22 m alleys) of each of the 25 varieties planted on 21 October 2009, (Appendix A). Only four plots were used for L-07-57 and US 93-15 because stalks failed to emerge in one replication of each variety. Beds were pulled on 20 October and opened just prior to planting in a field of Morey silt loam soil. All stalks were heat-treated prior to planting. Herbicides pendimethalin (Prowl[®]) 3.3EC at 9.615L/ha and atrazine (Atrex[®]) 4L at 9.615L/ha were applied on 21 October 2009 with a 3 nozzle spray boom (110°04 nozzles with 50 mesh screens) for pre-emergence control of grasses and broadleaves, respectively. Also, Mocap was applied at 11.5kg/ha with a hand-held gandy on non-buffer rows. Fields were exposed to natural stalkborer infestations for the remainder of the growing season. On 7-9 September 2010 ten stalk samples were collected from each plot with leaf sheaths removed for assessment of borer injury. Stalks were inspected externally for borer injury (entry and emergence holes). In addition, a stalk splitter was used to open stalks for internal examination. The number of bored internodes used in analysis included all internodes with either internal or external evidence of injury. The total number of internodes, bored internodes, and moth emergence holes were recorded for each stalk and summed for each plot. Relative survival was calculated as the no. emergence holes/no. bored internodes. Data was analyzed using generalized linear mixed models (Proc GLIMMIX, SAS Institute 2006) (Appendix D). The Kenward-Roger method

(Kenward and Roger 1997) was used to compute denominator degrees of freedom for the test of fixed effects for all variables. The proportion of bored internodes and relative survival data was analyzed with a binomial distribution and least square means are reported and separated with Tukey's HSD ($\alpha=0.05$) when differences among treatments were detected. Average emergence per stalk was calculated for each plot as the total no. emergence holes divided by the number of stalks. Emergence per stalk was analyzed with a Gaussian distribution.

Results

Differences were detected in the proportion of bored internodes between cultivars ($F = 17.68$; $df= 24, 94$; $P < 0.0001$). Injury ranged from 1.0-20.3% bored internodes (Table 3.1). *Eoreuma loftini* was responsible for >99% of bored internodes with *D. saccharalis* accounting for <1% of injury. Of the commercial varieties, HoCP 85-845 and L 01-299 were the most resistant, while L 03-371 and HoCP 96-540 were the most susceptible. HoCP 85-845 was the most resistant in terms of both injury and adult emergence (Table 3.1). HoCP 96-540, which is currently the most widely planted cultivar in Louisiana (Gravois *et al.* 2009), experienced nearly 8-fold more damage than the resistant cultivars, however, adult emergence for this cultivar was only 0.08 emergence holes/stalk. The experimental cultivars, Ho 06-563 and HoCP 05-902, were the most susceptible of all cultivars tested. All of the South African cultivars showed some level of resistance with N-21 (1.0 % bored) being the most resistant of all cultivars examined. The *D. saccharalis* resistant cultivars, US 08-9001 and US 08-9003, were also among the more resistant varieties at 5.2 and 2.6% bored, respectively. High fiber cultivars had a similar range of susceptibility (1.2 -5.8% bored) with US 93-15 being the most resistant. Adult emergence data followed the same trend as percent bored internodes (Table 3.1) however, differences in emergence between cultivars were not detected at $\alpha=0.05$ ($F=1.57$, $df= 24, 94$, $P=0.065$). The

commercial cultivar HoCP 96-540 had relatively low moth production, despite being among the most susceptible based on the proportion of bored internodes. Adult emergence ranged from < 0.01 to 0.38 emergence holes/stalk. Ho 06-463, HoCP 05-902, and L 07-57 were the most susceptible in terms of moth production with >0.30 emergence holes per stalk. Analysis of relative survival data did not converge and is not reported.

Discussion

This study demonstrates the importance evaluating commercial and experimental sugarcane cultivars for stalk borer resistance. The levels of resistance reported in this study are consistent with previous findings. Since its commercial release in 1993, HoCP 85-845 has consistently demonstrated a high level of resistance to both *E. loftini* and *D. saccharalis* (Reagan *et al.* 2003, 2004, 2005, Reay-Jones *et al.* 2003). It is a relatively high fiber cultivar and has pith (W. H. White, pers. comm.). Our findings indicate that HoCP 85-845 should continue to be viewed as a standard for stalk borer resistant sugarcane cultivars. However, Reay-Jones *et al.* (2003) found that under severe *E. loftini* pressure the level of resistance in this cultivar was reduced relative to other cultivars. One of the most susceptible cultivars evaluated in this study, HoCP 96-540, is currently the most widely planted cultivar in Louisiana representing >50% of planted acreage (Gravois *et al.* 2009). Sugarcane producers often opt to grow the highest yielding varieties, regardless of the level of stalkborer resistance.

While HoCP 96-540 has demonstrated intermediate levels of resistance to *D. saccharalis* (Reagan *et al.* 2005), it is considerably more susceptible to *E. loftini*. This appears to be due to differences in oviposition preferences between the two pest species. Despite having high levels of injury, adult emergence was relatively low for HoCP 96-540 which indicates this cultivar may be attractive for oviposition, but is among the more resistant in terms of larval development.

Table 3.1: Mean percent bored internodes and emergence per stalk. Evaluation of varietal resistance to sugarcane stalkborers. Beaumont, Texas. 2010

Variety	% Bored Internodes	Emergence per Stalk
Ho 06-563	20.3 A	0.38
HoCP 05-902	14.4 AB	0.32
HoCP 04-838	10.9 BC	0.20
Ho 07-612	10.0 BCD	0.18
L 03-371	9.5 BCD	0.14
HoCP 96-540	7.8 BCDE	0.08
L 07-57	7.1 CDEF	0.32
Ho 07-604	6.3 CDEF	0.04
US 01-40	5.8 CDEFG	0.06
N-27	5.7 CDEFG	0.12
Ho 06-537	5.7 CDEFG	0.19
Ho 07-613	5.4 CDEFG	0.02
N-17	5.4 DEFG	0.08
HoCP 05-961	5.2 DEFG	0.12
US 08-9001	5.2 DEFG	0.04
Ho 06-9610	4.9 DEFG	0.04
HoCP 00-950	4.5 DEFGH	0.04
L 07-68	4.0 EFGH	0.12
Ho 07-617	3.9 EFGH	0.06
US 08-9003	2.6 FGH	0.06
N-24	2.4 FGH	<0.01
L 01-299	2.2 FGH	0.04
US 93-15	1.2 GH	0.01
HoCP 85-845	1.0 H	<0.01
N-21	1.0 H	<0.01

*Means which share a letter are not significantly different (Tukey's HSD, $\alpha=0.05$).

LS means: $F=17.68$; $df=24, 94$; $P < 0.0001$

SE = 8.52 for all cultivars except L 07-57 and US93-15 (SE=9.51)

Although not currently planted on a large portion of sugarcane acreage in Louisiana, L 01-299 may offer a high yielding, stalkborer resistant variety which could be incorporated into *E. loftini* IPM programs.

Additionally, this research evaluated *E. loftini* resistance among several experimental cultivars in various levels of the sugarcane breeding programs. The two varieties which have been identified as having antibiosis to *D. saccharalis*, Ho 08-9001 and Ho 08-9003 (W. H. White, pers. comm.), demonstrated moderate to high levels of resistance despite *E. loftini* accounting for the vast majority of injury. These two cultivars are currently in the process of being registered with Crop Science as resistant germplasms and may be incorporated into future breeding programs. The high fiber cultivars, US 93-15, US 01-40 and Ho 06-9610, had varying levels of susceptibility indicating high fiber content alone may not be adequate to provide resistance to *E. loftini*. However, the high fiber cultivar US 93-15 was among the most resistant of the varieties evaluated. Fiber content is often negatively associated with cane yield (Gravois *et al.* 2009) and widespread commercial planting of high fiber cultivars may not be economical for Louisiana sugarcane growers. However, if high fiber cultivars demonstrate potential to impede larval entry into the stalk, they may be used to enhance efficacy of other control tactics, especially for energy canes (those developed for biomass). Of the experimental cultivars, only HoCP 05-961, HoCP 04-838, L 07-57 and Ho 07-613 remain in the Louisiana variety program. HoCP 05-961 demonstrated moderate levels of resistance while HoCP 04-838, L 07-57, and Ho 07-613 were among the more susceptible varieties. HoCP 04-838 was previously thought to be resistant, but our results indicate it is highly susceptible to *E. loftini*. The varieties developed by the South African Sugar Research Institute all demonstrated some level of stalkborer resistance. N-21 was as resistant as HoCP 85-845 both in terms of injury and adult emergence.

Identification of these varieties as resistant may lead to their incorporation into the Louisiana breeding programs as resistant germplasms. N-21 shows resistance to a broad range of sugarcane stalkborers and could potentially be used in both *E. loftini* and *D. saccharalis* variety development programs.

Continued assessment of varietal resistance to sugarcane stalkborers is critical to the development of effective IPM programs. Research examining the mechanisms behind resistance, particularly the role of free amino acids, could lead to the development of a non biological assay for assessing stalkborer resistance. In addition, identification of resistant cultivars which prolong larval exposure by impeding stalk entry may lead to development of improved *E. loftini* IPM. Evaluation of cultivar resistance in conjunction with insecticide applications is necessary to assess the effects of resistant cultivars on efficacy of chemical control. Cultivar resistance has the potential to keep low to moderate *E. loftini* infestations suppressed below economic injury levels as well as to reduce area-wide populations. The use of resistant cultivars on a commercial basis could slow the spread of *E. loftini*. Due to the severe pest history of *E. loftini* in sugarcane, stakeholders in Louisiana cannot afford to wait until this insect becomes an economic pest before developing resistant cultivars. Continued assessment of resistance, improved understanding of resistance mechanisms and increased emphasis on stalkborer resistance by sugarcane breeding institutions are critical to *E. loftini* IPM.

CHAPTER 4

SUMMARY AND CONCLUSIONS

Since its establishment in the Lower Rio Grande Valley (LRGV) in 1980, the Mexican rice borer, *Eoreuma loftini* (Dyar), has been the dominant pest of sugarcane in that area, where it is responsible for major revenue losses (Johnson and Van Leerdam 1981, Legaspi *et al.* 1999a). The species has since expanded its range across the Texas rice production area, reaching Louisiana in 2008 (Reay-Jones *et al.* 2007a, Hummel *et al.* 2010). Development of effective control tactics is critical to Louisiana agriculture as the invasive species is predicted to cause as much as \$220 million and \$48 million in annual revenue losses for sugarcane and rice, respectively, by 2035 (Reay-Jones *et al.* 2008).

Management of *E. loftini* in sugarcane is based on a balance of control tactics including irrigation and host plant resistance supplemented by insecticide applications (Reay-Jones *et al.* 2005). These efforts are often not able to reduce infestations below economic injury levels. Chemical control of *E. loftini* is limited by the sheltered nature of larvae which restricts their exposure to insecticides. Chemical and biological control agents target neonates which have not yet become protected within the stalk. This research investigated potential management strategies which may improve *E. loftini* management by focusing control tactics on vulnerable larvae.

A two-year aerial insecticide application study using commercial sugarcane fields conducted in the LRGV revealed chemical control strategies which both reduced *E. loftini* injury and improved subsequent sugar yield. This study highlighted the potential use of pheromone traps to increase scouting efficiency and enhance timing of insecticide applications. Regression analysis indicated that an action threshold of 20-25 male *E. loftini*/trap/wk is appropriate to initiate scouting for larval infestations. As with many similar pheromone trap studies, the attractive properties of *E. loftini* pheromone lures, dispersion distances and other behavioral relationships

are not well understood and need further examination. Over both years the insect growth regulator novaluron provided superior control compared to β -cyfluthrin and untreated plots. When applied as advised by thresholds, a single application of novaluron reduced *E. loftini* injury (proportion of bored internodes) by 60%, adult emergence by 58% and led to a 14% increase in sugar yield. The superior control of novaluron compared to pyrethroids is likely due to longer residual activity; by remaining on plant surfaces, novaluron can control neonates as soon as they eclose. Novaluron has been shown to have good residual activity remaining effective for 10–30 days depending on environmental conditions (Ishauya *et al.* 2002). β -cyfluthrin has demonstrated longer residual activity relative to other pyrethroids (Athanassiou *et al.* 2003), but its residual toxicity is negatively correlated with temperature and is greatly reduced at temperatures exceeding 25°C (Arthur 1999) with < 50% mortality one week after application. This study highlights management strategies utilizing pheromone traps to direct a single insecticide application and the improvement of chemical control of *E. loftini* in sugarcane. With the global price of sugar rising, yield reductions due to insect pests can lead to substantial decreases in revenue. The efficient chemical control demonstrated in this study has potential to greatly reduce revenue losses from *E. loftini* infestations in the LRGV.

A greenhouse study was conducted at the USDA-ARS Kika de la Garza Subtropical Agricultural Research Center in Weslaco, Texas on *E. loftini* neonate establishment and feeding behavior on two phenological stages of resistant (HoCP 85-845) and susceptible (HoCP 00-950) sugarcane cultivars. A substantial portion ($42 \pm 6.3\%$) of neonates mine into the mid-rib of sugarcane leaves within one day of eclosion where they become protected from insecticides, predators and parasitoids. The mean duration of exposure (time to stalk entry) was longer on the resistant cultivar HoCP 85-845 compared to HoCP 00-950 and on mature compared to immature

plants. This research shows the limited exposure time of *E. loftini* larvae to foliar-applied insecticides, and indicates that residual activity of insecticides may improve the efficacy of chemical control. In addition, results from this study suggest that resistant cultivars which impede stalk entry and prolong larval exposure on plant surfaces may be able to enhance the effects of other control tactics. Because neonate larvae are the target of *E. loftini* control strategies, additional research on neonate establishment could lead to the development of advanced management tactics. Continued research in this area should investigate neonate behavior on a diverse range of sugarcane cultivars and examine the relationships between larval establishment and physical characteristics of host plants. Host plant resistance will likely become more important to *E. loftini* management as it has potential to both mitigate revenue losses and reduce areawide populations.

A small plot field study was conducted at the Texas A&M AgriLife Research and Extension Center at Beaumont, Texas, which assessed relative stalkborer resistance among 25 commercial and experimental sugarcane cultivars. Although both *E. loftini* and *D. saccharalis* are present in Beaumont, *E. loftini* accounted for 99% of the infestations in this study. Differences in *E. loftini* injury were detected between cultivars. Consistent with previous findings (Reay-Jones *et al.* 2003, 2005, Reagan *et al.* 2005) the commercial cultivar HoCP 85-845 was the least injured variety examined. South African cultivars N-21 and N-24 and the high fiber cultivar US 93-15 also demonstrated high levels of resistance. The most widely planted cultivar in Louisiana, HoCP 96-540, was among the most susceptible varieties evaluated. Sugarcane breeding institutions should place more emphasis on selecting for stalkborer resistant cultivars. While *D. saccharalis* is largely controlled through the use of insecticides, the sheltered nature of *E. loftini* larvae will likely require a balance of multiple management tactics to achieve adequate control

(Reay-Jones *et al.* 2005). Host plant resistance will provide a critical element of *E. loftini* IPM programs because it may be used in conjunction with other management strategies and has the potential to slow the spread of this invasive species by reducing areawide populations. However, future research is necessary to better understand the mechanisms of *E. loftini* resistance in sugarcane. Investigation of nutritional factors which influence both *E. loftini* oviposition and larval behavior may lead to the development of a non-biological assay to evaluate levels of resistance. Examination of host plant characteristics which impede larval entry into the stalk can help identify cultivars with potential to enhance efficacy of *E. loftini* chemical control.

This research indicates that *E. loftini* management could be improved by development of control strategies which target the exposed neonate stage. The use of pheromone traps to improve timing of insecticide applications and the recently labeled novaluron both demonstrated potential to improve *E. loftini* chemical control. However, due to the extremely limited exposure time of *E. loftini* larvae, resistant cultivars should also be incorporated into management programs. Future *E. loftini* IPM programs should use a balance of irrigation (Reay-Jones *et al.* 2005, Showler and Castro 2010a), improved chemical control strategies, and host plant resistance.

REFERENCES

- Akbar, W., J. M. Beuzelin, T. E. Reagan, and B. J. Viator. 2008a.** Evaluation of insect growth regulators and a pyrethroid for sugarcane borer control through aerial application, 2007. *Arthrop. Manag. Tests* 33: F1
- Akbar, W., J. A. Ottea, J. M. Beuzelin, T. E. Reagan, and F. Huang. 2008b.** Selection and life history traits of tebufenozide-resistant sugarcane borer (Lepidoptera: Crambidae). *J. Econ. Entomol.* 101: 1903-1910.
- Akbar, W., J. M. Beuzelin, and T. E. Reagan. 2009.** Chemical control of the Mexican rice borer in the Lower Rio Grande Valley of Texas, 2008. *Arthrop. Manag. Tests* 34: F70.
- Arthur, F. 1999.** Effect of temperature on residual activity of Cyfluthrin wettable powder. *J. Econ. Entomol.* 93: 695-699.
- Athanassiou, C. G., N. G. Kavallieratos, B. J. Vayias, C. B. Dimizas, A. S. Papagregoriou, and C. T. Buchelos. 2004.** Residual toxicity of beta-cyfluthrin, alpha-cypermethrin, and deltamethrin against *Tribolium confusum* Jacquelin du Val (Coleoptera: Tenebrionidae) on stored wheat. *Appl. Entomol. Zool.* 39: 195-202
- Atkinson, P. R. and K. J. Nuss. 1989.** Associations between host-plant nitrogen and infestations of the sugarcane borer, *Eldana saccharina* Walker (Lepidoptera: Pyralidae). *Bull. Entomol. Research* 79: 489-506.
- Bessin, R. T., T. E. Reagan, and F. A. Martin. 1990.** A moth production index for evaluating sugarcane resistance to the sugarcane borer (Lepidoptera: Pyralidae). *J. Econ. Entomol.* 83: 221-225.
- Bessin, R. T., R. E. Stinner, and T. E. Reagan. 1991.** Modeling the areawide impact of sugarcane varieties and predation on sugarcane borer (Lepidoptera: Pyralidae) populations in Southern Louisiana. *Environ. Entomol.* 20: 252-257.
- Beuzelin, J., G. Coburn, W. Akbar, F.P.F. Reay-Jones, C. McAllister, and T. E. Reagan. 2005.** Aerial insecticidal control of the sugarcane borer and non-target arthropod assessment, 2004. *Arthrop. Manag. Tests* 30: F79.
- Beuzelin, J. M., W. Akbar, A. Meszaros, F.P.F. Reay-Jones, and T. E. Reagan. 2010a.** Field assessment of novaluron for sugarcane borer, *Diatraea saccharalis* (F.) (Lepidoptera: Crambidae), management in Louisiana sugarcane. *Crop Protection* 29: 1168-1176.
- Beuzelin, J. M., T. E. Reagan, M. O. Way, A. Meszaros, W. Akbar, and L.T. Wilson. 2010b.** Potential impact of Mexican ice bore non-crop hosts on sugarcane IPM. pp. 806-814 (Paper BE10). *In Proceedings, XXVII Congress of the International Society of Sugar Cane Technologists* March 7-11, Veracruz, Mexico.

- Boiteau, G. and C. Noronha. 2007.** Topical, residual, and ovicidal activity of three reduced risk insecticides against the European corn borer, *Ostrinia nubilalis* (Lepidoptera: Crambidae) on potato. *Pest Manag. Sci.* 36: 1230-1238.
- Brown, H. E., L. T. Wood, T. N. Shaver, and J. Worley. 1988.** Behavioral responses of male *Eoreuma loftini* (Lepidoptera: Pyralidae) to ovipositor extracts. *J. Econ. Entomol.* 81: 184-188.
- Browning, H. W., and C. W. Melton. 1987.** Indigenous and exotic Trichogrammatids (Hymenoptera: Trichogrammatidae) evaluated for biological control of *Eoreuma loftini* and *Diatraea saccharalis* (Lepidoptera: Pyralidae) borers on sugarcane. *Environ. Entomol.* 16: 360-364.
- Cetin, H., F. Erler, and A. Yanikoglu. 2006.** Larvicidal activity of novaluron, a chitin synthesis inhibitor, against the house fly, *Musca domestica*. *J. Insect Sci.* 6: 50
- Coburn, G. E. and S. D. Hensley. 1972.** Differential survival of *Diatraea saccharalis* larvae on two varieties of sugarcane. *Proc. Int. Soc. Sugarcane Technol.* 14: 440-444..
- Conlong, D. E., P. Sweet, and J. Piwalo. 2004.** Resistance of Southern African varieties of sugarcane to *Chilo Sacchariphagus* (Lepidoptera: Crambidae) in Mozambique, and development of a non-destructive field resistance rating system. *Proc. S. African Sugar Tech. Assn.* 78: 297-306.
- Dyar, H. G. 1917.** Seven new crambids from the United States. *Insecutor Inscitiae Mentrus* 5: 84-87.
- Hall, D. G. 1986.** Sampling for the sugarcane borer (Lepidoptera: Pyralidae) in sugarcane. *J. Econ. Entomol.* 79: 813-816
- Hensley, S.D. 1971.** Management of sugarcane borer populations in Louisiana, a decade of change. *Entomophaga* 16: 133-146.
- Hensley, S.D. and W.H. Long. 1969.** Differential yield responses of commercial sugarcane varieties to sugarcane borer damage. *J. Econ. Entomol.* 62: 620-623.
- Hummel, N. A., T. Hardy, T. E. Reagan, D. K. Pollet, C. E. Carlton, M. J. Stout, J. M. Beuzelin, W. Akbar, and W. H. White. 2010.** Monitoring and first discovery of the Mexican rice borer *Eoreuma loftini* (Lepidoptera: Crambidae) in Louisiana. *Florida Entomol.* 93: 123-124.
- Ishaaya, I., S. Kontsedalov, and A. R. Horowitz. 2003.** Novaluron (Rimon), a novel IGR: potency and cross-resistance. *Archives of Insect Biochemistry and Physiology* 54:157-164.

- Johnson, K.J.R. 1983.** Field evaluation of insecticides for rice borer control on sugarcane in Texas, 1982-1983. Report to the Lower Rio Grande Valley Sugar Growers, Inc.
- Johnson, K.J.R. 1984.** Identification of *Eoreuma loftini* (Dyar) (Lepidoptera: Pyralidae) in Texas, 1980: Forerunner for other sugarcane boring pest immigrants from Mexico? ESA Bulletin Fall 1984: 47-53.
- Johnson, K.J.R. 1985.** Seasonal occurrence and insecticidal suppression of *Eoreuma loftini* (Lepidoptera: Pyralidae) in sugarcane. J. Econ. Entomol. 78: 960-966.
- Johnson, K.J.R. and M. B. Van Leerdam. 1981.** Range extension of *Acigona loftini* into the Lower Rio Grande Valley of Texas. Sugar y Azucar 76: 34.
- Kennedy, G. G., and A. N. Kishaba. 1976.** Bionomics of *Aphis gossypii* on resistant and susceptible cantaloupe. Environ. Entomol. 5: 357-361.
- Kogan, M. 1994.** Plant resistance in pest management (Chapter 3) pp.73-128 in Introduction to Insect Pest Management (3rd ed) by R.L. Metcalf and W.H. Luckmann. Wiley Interscience, New York.
- Kenward, M. H. and J. H. Roger. 1997.** Small sample inference for fixed effects from restricted maximum likelihood. Biometrics 53: 983-997.
- Legaspi, J. C., B. C. Legaspi Jr., E. G. King, and R. R. Saldaña. 1997.** Mexican rice borer, *Eoreuma loftini* (Lepidoptera: Pyralidae) in the Lower Rio Grande Valley of Texas: Its history and control. Tropical Plant Sci. 49: 53-64.
- Legaspi, J. C., B. C. Legaspi Jr., J. E. Irvine, J. Johnson, R. L. Meagher Jr., and N. Rozeff. 1999a.** Stalkborer damage on yield and quality of sugarcane in Lower Rio Grande Valley of Texas. J. Econ. Entomol. 92: 228-234.
- Legaspi, J.C., B.C. Legaspi Jr, and R.R. Saldaña. 1999b.** Laboratory and field evaluations of biorational insecticides against the Mexican rice borer (Lepidoptera: Pyralidae) and a parasitoid (Hymenoptera: Braconidae). J. Econ. Entomol. 92: 804-810.
- Lopez, J. D., B. K. Fritz, D. E. Martin, W. C. Hoffman, Y. Lan, and M. A. Latheef. 2008.** Laboratory evaluation of novaluron for toxicity to nymphal instars of field collected southern green stink bugs on cotton. Southwestern Entomol. 33: 119-127.
- Martin, F.A., C. A. Richard, and S. D. Hensley. 1975.** Host resistance to *Diatraea saccharalis* F.: relationship between of sugarcane internode harness to larval damage. Environ. Entomol. 4: 687-688.

- Martinez, A. J., J. Baird, and T. Holler. 1988.** Mass rearing sugarcane borer and rice borer for production of parasites *Allorhagus pyralophagus* and *Rhaconotus roslinensis*. USDA dep Agric. APHIS-PPQ. APHIS 83-1.
- Mathes, R., and L. J. Charpentier. 1969.** Varietal resistance in sugar cane to stalk moth borers, pp. 175- 188. In J. R. Williams, J. R. Metcalfe, R.W. Mungomery, and R. Mathes [eds.], Pests of Sugar cane. Elsevier, New York.
- Meagher, R. L. Jr., J. W. Smith Jr., and K.J.R. Johnson. 1994.** Insecticidal management of *Eoreuma loftini* (Lepidoptera: Pyralidae) on Texas sugarcane: A critical review. J. Econ. Entomol. 87: 1332-1344.
- Meagher, R. L. Jr., J. E. Irvine, R.G. Breene, R. S. Pfannenstiel, and M. Gallo-Meagher. 1996a.** Resistance mechanisms of sugarcane to Mexican rice borer (Lepidoptera: Pyralidae). J. Econ. Entomol. 89: 536-543.
- Meagher, R. L. Jr., L. T. Wilson, and R. S. Pfannenstiel. 1996b.** Sampling *Eoreuma loftini* (Lepidoptera: Pyralidae) on Texas sugarcane. Environ. Entomol. 25: 7-16.
- Meagher, R. L. Jr., J. W. Smith Jr., H. W. Browning, and R. R. Saldaña. 1998.** Sugarcane stemborers and their parasites in southern Texas. Environ. Entomol. 27: 759-766.
- Nuss, K. J. 1991.** Screening sugarcane varieties for resistance to the Eldana borer. Proc. S. African Sugar Tech. Assn. 1991: 92-96.
- Pfannenstiel, R. S. and R. L. Meagher, Jr. 1991.** Sugarcane resistance to stalkborers (Lepidoptera: Pyralidae) in south Texas. Florida Entomol. 74:300-305.
- Pimentel, D., L. Lach, R. Zuniga, and D. Morrison. 2000.** Environmental and economic costs of nonindigenous species in the United States. BioScience 50: 53-65.
- Poisson, S. D. 1837.** Recherches sur la Probabilité des Jugements en Matière Criminelle et en Matière Civile: Précédées des Règles Générales du Calcul des Probabilités. , Bachelier, Paris
- Posey, F. R., W. H. White, F.P.F. Reay-Jones, K. Gravois, M. E. Salassi, B. R. Leonard, and T. E. Reagan. 2006.** Sugarcane borer (Lepidoptera: Crambidae) management threshold assessment on four sugarcane cultivars. J. Econ. Entomol. 99: 966-971.
- Ramaswamy, S. B. 1988.** Host finding by moths: sensory modalities and behaviours. J. Insect Physiol. 24: 235-249.
- Reagan, T. E., W. Akbar, C. D. McAllister, and F.P.F. Reay-Jones. 2003.** Assessment of varietal resistance to the sugarcane borer. Sugarcane Research: Annual Progress Report 2003. LSU AgCenter Publication p103-104.

- Reagan, T. E., W. Akbar, C. D. McAllister, and F.P.F. Reay-Jones. 2004.** Assessment of varietal resistance to the sugarcane borer. Sugarcane Research: Annual Progress Report 2004. LSU AgCenter Publication p123-124.
- Reagan, T. E., F. R. Posey, C. Blanco, R. Miller, and J. W. McGee. 2001.** Small plot assessment of insecticides against the Mexican rice borer in sugarcane, 2000. Arthrop. Manag. Tests 26: 110F.
- Reagan, T. E., F.P.F. Reay-Jones, B. L. Legendre, M. O. Way, and J. Amador. 2005.** Slowing down the Mexican rice borer. Louisiana Agric. 48: 6-8.
- Reagan, T. E., M. O. Way, J. M. Beuzelin, and W. Akbar. 2008.** Assessment of varietal resistance to the sugarcane borer and Mexican rice borer. Sugarcane Research: Annual Progress Report 2008. LSU AgCenter Publication. P120-121.
- Reay-Jones, F.P.F., M. O. Way, M. Sétamou, B. L. Legendre, and T. E. Reagan. 2003.** Resistance to the Mexican rice borer (Lepidoptera: Crambidae) among Louisiana and Texas sugarcane cultivars. J. Econ Entomol. 96: 1929-1934.
- Reay-Jones, F.P.F, A. T. Showler, T. E. Reagan, B. L. Legendre, M. O. Way, and E. B. Moser. 2005.** Integrated tactics for managing the Mexican rice borer (Lepidoptera: Crambidae) in sugarcane. Environ. Entomol. 34: 1558-1565.
- Reay-Jones, F.P.F., L. T. Wilson, M. O. Way, T. E. Reagan, and C. E. Carlton. 2007a.** Movement of Mexican rice borer (Lepidoptera: Crambidae) through the Texas rice belt. J. Econ. Entomol. 100: 54-60.
- Reay-Jones, F.P.F., L. T. Wilson, A. T. Showler, T. E. Reagan, and M. O. Way. 2007b.** Role of oviposition preference in an invasive crambid impacting two graminaceous host crops. Environ. Entomol. 36: 938-951.
- Reay-Jones, F. P. F., L. T. Wilson, T. E. Reagan, B. L. Legendre, and M. O. Way. 2008.** Predicting economic losses from the continued spread of the Mexican rice borer (Lepidoptera: Crambidae). J. Econ. Entomol. 101: 237-250.
- Ring, D. R., H. W. Browning, K.J.R. Johnson, J. W. Smith Jr., and C. E. Gates. 1991.** Age specific susceptibility of sugarcane internodes to attack by the Mexican rice borer (Lepidoptera: Pyralidae). J. Econ. Entomol. 84: 1001-1009.
- Robacker, D. C. and P. J. Landholt. 2002.** Importance and use of attractants, pp. 169-205. In *In* G. J. Hallman and C. P. Schwalbe [eds.], Invasive arthropods in agriculture, Science Publishers, Enfield, NH.

- Rodriguez, L. M., E. A. Ostheimer, A. E. Woolwine, and T. E. Reagan. 1995.** Efficacy of aerial application of selected insecticides against the sugarcane borer, 1994. *Arthro. Manage. Tests* 20 (131F), 254.
- Rodriguez, L. M., A. E. Woolwine, E. A. Ostheimer, H. P. Schexnayder Jr., and T. E. Reagan. 1998.** Insecticidal control of the sugarcane borer-aerial application test, 1997. *Arthro. Manage. Tests* 23 (140F), 287.
- Rodriguez, L. M., J. A. Ottea, and T. E. Reagan. 2001.** Selection, egg viability, and fecundity of the sugarcane borer (Lepidoptera: Crambidae) with tebufenozide. *J. Econ. Entomol.* 94: 1553-1557.
- SAS Institute. 2006.** User's Manual, Version 9.2. SAS institute, Cary, NC
- Shaver, T. N., H. E. Brown, H. J. Williams, L. T. Wood and J. Worley. 1988.** Components of female sex pheromone of *Eoreuma loftini* Dyar. *J. Chem. Ecol.* 14: 391-399.
- Shaver, T. N., H. E. Brown, and D. E. Hendricks. 1990.** Development of pheromone lure for monitoring field populations of *Eoreuma loftini* (Lepidoptera: Pyralidae). *J. Chem. Ecol.* 16: 2393-2399.
- Shaver, T. N., H. E. Brown, J. W. Bard, T. C. Holler, and D. E. Hendricks. 1991.** Field evaluations of pheromone-baited traps for monitoring Mexican rice borer (Lepidoptera: Pyralidae). *J. Econ. Entomol.* 84: 1216-1219.
- Showler, A. T. and B. A. Castro. 2010a.** Influence of drought stress on Mexican rice borer (Lepidoptera: Crambidae) oviposition preference in sugarcane. *Crop Protection* 29:415-421.
- Showler, A. T. and B. A. Castro. 2010b.** Mexican rice borer, *Eoreuma loftini* (Lepidoptera: Crambidae), oviposition site selection stimuli on sugarcane, and potential field applications. *J. Econ. Entomol.* 103:1180-1186.
- Toth, S. J., Jr., and T. C. Sparks. 1990.** Effect of temperature on toxicity and knockdown activity of *cis*-permethrin, esfenvalerate, and λ -cyhalothrin in the cabbage looper (Lepidoptera: Noctuidae). *J. Econ. Entomol.* 83: 342-346.
- Van Leerdam, M. B., K.J.R. Johnson , and J. W. Smith, Jr. 1894.** Effects of substrate physical characteristics and orientation on oviposition by *Eoreuma loftini* (Lepidoptera: Pyralidae). *Environ. Entomol.* 13: 800-802.
- Van Leerdam, M.B. 1986.** Bionomics of *Eoreuma loftini* a Pyralid stalk borer of sugarcane. PhD Dissertation. Texas A&M University.
- White, W. H. and S. D. Hensley. 1987.** Techniques to quantify *Diatraea saccharalis* (Lepidoptera: Pyralidae) on sugarcane quality. *Field Crop Protection* 15:341-345

APENDIX A

VARIETAL RESISTANCE TEST PLOT PLAN

		US 02-9010 (3 rows)		HoCP 91-552 (2 rows)		US 07-9027 (2 rows)	
V	US 04-9076	US 08-9003	Ho 06-563	HoCP 05-961	L07-57	HoCP 05-902	US 07-9019
		HoCP 85-845	Ho 07-604	Ho 07-612	US 08-9001	Ho 06-537	
		HoCP 04-838	L 03-371	Ho 06-9610	N-24	N-27	
		L 01-299	Ho 07-613	HoCP 00-950	HoCP 96-540	N-17	
		N-21	US 01-40	L 07-68	Ho 07-617	US 93-15	
IV	US 07-9612	HoCP 05-902	Ho 07-612	US 01-40	HoCP 05-961	Ho 07-604	HoCP 04-838
		US 93-15	Ho 07-613	L 01-299	US 08-9003	US 08-9001	
		L 07-57	L 07-68	HoCP 85-845	Ho 06-563	N-24	
		HoCP 00-950	L 03-371	HoCP 04-838	N-21	Ho 07-617	
		N-17	HoCP 96-540	N-27	Ho 06-537	Ho 06-9610	
III	US 07-9015	Ho 06-563	Ho 07-612	HoCP 05-961	US 08-9003	L 07-57	US 02-113
		Ho 06-9610	HoCP 00-950	N-21	HoCP 04-838	HoCP 96-540	
		Ho 07-617	N-24	N-17	US 93-15	N-27	
		HoCP 85-845	Ho 07-613	L 03-371	HoCP 05-902	US 01-40	
		US 08-9001	L 01-299	Ho 07-604	L 07-68	Ho 06-537	
II	US 07-9014	Ho 07-617	HoCP 04-838	HoCP 85-845	N-27	L 03-371	US 07-9017
		N-17	Ho 07-613	N-21	Ho 06-9610	HoCP 00-950	
		L 01-299	US 93-15	US 01-40	HoCP 96-540	N-24	
		Ho 06-563	Ho 06-537	Ho 07-612	US 08-9001	L 07-68	
		Ho 07-604	HoCP 05-961	US 08-9003	L 07-57	HoCP 05-902	
I	CP 44-155	HoCP 05-902	US 01-40	Ho 07-612	Ho 06-537	L 03-371	L 01-299
		L 07-57	L 07-68	HoCP 00-950	HoCP 85-845	US 08-9003	
		Ho 06-563	HoCP 04-838	N-17	HoCP 05-961	US 08-9001	
		L 01-299	N-21	N-27	HoCP 96-540	Ho 07-604	
		Ho 06-9610	N-24	US 93-15	Ho 07-617	Ho 07-613	
HoCP 85-845 (7 rows)							

↓
N

Plot size = 1 row, 5.25 ft row width, 12 ft long with 4 ft alley
Shaded plots = Seed increase as buffer

APENDIX B: AERIAL APPLICATION STUDY STATISTICAL ANALYSIS

INJURY AND EMERGENCY

```
dm'output;clear;log;clear';
Title1'LRGV All by Sample';
data data1;
data data1;
input Year$ Trt$ field$ Pos$ Bored Tot Emerg ;
cards;
2009 B 4 F 6 180 1
2009 B 5 B 31 205 8
2009 B 4 B 9 209 2
2009 B 5 F 24 192 4
2009 B 2 F 47 207 22
2009 B 1 F 36 174 10
2009 B 3 B 35 195 9
2009 B 3 F 36 192 14
2009 B 2 B 31 230 16
2009 B 1 B 33 231 3
2009 C 4 B 23 181 2
2009 C 5 F 27 220 11
2009 C 5 B 51 193 30
2009 C 2 F 27 207 12
2009 C 4 F 19 244 6
2009 C 3 B 17 207 1
2009 C 2 B 72 175 24
2009 C 3 F 24 219 3
2009 C 1 F 24 207 2
2009 C 1 B 42 188 11
2009 D 4 B 5 226 2
2009 D 5 B 23 219 14
2009 D 5 F 21 216 18
2009 D 2 B 30 202 8
2009 D 4 F 9 233 0
2009 D 2 F 66 209 27
2009 D 3 F 25 213 12
2009 D 3 B 11 199 0
2009 D 1 F 10 218 1
2009 D 1 B 9 220 2
2010 B 1 F 36 200 7
2010 B 1 B 38 222 16
2010 D 1 B 46 198 5
2010 D 1 F 46 219 4
2010 C 1 B 41 206 8
2010 C 1 F 76 185 14
2010 B 2 F 51 177 9
2010 B 2 B 39 189 8
2010 D 2 B 12 200 0
2010 D 2 F 14 187 2
2010 C 2 B 73 191 18
2010 C 2 F 139 363 36
2010 B 3 F 14 235 5
2010 B 3 B 7 197 1
```

2010	D	3	F	6	232	1
2010	D	3	B	22	233	7
2010	C	3	B	56	212	11
2010	C	3	F	42	222	9
2010	B	4	B	14	179	3
2010	B	4	F	4	202	0
2010	D	4	B	5	203	0
2010	D	4	F	6	220	2
2010	C	4	B	27	188	3
2010	C	4	F	39	200	12
2010	B	5	B	17	180	6
2010	B	5	F	16	209	2
2010	D	5	B	10	191	3
2010	D	5	F	13	205	1
2010	C	5	F	44	382	6
2010	C	5	B	39	188	8

```

;
proc glimmix data=data1 ;
class Year Trt field Pos;
model Bored/Tot = Trt / htype=3 ddfm=kr dist=binomial ;
random year field field*year field*year*trt;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

proc glimmix data=data1 ;
class Year Trt Field Pos;
model Emerg = Trt / htype=3 ddfm=kr dist=poisson ;
random year Field Field*year Field*year*trt ;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

proc glimmix data=data1 ;
class Year Trt Field Pos;
model Emerg/Bored = Trt / htype=3 ddfm=kr dist=binomial ;
random year Field Field*year Field*year*trt;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

proc glimmix data=data1 ;
class Year Trt Field Pos;

```



```

model Bored/Tot = Trt|year / htype=3 ddfm=kr dist=binomial ;
random year Field Field*year Field*year*trt;
lsmeans Trt|year / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

LRGV All 10:02 Thursday, March 31, 2011 10

The GLIMMIX Procedure

Probability of a Bored Internode

Model Information

Data Set	WORK.DATA1
Response Variable (Events)	Bored
Response Variable (Trials)	Tot
Response Distribution	Binomial
Link Function	Logit
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Residual PL
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Year	2	2009 2010
Trt	3	B C D
Field	5	1 2 3 4 5
Pos	2	B F

Number of Observations Read	60
Number of Observations Used	60
Number of Events	1815
Number of Trials	12626

Dimensions

G-side Cov. Parameters	4
Columns in X	4
Columns in Z	47
Subjects (Blocks in V)	1
Max Obs per Subject	60

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	4
Lower Boundaries	4
Upper Boundaries	0
Fixed Effects	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Subiterations	Objective		Max Gradient
			Function	Change	
0	0	5	187.02253872	0.11651819	41.77246
1	0	4	194.55097319	0.00527579	41.66498
2	0	2	194.8166387	0.00015906	41.6576
3	0	1	194.81735269	0.00004300	41.65712
4	0	1	194.81735262	0.00000582	41.65724
5	0	1	194.81735028	0.00000917	41.65713
6	0	1	194.81735109	0.00000683	41.65723
7	0	1	194.81735027	0.00000476	41.65716
8	0	1	194.8173512	0.00000491	41.65723
9	0	1	194.81735058	0.00000353	41.65717
10	0	1	194.81735125	0.00000361	41.65723
11	0	1	194.8173508	0.00000260	41.65719
12	0	1	194.8173513	0.00000266	41.65723
13	0	1	194.81735096	0.00000189	41.6572
14	0	1	194.81735132	0.00000195	41.65722
15	0	1	194.81735108	0.00000141	41.6572
16	0	1	194.81735134	0.00000145	41.65722
17	0	1	194.81735116	0.00000302	41.65718
18	0	1	194.81735173	0.00000279	41.65722
19	0	0	194.81735132	0.00000000	41.65722

Convergence criterion (PCONV=1.11022E-8) satisfied.

Estimated G matrix is not positive definite.

Fit Statistics

-2 Res Log Pseudo-Likelihood	194.82
Generalized Chi-Square	180.70
Gener. Chi-Square / DF	3.17

Covariance Parameter Estimates

Cov Parm	Estimate	Standard
		Error
Year	0	.
Field	0.2650	0.2318
Year*Field	0.03268	0.07646
Year*Trt*Field	0.2193	0.08169

Type III Tests of Fixed Effects

Effect	Num	Den	F Value	Pr > F
	DF	DF		
Trt	2	18.2	11.41	0.0006

----- Effect=Trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Trt	Estimate	Standard	Standard		Alpha	Lower	Upper	Lower	Upper	Letter
			Error	Mean	Mean						
1	C	-1.4435	0.2826	0.1910	0.04366	0.05	-2.1353	-0.7518	0.1057	0.3204	A

2	B	-2.0463	0.2848	0.1144	0.02886	0.05	-2.7384	-1.3542	0.06074	0.2052	B
3	D	-2.5000	0.2866	0.07586	0.02010	0.05	-3.1925	-1.8075	0.03945	0.1409	B

LRGV All by stalk 10:02 Thursday, March 31, 2011

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The GLIMMIX Procedure

Emergence

Model Information

Data Set	WORK.DATA1
Response Variable	Emerg
Response Distribution	Poisson
Link Function	Log
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Residual PL
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Year	2	2009 2010
Trt	3	B C D
Field	5	1 2 3 4 5
Pos	2	B F

Number of Observations Read	60
Number of Observations Used	60

Dimensions

G-side Cov. Parameters	4
Columns in X	4
Columns in Z	47
Subjects (Blocks in V)	1
Max Obs per Subject	60

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	4
Lower Boundaries	4
Upper Boundaries	0
Fixed Effects	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Subiterations	Objective Function	Change	Max Gradient
0	0	7	162.77438672	0.21549969	6.129949
1	0	5	180.08778862	0.04645105	6.25878
2	0	3	182.65464661	0.00529663	6.247226
3	0	2	182.73745635	0.00018619	6.245641
4	0	1	182.7375978	0.00007154	6.245852
5	0	1	182.73762526	0.00001618	6.245808
6	0	1	182.73759444	0.00001097	6.245846

7	0	1	182.73760536	0.00000588	6.245835
8	0	1	182.73759365	0.00000450	6.245851
9	0	1	182.73759797	0.00000230	6.245846
10	0	1	182.73759333	0.00000179	6.245852
11	0	1	182.73759502	0.00000090	6.245851
12	0	0	182.73759318	0.00000000	6.245851

Convergence criterion (PCONV=1.11022E-8) satisfied.

Estimated G matrix is not positive definite.

Fit Statistics

-2 Res Log Pseudo-Likelihood	182.74
Generalized Chi-Square	115.11
Gener. Chi-Square / DF	2.02

Covariance Parameter Estimates

Cov Parm	Estimate	Standard Error
Year	8.03E-19	.
Field	0.2170	0.2777
Year*Field	0.1412	0.1834
Year*Trt*Field	0.3418	0.1489

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
Trt	2	17.17	4.65	0.0244

----- Effect=Trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
1	C	2.2296	0.3131	9.2959	2.9106	0.05	1.4842	2.9750	4.4112	19.5894	A
2	B	1.7611	0.3194	5.8188	1.8583	0.05	1.0120	2.5102	2.7512	12.3069	AB
3	D	1.3125	0.3288	3.7155	1.2218	0.05	0.5564	2.0686	1.7444	7.9138	B

LRGV All by stalk 10:02 Thursday, March 31, 2011

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The GLIMMIX Procedure

Relative Survival

Model Information

Data Set	WORK.DATA1
Response Variable (Events)	Emerg
Response Variable (Trials)	Bored
Response Distribution	Binomial
Link Function	Logit
Variance Function	Default
Variance Matrix	Not blocked

Estimation Technique	Residual PL
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Year	2	2009 2010
Trt	3	B C D
Field	5	1 2 3 4 5
Pos	2	B F

Number of Observations Read	60
Number of Observations Used	60
Number of Events	482
Number of Trials	1815

Dimensions

G-side Cov. Parameters	4
Columns in X	4
Columns in Z	47
Subjects (Blocks in V)	1
Max Obs per Subject	60

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	4
Lower Boundaries	4
Upper Boundaries	0
Fixed Effects	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Subiterations	Objective Function	Change	Max Gradient
0	0	13	145.1021998	1.85589123	4.276594
1	0	5	149.91541795	0.08559157	4.218059
2	0	3	150.18429711	0.00136691	4.214693
3	0	1	150.18842522	0.00002217	4.214747
4	0	1	150.18849276	0.00001862	4.214536
5	0	1	150.1884777	0.00000549	4.214612
6	0	1	150.18848604	0.00000119	4.21461
7	0	1	150.18848422	0.00000086	4.214627
8	0	1	150.18848701	0.00000150	4.2146
9	0	1	150.18848311	0.00000092	4.214616
10	0	0	150.18848533	0.00000000	4.214616

Convergence criterion (PCONV=1.11022E-8) satisfied.

Estimated G matrix is not positive definite.

Fit Statistics

-2 Res Log Pseudo-Likelihood	150.19
Generalized Chi-Square	77.49
Gener. Chi-Square / DF	1.36

Cov Parm	Estimate	Standard Error
Year	0.08162	0.1866
Field	3.45E-20	.
Year*Field	0.1135	0.1224
Year*Trt*Field	0.2391	0.1168

Effect	Num	Den	F Value	Pr > F
	DF	DF		
Trt	2	18.65	0.11	0.8949

----- Effect=Trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Trt	Estimate	Standard Error	Standard Error of		Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
				Mean	Mean						
1	B	-1.0464	0.2998	0.2599	0.05768	0.05	-2.4184	0.3256	0.08178	0.5807	A
2	D	-1.1237	0.3125	0.2453	0.05786	0.05	-2.3563	0.1088	0.08657	0.5272	A
3	C	-1.1706	0.2900	0.2367	0.05240	0.05	-2.7014	0.3602	0.06289	0.5891	A

LRGV All by stalk 10:02 Thursday, March 31, 2011

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The GLIMMIX Procedure

Probability of a Bored Internode Year*Treatment

Model Information

Data Set	WORK.DATA1
Response Variable (Events)	Bored
Response Variable (Trials)	Tot
Response Distribution	Binomial
Link Function	Logit
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Residual PL
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Year	2	2009 2010
Trt	3	B C D
Field	5	1 2 3 4 5
Pos	2	B F

Number of Observations Read	60
Number of Observations Used	60

Number of Events 1815
 Number of Trials 12626

Dimensions
 G-side Cov. Parameters 4
 Columns in X 12
 Columns in Z 47
 Subjects (Blocks in V) 1
 Max Obs per Subject 60

Optimization Information
 Optimization Technique Dual Quasi-Newton
 Parameters in Optimization 4
 Lower Boundaries 4
 Upper Boundaries 0
 Fixed Effects Profiled
 Starting From Data

Iteration History

Iteration	Restarts	Subiterations	Objective Function	Change	Max Gradient
0	0	5	184.50505566	0.24271646	1.283E-6
1	0	4	192.05945964	0.00875641	0.000041
2	0	3	192.32627574	0.00017295	5.047E-6
3	0	1	192.32691833	0.00000090	5.942E-6
4	0	0	192.32691882	0.00000000	5.901E-6

Convergence criterion (PCONV=1.11022E-8) satisfied.

Estimated G matrix is not positive definite.

Fit Statistics
 -2 Res Log Pseudo-Likelihood 192.33
 Generalized Chi-Square 178.31
 Gener. Chi-Square / DF 3.30

Covariance Parameter Estimates
 Standard Error
 Cov Parm Estimate Error
 Year 0 .
 Field 0.2515 0.2341
 Year*Field 0.06898 0.1017
 Year*Trt*Field 0.1895 0.07637

Type III Tests of Fixed Effects
 Num Den F Value Pr > F
 Effect DF DF
 Trt 2 16.09 12.88 0.0005
 Year 1 4.088 0.02 0.8981
 Year*Trt 2 16.09 2.21 0.1414

LRGV All by stalk 10:02 Thursday, March 31, 2011

----- Effect=Trt Method=Tukey-Kramer(P<.05) Set=1 -----

Letter	Obs	Year	Trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	
	1		C	-1.4473	0.2789	0.1904	0.04300	0.05	-2.1383	-0.7563	0.1054	0.3195	A
	2		B	-2.0471	0.2812	0.1143	0.02848	0.05	-2.7382	-1.3560	0.06075	0.2049	B
	3		D	-2.4996	0.2830	0.07589	0.01985	0.05	-3.1909	-1.8082	0.03951	0.1409	B

----- Effect=Year Method=Tukey-Kramer(P<.05) Set=2 -----

Letter	Obs	Year	Trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	
	4	2010		-1.9818	0.2806	0.1211	0.02987	0.05	-2.6783	-1.2853	0.06427	0.2167	A
	5	2009		-2.0142	0.2803	0.1177	0.02911	0.05	-2.7107	-1.3177	0.06234	0.2112	A

----- Effect=Year*Trt Method=Tukey-Kramer(P<.05) Set=3 -----

Letter	Obs	Year	Trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	
	6	2010	C	-1.1836	0.3234	0.2344	0.05804	0.05	-1.9093	-0.4579	0.1291	0.3875	A
	7	2009	C	-1.7110	0.3256	0.1530	0.04220	0.05	-2.4388	-0.9832	0.08026	0.2723	AB
	8	2009	B	-1.9097	0.3274	0.1290	0.03679	0.05	-2.6395	-1.1799	0.06664	0.2351	AB
	9	2010	B	-2.1845	0.3294	0.1011	0.02995	0.05	-2.9164	-1.4526	0.05135	0.1896	B
	10	2009	D	-2.4219	0.3306	0.08152	0.02475	0.05	-3.1551	-1.6888	0.04089	0.1559	B
	11	2010	D	-2.5772	0.3325	0.07062	0.02182	0.05	-3.3125	-1.8419	0.03515	0.1368	B

LINEAR REGRESSION PHEROMONE TRAP CATCH VS LARVAL INFESTATION

```
dm'output;clear;log;clear';
Title'Catch vs Larval Infestation';
data data1;
input DATE$ FIELD$ Trap$ CATCH Infst;
cards;
1-Jun 1 A 3 0 1-Jun 4 A 2 0
1-Jun 1 B 1 0 1-Jun 4 B 1 5
1-Jun 2 A 2 0 1-Jun 5 A 0 0
1-Jun 2 B 0 0 1-Jun 5 B 0 0
1-Jun 3 A 0 0 8-Jun 1 A 2 0
1-Jun 3 B 0 0 8-Jun 1 B 1 0
```


8-Jun	2	A	2	0	13-Jul	5	A	3	0
8-Jun	2	B	5	0	13-Jul	5	B	7	0
8-Jun	3	A	1	0	20-Jul	1	A	11	5
8-Jun	3	B	4	0	20-Jul	1	B	9	0
8-Jun	4	A	0	0	20-Jul	2	A	8	0
8-Jun	4	B	0	0	20-Jul	2	B	14	10
8-Jun	5	A	0	0	20-Jul	3	A	19	10
8-Jun	5	B	2	0	20-Jul	3	B	22	10
15-Jun	1	A	1	0	20-Jul	4	A	7	0
15-Jun	1	B	2	0	20-Jul	4	B	10	0
15-Jun	2	A	2	0	20-Jul	5	A	6	2.5
15-Jun	2	B	4	0	20-Jul	5	B	8	0
15-Jun	3	A	5	0	27-Jul	1	A	17	2.5
15-Jun	3	B	3	0	27-Jul	1	B	13	0
15-Jun	4	A	2	0	27-Jul	2	A	10	5
15-Jun	4	B	0	0	27-Jul	2	B	29	10
15-Jun	5	A	1	0	27-Jul	3	A	31	10
15-Jun	5	B	5	0	27-Jul	3	B	17	5
22-Jun	1	A	4	0	27-Jul	4	A	12	0
22-Jun	1	B	6	0	27-Jul	4	B	9	2.5
22-Jun	2	A	5	0	27-Jul	5	A	14	2.5
22-Jun	2	B	4	0	27-Jul	5	B	19	0
22-Jun	3	A	7	0	4-Aug	1	A	21	7.5
22-Jun	3	B	8	0	4-Aug	1	B	29	10
22-Jun	4	A	3	0	4-Aug	2	A	31	10
22-Jun	4	B	4	0	4-Aug	2	B	11	0
22-Jun	5	A	2	0	4-Aug	3	A	24	2.5
22-Jun	5	B	6	0	4-Aug	3	B	37	5
29-Jun	1	A	8	0	4-Aug	4	A	17	5
29-Jun	1	B	7	0	4-Aug	4	B	21	2.5
29-Jun	2	A	6	0	4-Aug	5	A	9	0
29-Jun	2	B	4	0	4-Aug	5	B	24	7.5
29-Jun	3	A	6	0	11-Aug	1	A	33	10
29-Jun	3	B	8	0	11-Aug	1	B	17	7.5
6-Jul	1	A	5	0	11-Aug	2	A	22	12.5
6-Jul	1	B	8	0	11-Aug	2	B	41	10
6-Jul	2	A	6	0	11-Aug	3	A	53	20
6-Jul	2	B	3	0	11-Aug	3	B	36	17.5
6-Jul	3	A	6	0	11-Aug	4	A	22	5
6-Jul	3	B	4	0	11-Aug	4	B	27	2.5
6-Jul	4	A	2	0	11-Aug	5	A	14	0
6-Jul	4	B	3	0	11-Aug	5	B	26	5
6-Jul	5	A	3	0	13-Aug	1	A	56	12.5
6-Jul	5	B	4	0	13-Aug	1	B	40	10
13-Jul	1	A	9	5	13-Aug	2	A	31.5	5
13-Jul	1	B	7	5	13-Aug	2	B	77	10
13-Jul	2	A	8	2.5	13-Aug	3	A	108.5	22.5
13-Jul	2	B	11	2.5	13-Aug	3	B	73.5	15
13-Jul	3	A	12	5	13-Aug	4	A	49	7.5
13-Jul	3	B	16	5	13-Aug	4	B	73.5	10
13-Jul	4	A	6	0	13-Aug	5	A	56	7.5
13-Jul	4	B	4	0	13-Aug	5	B	70	12.5

```

;
proc reg data=data1;
model Infst = catch;

```

run;

Catch vs Larval Infestation

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2011

The REG Procedure
Model: MODEL1
Dependent Variable: Infst

Number of Observations Read 116
Number of Observations Used 116

Analysis of Variance

Source	DF	Sum of Squares	Mean Square	F Value	Pr > F
Model	1	1896.74901	1896.74901	280.74	<.0001
Error	114	770.22297	6.75634		
Corrected Total	115	2666.97198			

Root MSE 2.59930 R-Square 0.7112
Dependent Mean 3.08190 Adj R-Sq 0.7087
Coeff Var 84.34081

Parameter Estimates

Variable	DF	Parameter Estimate	Standard Error	t Value	Pr > t
Intercept	1	-0.03833	0.30483	-0.13	0.9002
CATCH	1	0.21278	0.01270	16.76	<.0001

2010 YIELD AND QUALITY

SUGAR PER ACRE

```
dm'output;clear;log;clear';
Title1'LRGV 2010 Yield';
data data1;
input field$ trt$ Pur Fib Brix POL CRS TCA TSA;
```

cards;

1	D	86.98	17.48	16.64	14.47	208	28.14	2.93
1	C	85.92	18.12	16.77	14.40	204	29.69	3.04
1	B	85.45	17.64	17.34	14.82	211	28.79	3.04
2	D	84.78	18.92	17.25	14.62	205	27.19	2.79
2	C	81.43	18.71	16.58	13.51	185	21.40	1.99
2	B	84.67	18.71	16.48	13.95	195	18.12	1.77
4	D	84.94	15.39	17.22	14.63	211	34.51	3.65
4	C	84.54	15.36	16.65	14.07	203	31.36	3.19
4	B	85.34	15.01	16.47	14.05	204	30.53	3.13
5	D	84.82	14.99	17.56	14.90	216	28.24	3.06
5	C	84.21	15.87	17.13	14.42	207	27.59	2.86
5	B	85.11	15.59	16.71	14.22	206	25.84	2.66

3	D	84.85	17.34	16.52	14.02	199	33.90	3.38
3	C	85.14	19.30	15.65	13.32	186	29.59	2.76
3	B	84.77	18.06	16.40	13.90	196	24.01	2.36

```

;
proc glimmix data=data1 ;
class Field Trt ;
model TSA = Trt / htype=3 ddfm=kr dist=Gaussian ;
random Field ;
lsmeans Trt / ilink diff cl ;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

```

proc glimmix data=data1 ;
class Field Trt ;
model TCA = Trt / htype=3 ddfm=kr dist=Gaussian ;
random Field ;
lsmeans Trt / ilink diff cl ;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

LRGV 2010 Yield 10:02 Thursday, March 31, 2011 25
The GLIMMIX Procedure

Sugar/Acre

Model Information

Data Set	WORK.DATA1
Response Variable	TSA
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
field	5	1 2 3 4 5
trt	3	B C D

Number of Observations Read	15
Number of Observations Used	15

Dimensions

G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	4
Columns in Z	5
Subjects (Blocks in V)	1
Max Obs per Subject	15

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	14.120093624	.	1.89E-15

Convergence criterion (ABSGCONV=0.00001) satisfied.

Fit Statistics

-2 Res Log Likelihood	14.12
AIC (smaller is better)	18.12
AICC (smaller is better)	19.45
BIC (smaller is better)	17.34
CAIC (smaller is better)	19.34
HQIC (smaller is better)	16.02
Generalized Chi-Square	0.75
Gener. Chi-Square / DF	0.06

Covariance Parameter Estimates

Cov Parm	Estimate	Standard Error
field	0.1520	0.1228
Residual	0.06283	0.03141

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
trt	2	8	6.78	0.0190

----- Effect=trt Method=LSD(P<.05) Set=1 -----

Standard	Standard Error of	Lower	Upper	Letter
----------	-------------------	-------	-------	--------

Obs	trt	Estimate	Error	Mean	Mean	Alpha	Lower	Upper	Mean	Mean	Group
1	D	3.1620	0.2073	3.1620	0.2073	0.05	2.6547	3.6693	2.6547	3.6693	A
2	C	2.7680	0.2073	2.7680	0.2073	0.05	2.2607	3.2753	2.2607	3.2753	B
3	B	2.5920	0.2073	2.5920	0.2073	0.05	2.0847	3.0993	2.0847	3.0993	B

LRGV 2010 Yield

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Cane/acre

The GLIMMIX Procedure
Model Information

Data Set WORK.DATA1
 Response Variable TCA
 Response Distribution Gaussian
 Link Function Identity
 Variance Function Default
 Variance Matrix Not blocked
 Estimation Technique Restricted Maximum Likelihood
 Degrees of Freedom Method Kenward-Roger
 Fixed Effects SE Adjustment Kenward-Roger

Class Level Information

Class	Levels	Values
field	5	1 2 3 4 5
trt	3	B C D

Number of Observations Read 15
 Number of Observations Used 15

Dimensions

G-side Cov. Parameters 1
 R-side Cov. Parameters 1
 Columns in X 4
 Columns in Z 5
 Subjects (Blocks in V) 1
 Max Obs per Subject 15

Optimization Information

Optimization Technique Dual Quasi-Newton
 Parameters in Optimization 1
 Lower Boundaries 1
 Upper Boundaries 0
 Fixed Effects Profiled
 Residual Variance Profiled
 Starting From Data

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	67.161408321	.	8.33E-16

Convergence criterion (ABSGCONV=0.00001) satisfied.

Fit Statistics	
-2 Res Log Likelihood	67.16
AIC (smaller is better)	71.16
AICC (smaller is better)	72.49
BIC (smaller is better)	70.38
CAIC (smaller is better)	72.38
HQIC (smaller is better)	69.06
Generalized Chi-Square	65.26
Gener. Chi-Square / DF	5.44

Covariance Parameter Estimates		
Cov Parm	Estimate	Standard Error
field	11.4373	9.4130
Residual	5.4385	2.7193

Type III Tests of Fixed Effects				
Effect	Num DF	Den DF	F Value	Pr > F
trt	2	8	5.60	0.0301

----- Effect=trt Method=LSD(P<.05) Set=1 -----

Obs	trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
1	D	30.3960	1.8372	30.3960	1.8372	0.05	25.9446	34.8474	25.9446	34.8474	A
2	C	27.9260	1.8372	27.9260	1.8372	0.05	23.4746	32.3774	23.4746	32.3774	AB
3	B	25.4580	1.8372	25.4580	1.8372	0.05	21.0066	29.9094	21.0066	29.9094	B

QUALITY PARAMETERS

```
dm'output;clear;log;clear';
Title1'LRGV 2010 Yield Quality';
data data1;
input field$ trt$ Pur Brix POL CRS ;

cards;
5 D 86.58 17.80 15.42 228
5 D 86.17 17.03 14.68 212
5 D 86.00 17.87 15.37 227
5 D 84.92 17.68 15.01 219
5 D 82.06 17.78 14.59 210
5 D 84.47 17.63 14.89 214
5 D 84.78 17.46 14.80 216
5 D 83.59 17.23 14.40 208
5 C 83.76 17.73 14.85 213
5 C 82.74 17.34 14.35 203
5 C 84.00 17.67 14.84 216
5 C 84.08 17.42 14.64 210
```

5	C	84.04	17.13	14.39	205
5	C	85.09	17.26	14.69	211
5	C	84.41	16.37	13.82	197
5	C	82.70	16.97	14.03	201
5	C	84.63	17.13	14.50	210
5	C	82.45	17.11	14.10	203
5	C	83.96	16.64	13.97	199
5	C	87.47	17.17	15.01	222
5	C	85.35	16.70	14.25	205
5	B	85.45	15.51	13.26	191
5	B	85.02	17.04	14.49	211
5	B	85.27	16.93	14.44	209
5	B	85.71	16.65	14.27	203
5	B	84.31	17.30	14.59	211
5	B	84.89	16.80	14.26	211
4	D	85.11	18.26	15.54	226
4	D	85.02	17.62	14.98	215
4	D	86.66	17.23	14.93	218
4	D	85.77	16.55	14.19	204
4	D	86.16	17.20	14.82	214
4	D	83.35	17.12	14.27	207
4	D	84.09	16.64	13.99	204
4	D	83.33	17.14	14.29	206
4	C	82.60	17.90	14.79	215
4	C	83.06	17.26	14.34	207
4	C	84.71	16.52	13.99	203
4	C	85.62	16.16	13.83	201
4	C	85.64	16.11	13.80	199
4	C	85.60	15.94	13.65	197
4	B	85.61	16.50	14.13	205
4	B	84.58	16.23	13.73	198
4	B	84.76	16.45	13.94	203
4	B	83.45	17.00	14.18	209
4	B	83.52	15.65	13.07	186
4	B	87.02	16.60	14.44	213
4	B	86.42	16.63	14.37	213
4	B	85.19	16.88	14.38	207
4	B	86.66	16.36	14.18	206
4	B	86.17	16.35	14.09	208
3	D	84.61	16.20	13.71	197.00
3	D	85.12	17.12	14.57	210.00
3	D	85.76	17.01	14.59	211.00
3	D	85.45	16.21	13.85	198.00
3	D	85.62	16.56	14.18	204.00
3	D	85.27	16.77	14.30	204.00
3	D	83.69	16.03	13.41	189.00
3	D	83.49	17.24	14.39	205.00
3	D	83.61	16.57	13.85	195.00
3	D	84.82	15.97	13.54	187.00
3	D	85.91	16.04	13.78	196.00
3	C	85.41	15.81	13.51	192.00
3	C	85.43	15.35	13.11	180.00
3	C	85.80	15.61	13.40	189.00
3	C	83.18	16.59	13.80	194.00
3	C	83.06	16.55	13.74	197.00

3	C	83.86	15.84	13.28	190.00
3	C	83.22	15.15	12.61	173.00
3	C	84.12	15.07	12.67	175.00
3	C	87.73	14.86	13.04	177.00
3	C	86.58	16.16	13.99	196.00
3	C	87.23	14.77	12.89	176.00
3	C	86.07	16.06	13.82	197.00
3	B	82.94	16.23	13.46	189.00
3	B	86.42	16.58	14.33	204.00
3	B	85.65	16.45	14.09	197.00
3	B	84.05	16.34	13.73	197.00
2	C	84.30	17.06	14.38	202
2	C	79.84	15.44	12.33	164
2	C	80.81	15.87	12.82	174
2	C	82.64	17.08	14.12	197
2	C	82.72	17.17	14.20	198
2	C	78.25	16.88	13.21	179
2	D	82.95	17.50	14.51	196
2	D	85.07	16.73	14.23	197
2	D	84.24	16.70	14.07	197
2	D	83.53	16.28	13.60	189
2	D	86.03	17.25	14.84	211
2	D	86.80	17.05	14.80	211
2	D	85.00	18.13	15.41	219
2	D	84.52	18.33	15.49	219
2	D	83.99	17.52	14.71	204
2	D	85.70	16.99	14.56	207
2	B	85.14	16.62	14.15	195
2	B	85.77	16.20	13.90	195
2	B	84.69	16.12	13.65	192
2	B	84.36	16.33	13.78	196
2	B	84.95	16.16	13.72	192
2	B	83.08	17.46	14.51	204
1	D	85.97	17.23	14.81	214
1	D	84.42	16.64	14.04	199
1	D	85.60	17.04	14.59	210
1	D	88.30	16.06	14.18	201
1	D	87.71	16.57	14.54	210
1	D	87.98	17.15	15.09	223
1	D	88.34	17.07	15.08	220
1	D	86.40	17.03	14.71	214
1	D	85.97	16.44	14.14	203
1	D	89.07	15.20	13.54	189
1	C	88.42	16.16	14.29	206
1	C	87.73	16.25	14.25	204
1	C	86.08	16.51	14.21	198
1	C	88.50	15.71	13.90	199
1	C	84.75	16.68	14.13	203
1	C	84.15	16.75	14.09	202
1	C	85.25	16.46	14.03	201
1	C	84.23	17.42	14.67	204
1	C	84.05	17.53	14.73	205
1	C	86.08	18.24	15.70	224
1	B	86.48	17.28	14.95	210
1	B	85.11	17.67	15.04	213

1	B	86.40	18.23	15.75	229
1	B	86.00	17.31	14.89	211
1	B	83.22	17.89	14.88	214
1	B	86.91	17.50	15.21	220
1	B	84.92	16.83	14.29	203
1	B	85.55	17.20	14.72	210
1	B	84.75	16.63	14.09	197
1	B	85.17	16.87	14.37	206

```

;
ODS HTML FILE='C:\Documents and Settings\treagan\Desktop\Blake Wilson\LRGV
2010 Yield Quality.html' style = minimal
;
proc glimmix data=data1 ;
class Field Trt ;
model Pol = Trt / htype=3 ddfm=kr dist=Gaussian ;
random Field ;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;
proc glimmix data=data1 ;
class Field Trt ;
model Pur = Trt / htype=3 ddfm=kr dist=Gaussian ;
random Field ;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;
proc glimmix data=data1 ;
class Field Trt ;
model brix = Trt / htype=3 ddfm=kr dist=Gaussian ;
random Field ;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;
proc glimmix data=data1 ;
class Field Trt ;
model CRS = Trt / htype=3 ddfm=kr dist=Gaussian ;
random Field ;
lsmeans Trt / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);

```

run;

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The GLIMMIX Procedure
Percent Sucrose (POL)
Model Information

Data Set	WORK.DATA1
Response Variable	POL
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information
Class Levels Values

field	5	1	2	3	4	5
trt	3	B	C	D		

Number of Observations Read	130
Number of Observations Used	130

Dimensions

G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	4
Columns in Z	5
Subjects (Blocks in V)	1
Max Obs per Subject	130

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	209.65163212	.	0.786074
1	0	3	209.62438108	0.02725105	0.27281
2	0	2	209.62176364	0.00261744	0.087835
3	0	2	209.62150377	0.00025987	0.006761
4	0	2	209.62150217	0.00000160	0.000153
5	0	2	209.62150216	0.00000000	2.74E-7

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics

-2 Res Log Likelihood	209.62
AIC (smaller is better)	213.62
AICC (smaller is better)	213.72
BIC (smaller is better)	212.84
CAIC (smaller is better)	214.84
HQIC (smaller is better)	211.53
Generalized Chi-Square	32.67
Gener. Chi-Square / DF	0.26

Covariance Parameter Estimates

		Standard
Cov Parm	Estimate	Error
field	0.1244	0.09467
Residual	0.2573	0.03280

Type III Tests of Fixed Effects

	Num	Den		
Effect	DF	DF	F Value	Pr > F
trt	2	123.6	13.94	<.0001

----- Effect=trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
1	D	14.5192	0.1743	14.5192	0.1743	0.05	14.0749	14.9634	14.0749	14.9634	A
2	B	14.1965	0.1796	14.1965	0.1796	0.05	13.7530	14.6400	13.7530	14.6400	B
3	C	13.9637	0.1748	13.9637	0.1748	0.05	13.5194	14.4081	13.5194	14.4081	B

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The GLIMMIX Procedure
Juice Purity

Model Information

Data Set	WORK.DATA1
Response Variable	Pur
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
field	5	1 2 3 4 5
trt	3	B C D

Number of Observations Read 130
 Number of Observations Used 130

Dimensions
 G-side Cov. Parameters 1
 R-side Cov. Parameters 1
 Columns in X 4
 Columns in Z 5
 Subjects (Blocks in V) 1
 Max Obs per Subject 130

Optimization Information
 Optimization Technique Dual Quasi-Newton
 Parameters in Optimization 1
 Lower Boundaries 1
 Upper Boundaries 0
 Fixed Effects Profiled
 Residual Variance Profiled
 Starting From Data

Iteration History						
Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient	
0	0	4	477.68892338	.	0.433986	
1	0	4	477.68602878	0.00289460	0.020752	
2	0	2	477.6860228	0.00000598	0.001662	
3	0	2	477.68602276	0.00000004	5.776E-6	

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics
 -2 Res Log Likelihood 477.69
 AIC (smaller is better) 481.69
 AICC (smaller is better) 481.78
 BIC (smaller is better) 480.90
 CAIC (smaller is better) 482.90
 HQIC (smaller is better) 479.59
 Generalized Chi-Square 273.67
 Gener. Chi-Square / DF 2.15

Covariance Parameter Estimates

Cov Parm	Estimate	Standard Error
field	0.6233	0.5003
Residual	2.1549	0.2748

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
trt	2	123.9	4.15	0.0180

----- Effect=trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
1	D	85.2887	0.4133	85.2887	0.4133	0.05	84.2698	86.3077	84.2698	86.3077	A
2	B	85.0371	0.4317	85.0371	0.4317	0.05	84.0127	86.0615	84.0127	86.0615	AB
3	C	84.4252	0.4151	84.4252	0.4151	0.05	83.4049	85.4455	83.4049	85.4455	B

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The GLIMMIX Procedure
Brix
Model Information

Data Set WORK.DATA1
Response Variable Brix
Response Distribution Gaussian
Link Function Identity
Variance Function Default
Variance Matrix Not blocked
Estimation Technique Restricted Maximum Likelihood
Degrees of Freedom Method Kenward-Roger
Fixed Effects SE Adjustment Kenward-Roger

Class Level Information
Class Levels Values
field 5 1 2 3 4 5
trt 3 B C D

Number of Observations Read 130
Number of Observations Used 130

Dimensions
G-side Cov. Parameters 1
R-side Cov. Parameters 1
Columns in X 4
Columns in Z 5
Subjects (Blocks in V) 1
Max Obs per Subject 130

Optimization Information
Optimization Technique Dual Quasi-Newton
Parameters in Optimization 1
Lower Boundaries 1
Upper Boundaries 0
Fixed Effects Profiled
Residual Variance Profiled
Starting From Data

Iteration History
Iteration Restarts Evaluations Objective Function Change Max Gradient
0 0 4 255.41715996 . 0.774658
1 0 3 255.39806588 0.01909407 0.036456

2	0	2	255.39803462	0.00003126	0.008119
3	0	2	255.39803302	0.00000161	0.000066
4	0	2	255.39803302	0.00000000	1.183E-7

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics

-2 Res Log Likelihood	255.40
AIC (smaller is better)	259.40
AICC (smaller is better)	259.49
BIC (smaller is better)	258.62
CAIC (smaller is better)	260.62
HQIC (smaller is better)	257.30
Generalized Chi-Square	47.12
Gener. Chi-Square / DF	0.37

Covariance Parameter Estimates

Cov Parm	Estimate	Standard Error
field	0.1477	0.1142
Residual	0.3710	0.04730

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
trt	2	123.7	7.47	0.0009

----- Effect=trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
1	D	17.0267	0.1936	17.0267	0.1936	0.05	16.5393	17.5141	16.5393	17.5141	A
2	B	16.6949	0.2004	16.6949	0.2004	0.05	16.2074	17.1825	16.2074	17.1825	B
3	C	16.5469	0.1943	16.5469	0.1943	0.05	16.0593	17.0346	16.0593	17.0346	B

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The GLIMMIX Procedure

CRS

Model Information

Data Set	WORK.DATA1
Response Variable	CRS
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
field	5	1 2 3 4 5
trt	3	B C D

Number of Observations Read 130
Number of Observations Used 130

Dimensions

G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	4
Columns in Z	5
Subjects (Blocks in V)	1
Max Obs per Subject	130

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	937.43470262	.	0.09258
1	0	4	937.43405478	0.00064784	0.004957
2	0	2	937.434053	0.00000178	0.000183

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics

-2 Res Log Likelihood	937.43
AIC (smaller is better)	941.43
AICC (smaller is better)	941.53
BIC (smaller is better)	940.65
CAIC (smaller is better)	942.65
HQIC (smaller is better)	939.34
Generalized Chi-Square	9957.28
Gener. Chi-Square / DF	78.40

Covariance Parameter Estimates

Cov Parm	Estimate	Standard Error
field	55.5866	41.4728
Residual	78.4038	9.9975

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
trt	2	123.4	16.03	<.0001

----- Effect=trt Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	trt	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
1	D	208.24	3.5771	208.24	3.5771	0.05	198.90	217.58	198.90	217.58	A
2	B	202.99	3.6556	202.99	3.6556	0.05	193.70	212.29	193.70	212.29	B
3	C	197.79	3.5847	197.79	3.5847	0.05	188.46	207.13	188.46	207.13	C

APENDIX C

NEONATE ESTABLISHMENT STATISTICAL ANALYSIS

```

Dm 'output;clear;log;clear';
Title1'GH by plant';
data data1;
input Age$ Var$ Rep$ Eggs Hatch Midrib Vis Est Entry Surv;
cards;
I 950 1 39 18 2 1 3 3 1 M 950 1 73 56 7 11 18 8 0
I 950 2 22 12 2 1 3 1 1 M 950 2 37 34 6 14 20 13 1
I 950 3 46 31 11 1 12 2 1 M 950 3 86 51 4 6 10 1 0
I 950 4 35 19 2 0 2 1 1 M 950 4 20 20 6 8 14 5 0
I 950 5 24 16 1 2 3 3 2 M 950 5 43 19 5 7 12 4 0
I 950 6 47 22 1 3 4 3 1 M 950 6 77 58 8 6 14 8 0
I 950 7 14 8 1 0 1 1 1 M 950 7 64 39 4 4 8 3 0
I 950 8 14 13 1 2 3 3 1 M 950 8 37 22 2 6 8 1 0
I 950 9 24 15 2 0 2 1 1 M 950 9 48 33 5 4 9 0 0
I 950 10 42 26 1 1 2 2 1 M 950 10 63 51 4 2 6 2 0
I 950 11 47 26 1 1 2 2 1 M 950 11 41 17 3 2 5 1 0
I 950 12 20 14 2 1 3 2 0 M 950 12 40 33 3 7 10 9 0
I 845 1 29 27 1 4 5 2 1 M 845 1 70 59 0 9 9 3 0
I 845 2 20 16 1 2 3 2 0 M 845 2 50 27 2 4 6 1 0
I 845 3 27 13 0 1 1 1 0 M 845 3 48 41 0 3 3 1 0
I 845 4 26 21 2 1 3 2 1 M 845 4 45 34 4 4 8 2 0
I 845 5 23 16 2 0 2 1 0 M 845 5 32 19 3 4 7 2 0
I 845 6 18 9 0 1 1 1 1 M 845 6 30 16 1 3 4 0 0
I 845 7 22 6 0 2 2 2 1 M 845 7 49 29 3 5 8 2 0
I 845 8 27 17 0 2 2 2 2 M 845 8 29 21 1 3 4 1 0
I 845 9 54 26 0 2 2 2 1 M 845 9 36 27 0 3 3 1 0
I 845 10 20 19 0 3 3 2 1 M 845 10 48 28 6 4 10 4 0
I 845 11 26 21 0 3 3 3 2 M 845 11 34 10 1 3 4 1 0
I 845 12 22 15 1 1 2 1 0 M 845 12 38 23 3 4 7 2 0
;proc glimmix data=data1 ;
class Age Var Rep;
model Midrib/est = Age|Var / htype=3 ddfm=kr dist=binomial ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

proc glimmix data=data1 ;
class Age Var Rep;
model Est/Hatch = Age|Var / htype=3 ddfm=kr dist=binomial ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

```

proc glimmix data=data1 ;
class Age Var Rep;
model Entry/Est = Age|Var / htype=3 ddfm=kr dist=binomial ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

Proc glimmix data=data1 ;
class Age Var Rep;
model Surv/Est = Age|Var / htype=3 ddfm=kr dist=binomial ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

proc glimmix data=data1 ;
class Age Var Rep;
model Entry/Hatch = Age|Var / htype=3 ddfm=kr dist=binomial ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

GH by plant

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The GLIMMIX Procedure

Midrib Entry

Model Information

Data Set	WORK.DATA1
Response Variable (Events)	Midrib
Response Variable (Trials)	Est
Response Distribution	Binomial
Link Function	Logit
Variance Function	Default
Variance Matrix	Diagonal
Estimation Technique	Maximum Likelihood
Degrees of Freedom Method	Residual

Class Level Information

Class	Levels	Values
Age	2	I M
Var	2	845 950
Rep	12	1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read	48
Number of Observations Used	48

Number of Events 115
 Number of Trials 276

Dimensions
 Columns in X 9
 Columns in Z 0
 Subjects (Blocks in V) 1
 Max Obs per Subject 48

Optimization Information
 Optimization Technique Newton-Raphson
 Parameters in Optimization 4
 Lower Boundaries 0
 Upper Boundaries 0
 Fixed Effects Not Profiled

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	65.067131257	.	1.489971
1	0	3	64.831676436	0.23545482	0.029752
2	0	3	64.831579122	0.00009731	0.000018
3	0	3	64.831579122	0.00000000	5.72E-12

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics

-2 Log Likelihood	129.66
AIC (smaller is better)	137.66
AICC (smaller is better)	138.59
BIC (smaller is better)	145.15
CAIC (smaller is better)	149.15
HQIC (smaller is better)	140.49
Pearson Chi-Square	47.21
Pearson Chi-Square / DF	1.07

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
Age	1	44	0.91	0.3447
Var	1	44	13.27	0.0007
Age*Var	1	44	5.42	0.0246

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Mean	Alpha	Lower	Upper	Lower	Upper	Ltter
										Mean	Mean	Group
1	I		-0.2071	0.2749	0.4484	0.06799	0.05	-0.7611	0.3469	0.3184	0.5859	A
2	M		-0.5073	0.1522	0.3758	0.03569	0.05	-0.8139	-0.2006	0.3071	0.4500	A

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Standard

Letter	Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Group
	3		950	0.2151	0.1901	0.5536	0.04697	0.05	-0.1680	0.5981	0.4581	0.6452	A
	4		845	-0.9294	0.2502	0.2830	0.05077	0.05	-1.4337	-0.4252	0.1925	0.3953	B

----- Effect=Age*Var Method=Tukey-Kramer (P<.05) Set=3 -----

Letter	Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Group
	5	I	950	0.7309	0.3376	0.6750	0.07406	0.05	0.05054	1.4112	0.5126	0.8040	A
	6	M	950	-0.3008	0.1747	0.4254	0.04271	0.05	-0.6529	0.05139	0.3423	0.5128	B
	7	M	845	-0.7138	0.2491	0.3288	0.05498	0.05	-1.2159	-0.2116	0.2287	0.4473	B
	8	I	845	-1.1451	0.4339	0.2414	0.07946	0.05	-2.0197	-0.2706	0.1172	0.4328	B

The GLIMMIX Procedure
Percent Established

Model Information

```

Data Set                WORK.DATA1
Response Variable (Events) Est
Response Variable (Trials) Hatch
Response Distribution   Binomial
Link Function           Logit
Variance Function      Default
Variance Matrix        Diagonal
Estimation Technique   Maximum Likelihood
Degrees of Freedom Method Residual
  
```

Class Level Information

```

Class   Levels   Values
Age      2      I M
Var      2      845 950
Rep      12     1 10 11 12 2 3 4 5 6 7 8 9
  
```

```

Number of Observations Read      48
Number of Observations Used      48
Number of Events                  276
Number of Trials                  1193
  
```

Dimensions

```

Columns in X      9
Columns in Z      0
Subjects (Blocks in V) 1
Max Obs per Subject 48
  
```

Optimization Information

```

Optimization Technique   Newton-Raphson
Parameters in Optimization 4
Lower Boundaries        0
Upper Boundaries        0
Fixed Effects           Not Profiled
  
```

Iteration History						
Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient	
0	0	4	117.40174713	.	13.5412	
1	0	3	116.92037584	0.48137129	0.251043	
2	0	3	116.92013648	0.00023935	0.000143	
3	0	3	116.92013648	0.00000000	7.39E-11	

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics	
-2 Log Likelihood	233.84
AIC (smaller is better)	241.84
AICC (smaller is better)	242.77
BIC (smaller is better)	249.33
CAIC (smaller is better)	253.33
HQIC (smaller is better)	244.67
Pearson Chi-Square	87.92
Pearson Chi-Square / DF	2.00

Type III Tests of Fixed Effects				
Effect	Num DF	Den DF	F Value	Pr > F
Age	1	44	15.43	0.0003
Var	1	44	6.08	0.0176
Age*Var	1	44	0.28	0.5990

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
1	M		-1.0383	0.08417	0.2615	0.01625	0.05	-1.2080	-0.8687	0.2301	0.2955	A
2	I		-1.6565	0.1329	0.1602	0.01789	0.05	-1.9244	-1.3886	0.1274	0.1996	B

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
3		950	-1.1533	0.1017	0.2399	0.01854	0.05	-1.3583	-0.9484	0.2045	0.2792	A
4		845	-1.5415	0.1201	0.1763	0.01744	0.05	-1.7834	-1.2995	0.1439	0.2143	B

----- Effect=Age*Var Method=Tukey-Kramer(P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
5	M	950	-0.8026	0.1040	0.3095	0.02222	0.05	-1.0121	-0.5931	0.2666	0.3559	A
6	M	845	-1.2741	0.1324	0.2186	0.02261	0.05	-1.5409	-1.0072	0.1764	0.2675	B
7	I	950	-1.5041	0.1748	0.1818	0.02600	0.05	-1.8564	-1.1518	0.1351	0.2402	B
8	I	845	-1.8089	0.2003	0.1408	0.02423	0.05	-2.2126	-1.4051	0.09863	0.1970	B

The GLIMMIX Procedure
Percent Surviving to Stalk Entry

Model Information

Data Set	WORK.DATA1
Response Variable (Events)	Entry
Response Variable (Trials)	Est
Response Distribution	Binomial
Link Function	Logit
Variance Function	Default
Variance Matrix	Diagonal
Estimation Technique	Maximum Likelihood
Degrees of Freedom Method	Residual

Class Level Information

Class	Levels	Values
Age	2	I M
Var	2	845 950
Rep	12	1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read	48
Number of Observations Used	48
Number of Events	120
Number of Trials	276

Dimensions

Columns in X	9
Columns in Z	0
Subjects (Blocks in V)	1
Max Obs per Subject	48

Optimization Information

Optimization Technique	Newton-Raphson
Parameters in Optimization	4
Lower Boundaries	0
Upper Boundaries	0
Fixed Effects	Not Profiled

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	73.452876663	.	2.368632
1	0	3	73.362663278	0.09021339	0.005842
2	0	3	73.362657543	0.00000574	1.215E-6

Convergence criterion (ABSGCONV=0.00001) satisfied.

Fit Statistics

-2 Log Likelihood	146.73
AIC (smaller is better)	154.73
AICC (smaller is better)	155.66
BIC (smaller is better)	162.21
CAIC (smaller is better)	166.21
HQIC (smaller is better)	157.55
Pearson Chi-Square	61.92
Pearson Chi-Square / DF	1.41

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
Age	1	44	19.47	<.0001
Var	1	44	0.01	0.9318
Age*Var	1	44	3.65	0.0626

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	UpperLetter
1	I		0.6853	0.2631	0.6649	0.05861	0.05	0.1551	1.2154	0.5387	0.7713	A
2	M		-0.6683	0.1579	0.3389	0.03537	0.05	-0.9865	-0.3501	0.2716	0.4133	B

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	UpperLetter
3		950	0.02168	0.1837	0.5054	0.04592	0.05	-0.3486	0.3919	0.4137	0.5967	A
4		845	-0.00474	0.2457	0.4988	0.06143	0.05	-0.4999	0.4904	0.3776	0.6202	A

----- Effect=Age*Var Method=Tukey-Kramer(P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	UpperLetter
5	I	845	0.9651	0.4155	0.7241	0.08300	0.05	0.1277	1.8024	0.5319	0.8584	A
6	I	950	0.4055	0.3227	0.6000	0.07746	0.05	-0.2450	1.0559	0.4391	0.7419	AB
7	M	950	-0.3621	0.1756	0.4104	0.04249	0.05	-0.7160	-0.00819	0.3283	0.4980	BC
8	M	845	-0.9746	0.2624	0.2740	0.05220	0.05	-1.5034	-0.4457	0.1819	0.3904	C

The GLIMMIX Procedure
 Percent Surviving to Pupation

Model Information

Data Set WORK.DATA1
 Response Variable (Events) Surv
 Response Variable (Trials) Est
 Response Distribution Binomial
 Link Function Logit
 Variance Function Default
 Variance Matrix Diagonal
 Estimation Technique Maximum Likelihood
 Degrees of Freedom Method Residual

Class Level Information

Class Levels Values
 Age 2 I M

Var 2 845 950
 Rep 12 1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read 48
 Number of Observations Used 48
 Number of Events 23
 Number of Trials 276

Dimensions
 Columns in X 9
 Columns in Z 0
 Subjects (Blocks in V) 1
 Max Obs per Subject 48

Optimization Information
 Optimization Technique Newton-Raphson
 Parameters in Optimization 4
 Lower Boundaries 0
 Upper Boundaries 0
 Fixed Effects Not Profiled

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	31.487305762	.	4.877972
1	0	3	29.465963171	2.02134259	1.198385
2	0	3	28.93929815	0.52666502	0.322946
3	0	3	28.777136788	0.16216136	0.094843
4	0	3	28.7185603	0.05857649	0.034036
5	0	3	28.697037756	0.02152254	0.012518
6	0	3	28.689123509	0.00791425	0.004605
7	0	3	28.686212488	0.00291102	0.001694
8	0	3	28.685141646	0.00107084	0.000623
9	0	3	28.684747714	0.00039393	0.000229
10	0	3	28.684602796	0.00014492	0.000084
11	0	3	28.684549483	0.00005331	0.000031
12	0	3	28.684529871	0.00001961	0.000011
13	0	3	28.684522656	0.00000722	4.199E-6

Convergence criterion (ABSGCONV=0.00001) satisfied.

Fit Statistics

-2 Log Likelihood	57.37
AIC (smaller is better)	65.37
AICC (smaller is better)	66.30
BIC (smaller is better)	72.85
CAIC (smaller is better)	76.85
HQIC (smaller is better)	68.20
Pearson Chi-Square	27.76
Pearson Chi-Square / DF	0.63

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
Age	1	44	0.00	0.9674
Var	1	44	0.00	0.9812
Age*Var	1	44	0.00	0.9805

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	UpperLetter Mean	Group
1	I		-0.7446	0.2606	0.3220	0.05690	0.05	-1.2698	-0.2193	0.2193	0.4454	A
2	M		-10.7807	244.01	0.000021	0.005074	0.05	-502.54	480.98	561E-221	1.0000	A

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	UpperLetter Mean	Group
3		950	-2.8688	0.5307	0.05372	0.02698	0.05	-3.9384	-1.7993	0.01911	0.1419	A
4		845	-8.6565	244.00	0.000174	0.04244	0.05	-500.42	483.10	47E-219	1.0000	A

----- Effect=Age*Var Method=Tukey-Kramer(P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	UpperLetter Mean	Group
5	I	845	-0.6419	0.3907	0.3448	0.08826	0.05	-1.4292	0.1455	0.1932	0.5363	A
6	I	950	-0.8473	0.3450	0.3000	0.07246	0.05	-1.5427	-0.1519	0.1761	0.4621	A
7	M	950	-4.8903	1.0038	0.007463	0.007435	0.05	-6.9133	-2.8674	0.000994	0.05379	B
8	M	845	-16.6711	488.01	5.752E-8	0.000028	0.05	-1000.19	966.85	0	1.0000	AB

The GLIMMIX Procedure
Percent (Hatch) Stalk Entry

Model Information

Data Set WORK.DATA1
 Response Variable (Events) Entry
 Response Variable (Trials) Hatch
 Response Distribution Binomial
 Link Function Logit
 Variance Function Default
 Variance Matrix Diagonal
 Estimation Technique Maximum Likelihood
 Degrees of Freedom Method Residual

Class Level Information

Class	Levels	Values
Age	2	I M
Var	2	845 950
Rep	12	1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read 48
 Number of Observations Used 48
 Number of Events 120
 Number of Trials 1193

Dimensions

Columns in X 9
 Columns in Z 0
 Subjects (Blocks in V) 1
 Max Obs per Subject 48

Optimization Information

Optimization Technique Newton-Raphson
 Parameters in Optimization 4
 Lower Boundaries 0
 Upper Boundaries 0
 Fixed Effects Not Profiled

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	92.609074526	.	20.9784
1	0	3	90.182780032	2.42629449	1.546366
2	0	3	90.163709811	0.01907022	0.014015
3	0	3	90.163707878	0.00000193	1.442E-6

Convergence criterion (ABSGCONV=0.00001) satisfied.

Fit Statistics

-2 Log Likelihood 180.33
 AIC (smaller is better) 188.33
 AICC (smaller is better) 189.26
 BIC (smaller is better) 195.81
 CAIC (smaller is better) 199.81
 HQIC (smaller is better) 191.16
 Pearson Chi-Square 65.47
 Pearson Chi-Square / DF 1.49

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
Age	1	44	0.95	0.3362
Var	1	44	4.68	0.0360
Age*Var	1	44	3.24	0.0788

----- Effect=Age Method=Tukey-Kramer (P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
1	I		-2.1379	0.1580	0.1055	0.01490	0.05	-2.4563	-1.8196	0.07898	0.1395	A
2	M		-2.3406	0.1360	0.08781	0.01090	0.05	-2.6148	-2.0665	0.06819	0.1124	A

----- Effect=Var Method=Tukey-Kramer (P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
3		950	-2.0138	0.1300	0.1178	0.01351	0.05	-2.2758	-1.7518	0.09315	0.1478	A
4		845	-2.4647	0.1629	0.07837	0.01177	0.05	-2.7931	-2.1363	0.05770	0.1056	B

----- Effect=Age*Var Method=Tukey-Kramer (P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
5	M	950	-1.9276	0.1443	0.1270	0.01600	0.05	-2.2184	-1.6367	0.09811	0.1629	A

6	I	950	-2.1001	0.2163	0.1091	0.02102	0.05	-2.5359	-1.6642	0.07338	0.1592	AB
7	I	845	-2.1758	0.2303	0.1019	0.02108	0.05	-2.6399	-1.7118	0.06661	0.1529	AB
8	M	845	-2.7537	0.2306	0.05988	0.01298	0.05	-3.2184	-2.2889	0.03848	0.09205	B

DISPERSAL AND EXPOSURE (ALL LARVAE)

```
dm'output;clear;log;clear';
Title1'GH All Larvae';
data data1;
input Age$ Var$ Rep$ Midrib Stalk Exp Disp AbsDisp Surv;
cards;
```

I 845 1	1	1	1	0	0	1	I 950 3	0	1	4	3	3	0
I 845 2	1	1	1	0	0	0	I 950 5	0	1	4	1	1	0
I 845 4	1	1	1	0	0	1	I 950 6	0	1	4	1	1	0
I 845 5	1	1	1	0	0	0	I 950 10	0	1	4	3	3	1
I 950 1	1	1	1	0	0	0	I 950 11	0	1	4	4	4	0
I 950 1	1	1	1	1	1	0	I 845 10	0	1	5	5	5	0
I 950 3	1	1	1	0	0	1	I 950 5	0	1	5	3	3	1
I 950 5	1	1	1	0	0	1	M 950 1	0	1	5	-1	1	0
I 950 6	1	1	1	0	0	1	M 950 4	0	1	5	0	0	0
I 950 7	1	1	1	0	0	1	M 950 12	0	1	5	-1	1	0
I 950 8	1	1	1	0	0	0	I 845 10	0	1	6	1	1	1
I 950 9	1	1	1	0	0	1	I 845 1	0	1	6	1	1	0
I 950 9	1	0	1	0	0	0	I 845 9	0	1	6	1	1	1
I 950 10	1	1	1	0	0	0	I 950 2	0	1	6	3	3	1
I 950 11	1	1	1	1	1	1	I 950 8	0	1	6	2	2	0
I 950 12	1	1	1	0	0	0	M 845 5	0	1	6	1	1	0
M 845 2	1	1	1	0	0	0	M 950 1	0	1	6	-3	3	0
M 845 4	1	1	1	0	0	0	M 950 1	0	1	6	-2	2	0
M 845 10	1	1	1	0	0	0	M 950 2	0	1	6	-2	2	0
M 845 10	1	1	1	0	0	0	M 950 2	0	1	6	-1	1	0
M 845 10	1	1	1	0	0	0	M 950 2	0	1	6	0	0	0
M 950 2	1	1	1	0	0	0	M 950 6	0	1	6	0	0	0
M 950 2	1	1	1	0	0	0	M 950 7	0	1	6	-1	1	0
M 950 2	1	1	1	0	0	0	I 845 9	0	1	7	5	5	0
M 950 4	1	1	1	0	0	0	I 845 11	0	1	7	0	0	0
M 950 4	1	1	1	0	0	0	I 845 11	0	1	7	5	5	1
M 950 4	1	1	1	0	0	0	I 845 12	0	1	7	2	2	0
M 950 5	1	1	1	0	0	0	I 845 6	0	1	7	0	0	1
M 950 5	1	1	1	0	0	0	I 845 8	0	1	7	0	0	1
M 950 5	1	1	1	0	0	0	I 845 8	0	1	7	2	2	1
M 950 6	1	1	1	0	0	0	I 950 1	0	1	7	4	4	1
M 950 6	1	1	1	0	0	0	I 950 4	0	1	7	4	4	1
M 950 6	1	1	1	0	0	0	I 950 6	0	1	7	3	3	0
M 950 6	1	1	1	0	0	0	I 950 12	0	1	7	4	4	0
M 950 6	1	1	1	0	0	0	M 845 1	0	1	7	-1	1	0
M 950 7	1	1	1	0	0	0	M 845 7	0	1	7	1	1	0
M 950 7	1	1	1	0	0	0	M 845 9	0	1	7	0	0	0
M 950 10	1	1	1	0	0	0	M 845 11	0	1	7	2	2	0
M 950 11	1	1	1	0	0	0	M 845 12	0	1	7	0	0	0
M 950 12	1	1	1	0	0	0	M 950 1	0	1	7	-4	4	0
M 950 12	1	1	1	0	0	0	M 950 2	0	1	7	-2	2	0
M 950 12	1	1	1	0	0	0	M 950 2	0	1	7	-1	1	0

M 950 4	0	1	7	-2	2	0	I 950 3	1	0	.	0	0	0
M 950 5	0	1	7	2	2	0	I 950 3	1	0	.	0	0	0
M 950 5	0	1	7	3	3	0	I 950 3	1	0	.	0	0	0
M 950 6	0	1	7	1	1	0	I 950 3	1	0	.	0	0	0
M 950 12	0	1	7	-2	2	0	I 950 3	1	0	.	0	0	0
M 950 12	0	1	7	0	0	0	I 950 3	1	0	.	0	0	0
M 950 12	0	1	7	1	1	0	I 950 3	1	0	.	0	0	0
I 845 2	0	1	8	6	6	0	I 950 3	1	0	.	0	0	0
I 845 3	0	1	8	0	0	0	I 950 4	1	0	.	0	0	0
I 845 4	0	1	8	4	4	0	I 950 6	0	0	.	4	4	0
I 845 7	0	1	8	0	0	1	I 950 12	1	0	.	0	0	0
I 845 7	0	1	8	3	3	0	M 845 4	1	0	.	0	0	0
I 950 8	0	1	8	5	5	1	M 845 4	1	0	.	0	0	0
M 845 1	0	1	8	0	0	0	M 845 1	0	0	.	-3	3	0
M 845 3	0	1	8	0	0	0	M 845 1	0	0	.	-2	2	0
M 845 8	0	1	8	0	0	0	M 845 1	0	0	.	1	1	0
M 845 10	0	1	8	0	0	0	M 845 1	0	0	.	2	2	0
M 845 10	0	1	8	1	1	0	M 845 1	0	0	.	2	2	0
M 845 12	0	1	8	-1	1	0	M 845 1	0	0	.	3	3	0
M 950 1	0	1	8	-5	5	0	M 845 2	0	0	.	-1	1	0
M 950 1	0	1	8	-5	5	0	M 845 2	0	0	.	1	1	0
M 950 1	0	1	8	-3	3	0	M 845 2	0	0	.	4	4	0
M 950 2	0	1	8	1	1	0	M 845 2	1	0	.	0	0	0
M 950 2	0	1	8	1	1	0	M 845 3	0	0	.	-2	2	0
M 950 2	0	1	8	3	3	0	M 845 3	0	0	.	1	1	0
M 950 4	0	1	8	3	3	0	M 845 4	0	0	.	-1	1	0
M 950 5	0	1	8	4	4	0	M 845 4	0	0	.	2	2	0
M 950 8	1	0	8	0	0	0	M 845 4	0	0	.	3	3	0
M 950 10	0	1	8	1	1	0	M 845 4	1	0	.	0	0	0
M 950 12	0	1	8	-2	2	0	M 845 5	0	0	.	1	1	0
M 950 12	0	1	8	1	1	0	M 845 5	0	0	.	2	2	0
I 845 11	0	1	9	1	1	1	M 845 5	1	0	.	0	0	0
M 845 1	0	1	9	-2	2	0	M 845 5	1	0	.	0	0	0
M 845 2	0	1	9	0	0	0	M 845 5	1	0	.	0	0	0
M 845 5	0	1	9	5	5	0	M 845 6	0	0	.	-1	1	0
M 845 7	0	1	9	3	3	0	M 845 6	0	0	.	0	0	0
M 950 1	0	1	9	-3	3	0	M 845 6	0	0	.	2	2	0
M 950 2	0	1	9	-2	2	0	M 845 6	1	0	.	0	0	0
M 950 2	0	1	9	3	3	0	M 845 7	0	0	.	-2	2	0
M 950 4	0	1	9	-2	2	0	M 845 7	0	0	.	-1	1	0
M 950 5	0	1	9	2	2	0	M 845 7	0	0	.	0	0	0
M 950 6	0	1	9	-3	3	0	M 845 7	1	0	.	0	0	0
M 950 3	0	1	10	2	2	0	M 845 7	1	0	.	0	0	0
M 950 4	0	1	10	3	3	0	M 845 7	1	0	.	0	0	0
M 950 6	0	1	10	3	3	1	M 845 8	0	0	.	-1	1	0
M 845 4	0	1	11	5	5	0	M 845 8	0	0	.	1	1	0
M 950 8	0	1	11	3	3	0	M 845 8	1	1	.	0	0	0
I 845 10	0	0	.	3	3	0	M 845 9	0	0	.	2	2	0
I 845 12	1	0	.	0	0	0	M 845 9	0	0	.	3	3	0
I 845 1	0	0	.	2	2	0	M 845 10	0	0	.	2	2	0
I 845 1	0	0	.	3	3	0	M 845 10	0	0	.	5	5	0
I 845 1	0	0	.	5	5	0	M 845 10	1	0	.	0	0	0
I 845 2	0	0	.	2	2	0	M 845 10	1	0	.	0	0	0
I 845 4	1	0	.	0	0	0	M 845 10	1	0	.	0	0	0
I 845 5	1	0	.	0	0	0	M 845 11	1	0	.	0	0	0
I 950 2	1	0	.	0	0	0	M 845 11	0	0	.	0	0	0
I 950 2	1	0	.	0	0	0	M 845 11	0	0	.	3	3	0
I 950 3	1	0	.	0	0	0	M 845 12	0	0	.	-2	2	0
I 950 3	1	0	.	0	0	0	M 845 12	0	0	.	1	1	0

M 845 12	1	0	.	0	0	0	M 950 5	1	0	.	0	0	0
M 845 12	1	0	.	0	0	0	M 950 5	1	0	.	0	0	0
M 845 12	1	0	.	0	0	0	M 950 6	0	0	.	-2	20	
M 950 1	0	0	.	0	0	0	M 950 6	0	0	.	2	2	0
M 950 1	0	0	.	2	2	0	M 950 6	1	0	.	0	0	0
M 950 1	0	0	.	3	3	0	M 950 6	1	0	.	0	0	0
M 950 1	1	0	.	0	0	0	M 950 6	1	0	.	0	0	0
M 950 1	1	0	.	0	0	0	M 950 7	0	0	.	-2	2	0
M 950 1	1	0	.	0	0	0	M 950 7	0	0	.	0	0	0
M 950 1	1	0	.	0	0	0	M 950 7	0	0	.	2	2	0
M 950 1	1	0	.	0	0	0	M 950 7	0	0	.	5	5	0
M 950 1	1	0	.	0	0	0	M 950 7	1	0	.	0	0	0
M 950 1	1	0	.	0	0	0	M 950 7	1	0	.	0	0	0
M 950 2	0	0	.	-3	3	0	M 950 8	0	0	.	-1	1	0
M 950 2	0	0	.	2	2	0	M 950 8	0	0	.	0	0	0
M 950 2	0	0	.	4	4	0	M 950 8	0	0	.	1	1	0
M 950 2	0	0	.	6	6	0	M 950 8	0	0	.	2	2	0
M 950 2	1	0	.	0	0	0	M 950 8	0	0	.	5	5	0
M 950 2	1	0	.	0	0	0	M 950 8	1	0	.	0	0	0
M 950 2	1	0	.	0	0	0	M 950 9	0	0	.	-1	1	0
M 950 3	0	0	.	-2	2	0	M 950 9	0	0	.	0	0	0
M 950 3	0	0	.	-1	1	0	M 950 9	0	0	.	1	1	0
M 950 3	0	0	.	0	0	0	M 950 9	0	0	.	4	4	0
M 950 3	0	0	.	1	1	0	M 950 9	1	0	.	0	0	0
M 950 3	0	0	.	4	4	0	M 950 9	1	0	.	0	0	0
M 950 3	1	0	.	0	0	0	M 950 9	1	0	.	0	0	0
M 950 3	1	0	.	0	0	0	M 950 9	1	0	.	0	0	0
M 950 3	1	0	.	0	0	0	M 950 9	1	0	.	0	0	0
M 950 3	1	0	.	0	0	0	M 950 10	0	0	.	-2	2	0
M 950 4	0	0	.	-1	1	0	M 950 10	1	0	.	0	0	0
M 950 4	0	0	.	1	1	0	M 950 10	1	0	.	0	0	0
M 950 4	0	0	.	2	2	0	M 950 10	1	0	.	0	0	0
M 950 4	1	0	.	0	0	0	M 950 11	0	0	.	-2	2	0
M 950 4	1	0	.	0	0	0	M 950 11	0	0	.	1	1	0
M 950 4	1	0	.	0	0	0	M 950 11	1	0	.	0	0	0
M 950 5	0	0	.	1	1	0	M 950 11	1	0	.	0	0	0
M 950 5	0	0	.	6	6	0	M 950 12	0	0	.	3	3	0
M 950 5	0	0	.	7	7	0							

```

;
proc glimmix data=data1 ;
class Age Var Rep ;
model AbsDisp = Age|Var / htype=3 ddfm=kr dist=Gaussian ;
random Rep(Age*Var) ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;
proc glimmix data=data1 ;
class Age Var Rep ;
model Exp = Age|Var / htype=3 ddfm=kr dist=Gaussian ;
random Rep(Age*Var) ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
;%include 'E:\Stats\pdmix800.sas';

```

```
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;
```

The GLIMMIX Procedure

Dispersal

Model Information

Data Set	WORK.DATA1
Response Variable	AbsDisp
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Age	2	I M
Var	2	845 950
Rep	12	1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read	277
Number of Observations Used	277

Dimensions

G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	9
Columns in Z	48
Subjects (Blocks in V)	1
Max Obs per Subject	277

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	1037.610061	.	3.612149
1	0	5	1037.6071724	0.00288868	0.054904
2	0	2	1037.6071717	0.00000064	0.001855

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics

-2 Res Log Likelihood	1037.61
AIC (smaller is better)	1041.61
AICC (smaller is better)	1041.65
BIC (smaller is better)	1045.35
CAIC (smaller is better)	1047.35

HQIC (smaller is better) 1043.02
 Generalized Chi-Square 670.82
 Gener. Chi-Square / DF 2.46

Covariance Parameter Estimates

Cov Parm	Estimate	Standard Error
Rep(Age*Var)	0.01189	0.07124
Residual	2.4572	0.2194

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
Age	1	88.76	1.81	0.1815
Var	1	88.76	0.99	0.3214
Age*Var	1	88.76	2.65	0.1073

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
1	I		1.4584	0.1942	1.4584	0.1942	0.05	1.0744	1.8424	1.0744	1.8424	A
2	M		1.1531	0.1172	1.1531	0.1172	0.05	0.9151	1.3910	0.9151	1.3910	A

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
3		845	1.4188	0.1740	1.4188	0.1740	0.05	1.0746	1.7631	1.0746	1.7631	A
4		950	1.1927	0.1454	1.1927	0.1454	0.05	0.9011	1.4843	0.9011	1.4843	A

----- Effect=Age*Var Method=Tukey-Kramer(P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
5	I	845	1.7560	0.2934	1.7560	0.2934	0.05	1.1773	2.3347	1.1773	2.3347	A
6	M	950	1.2245	0.1409	1.2245	0.1409	0.05	0.9282	1.5208	0.9282	1.5208	A
7	I	950	1.1609	0.2544	1.1609	0.2544	0.05	0.6551	1.6667	0.6551	1.6667	A
8	M	845	1.0817	0.1872	1.0817	0.1872	0.05	0.7066	1.4568	0.7066	1.4568	A

The GLIMMIX Procedure

Duration of Exposure

Model Information

Data Set	WORK.DATA1
Response Variable	Exp
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Age	2	I M
Var	2	845 950
Rep	12	1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read	277
Number of Observations Used	131

Dimensions	
G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	9
Columns in Z	48
Subjects (Blocks in V)	1
Max Obs per Subject	131

Optimization Information	
Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

Iteration History					
Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	658.83161216	.	0

Convergence criterion (ABSGCONV=0.00001) satisfied.

Estimated G matrix is not positive definite.

Fit Statistics	
-2 Res Log Likelihood	658.83
AIC (smaller is better)	660.83
AICC (smaller is better)	660.86
BIC (smaller is better)	662.70
CAIC (smaller is better)	663.70
HQIC (smaller is better)	661.54
Generalized Chi-Square	1197.13
Gener. Chi-Square / DF	9.43

Covariance Parameter Estimates		
Cov Parm	Estimate	Standard Error
Rep(Age*Var)	0	.
Residual	9.4262	1.1829

Type III Tests of Fixed Effects				
Effect	Num DF	Den DF	F Value	Pr > F
Age	1	127	4.23	0.0417
Var	1	127	9.13	0.0030
Age*Var	1	127	1.65	0.2006

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	UpperLetter Mean	Group
1	M		5.8950	0.3802	5.8950	0.3802	0.05	5.1428	6.6473	5.1428	6.6473	A
2	I		4.6762	0.4544	4.6762	0.4544	0.05	3.7770	5.5754	3.7770	5.5754	B

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	UpperLetter Mean	Group
3		845	6.1807	0.4683	6.1807	0.4683	0.05	5.2540	7.1075	5.2540	7.1075	A
4		950	4.3905	0.3629	4.3905	0.3629	0.05	3.6724	5.1085	3.6724	5.1085	B

----- Effect=Age*Var Method=Tukey-Kramer(P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	UpperLetter Mean	Group
5	M	845	6.4091	0.6546	6.4091	0.6546	0.05	5.1138	7.7044	5.1138	7.7044	A
6	I	845	5.9524	0.6700	5.9524	0.6700	0.05	4.6266	7.2781	4.6266	7.2781	A
7	M	950	5.3810	0.3868	5.3810	0.3868	0.05	4.6155	6.1464	4.6155	6.1464	A
8	I	950	3.4000	0.6140	3.4000	0.6140	0.05	2.1849	4.6151	2.1849	4.6151	B

EXPOSURE: SHEATH LARVAE

```
dm'output;clear;log;clear';
Title1'GH Sheath Larvae';
data data1;
input Age$ Var$ Rep$ Stalk Exp ;
cards;
```

I	845	10	1	5	I	845	6	1	7
I	845	10	1	6	I	845	8	1	7
I	845	9	1	7	I	845	8	1	7
I	845	11	1	7	I	845	2	1	8
I	845	11	1	7	I	845	3	1	8
I	845	12	1	7	I	845	4	1	8
I	845	11	1	9	I	845	7	1	8
I	950	3	1	4	I	845	7	1	8
I	950	5	1	4	M	845	5	1	6
I	950	6	1	4	M	845	1	1	7
I	950	10	1	4	M	845	7	1	7
I	950	11	1	4	M	845	9	1	7
I	950	5	1	5	M	845	11	1	7
I	950	2	1	6	M	845	12	1	7
I	950	8	1	6	M	845	1	1	8
I	950	1	1	7	M	845	3	1	8
I	950	4	1	7	M	845	8	1	8
I	950	6	1	7	M	845	10	1	8
I	950	12	1	7	M	845	10	1	8
I	950	8	1	8	M	845	12	1	8
I	845	1	1	6	M	845	1	1	9
I	845	9	1	6	M	845	2	1	9

M	845	5	1	9
M	845	7	1	9
M	845	4	1	11
M	950	1	1	5
M	950	4	1	5
M	950	12	1	5
M	950	1	1	6
M	950	1	1	6
M	950	2	1	6
M	950	2	1	6
M	950	2	1	6
M	950	6	1	6
M	950	7	1	6
M	950	1	1	7
M	950	2	1	7
M	950	2	1	7
M	950	4	1	7
M	950	5	1	7
M	950	5	1	7
M	950	6	1	7
M	950	12	1	7
M	950	12	1	7
M	950	12	1	7
M	950	1	1	8
M	950	1	1	8
M	950	1	1	8
M	950	2	1	8
M	950	2	1	8
M	950	2	1	8
M	950	4	1	8
M	950	5	1	8
M	950	8	1	8
M	950	10	1	8
M	950	12	1	8
M	950	12	1	8
M	950	1	1	9
M	950	2	1	9
M	950	2	1	9
M	950	4	1	9
M	950	5	1	9
M	950	6	1	9
M	950	3	1	10
M	950	4	1	10
M	950	6	1	10
M	950	8	1	11

```

;
proc glimmix data=data1 ;
class Age Var Rep ;
model Exp = Age|Var / htype=3 ddfm=kr dist=Gaussian ;
random Rep(Age*Var) ;
lsmeans Age|Var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

The GLIMMIX Procedure

Model Information

Data Set	WORK.DATA1
Response Variable	Exp
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
Age	2	I M
Var	2	845 950
Rep	12	1 10 11 12 2 3 4 5 6 7 8 9

Number of Observations Read	89
Number of Observations Used	89

Dimensions

G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	9
Columns in Z	42
Subjects (Blocks in V)	1
Max Obs per Subject	89

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

The GLIMMIX Procedure

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	300.97128996	.	0

Convergence criterion (ABSGCONV=0.00001) satisfied.

Estimated G matrix is not positive definite.

Fit Statistics	
-2 Res Log Likelihood	300.97
AIC (smaller is better)	302.97
AICC (smaller is better)	303.02
BIC (smaller is better)	304.71
CAIC (smaller is better)	305.71
HQIC (smaller is better)	303.61
Generalized Chi-Square	149.13
Gener. Chi-Square / DF	1.75

Covariance Parameter Estimates		
Cov Parm	Estimate	Standard Error
Rep(Age*Var)	0	.
Residual	1.7544	0.2691

Type III Tests of Fixed Effects				
Effect	Num DF	Den DF	F Value	Pr > F
Age	1	85	21.03	<.0001
Var	1	85	9.73	0.0025
Age*Var	1	85	3.01	0.0864

----- Effect=Age Method=Tukey-Kramer(P<.05) Set=1 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
1	M		7.7857	0.1904	7.7857	0.1904	0.05	7.4072	8.1642	7.4072	8.1642	A
2	I		6.3665	0.2440	6.3665	0.2440	0.05	5.8814	6.8517	5.8814	6.8517	B

----- Effect=Var Method=Tukey-Kramer(P<.05) Set=2 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
3		845	7.5588	0.2272	7.5588	0.2272	0.05	7.1072	8.0105	7.1072	8.0105	A
4		950	6.5934	0.2102	6.5934	0.2102	0.05	6.1755	7.0113	6.1755	7.0113	B

----- Effect=Age*Var Method=Tukey-Kramer(P<.05) Set=3 -----

Obs	Age	Var	Estimate	Standard Error	Mean	Standard Error of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter
5	M	845	8.0000	0.3213	8.0000	0.3213	0.05	7.3613	8.6387	7.3613	8.6387	A
6	M	950	7.5714	0.2044	7.5714	0.2044	0.05	7.1651	7.9778	7.1651	7.9778	A
7	I	845	7.1176	0.3213	7.1176	0.3213	0.05	6.4789	7.7564	6.4789	7.7564	A
8	I	950	5.6154	0.3674	5.6154	0.3674	0.05	4.8850	6.3458	4.8850	6.3458	B

APENDIX D

VARIETAL RESISTANCE TEST STATISTICAL ANALYSIS

PERCENT BORED INTERNODES

```

dm'output;clear;log;clear';
Title1'Variety Test by Plot';
data data1;
input VAR$ rep$ tot bored Emerg;
cards;
US9315      1      102    0      0
US9315      5       74    1      0
US9315      3      112    0      0
US9315      4      125    5      0
L0768 1     149     28    4
L0768 2     165     1     0
L0768 3     143     0     0
L0768 4     146     2     0
L0768 5     119     3     2
N21 1       118     0     0
N21 2       121     0     0
N21 3       130     3     0
N21 4       124     4     0
N21 5       117     0     0
HO07617    1      130    14     1
HO07617    2      102     6     1
HO07617    3      123     6     1
HO07617    4      125     1     0
HO07617    5      124     0     0
US089001   1      124    12     0
US089001   2      126     6     0
US089001   3      112    17     2
US089001   4      127     1     0
US089001   5      121     0     0
HO07613    1      124    27     1
HO07613    2      135     5     0
HO07613    3      113     4     0
HO07613    4      131     2     0
HO07613    5      108     0     0
HoCP05902  1       96    15     0
HoCP05902  2      109    12     6
HoCP05902  3       98    40    10
HoCP05902  4      109     6     0
HoCP05902  5      108     7     0
L03371     1      126    28     0
L03371     2       98     5     1
L03371     3      125     5     1
L03371     4      128    15     1
L03371     5      115    10     4
N17 1       135    18     2
N17 2       130     0     0
N17 3       131    14     2
N17 4       124     4     0
N17 5       124     4     0
HoCP96540  1      120    17     0
HoCP96540  2      126    17     2
HoCP96540  3      142    11     1
HoCP96540  4      122    11     1
HoCP96540  5      133     0     0
L01299     1      107     6     0
L01299     2      114     0     0
L01299     3       98     5     2
L01299     4       87     2     0
L01299     5       75     0     0
Ho07612    1      103    21     0
Ho07612    2      107    17     5
Ho07612    3      131     9     1
Ho07612    4      116    12     3
Ho07612    5      109     3     0
HoCP00950  1      113    20     0
HoCP00950  2      115     2     0
HoCP00950  3      100     3     0
HoCP00950  4      129     2     1
HoCP00950  5      115     2     1
L0757 1     132    13     0
L0757 2     135    16     8
L0757 3     135     3     0
L0757 5     119    15     5
Ho069610   1      116    21     1
Ho069610   2      122     7     0
Ho069610   3      107     2     0
Ho069610   4      109     2     1
Ho069610   5      113     0     0
HoCP04838  1      153    37     0
HoCP04838  2      127     7     0
HoCP04838  3      132    26     7
HoCP04838  4      152    10     2
HoCP04838  5      127     5     1
Ho07604    1      128    15     0
Ho07604    2      120    16     2
Ho07604    3      116     6     0
Ho07604    4      111     1     0
Ho07604    5      118     5     0
N27 1       133    11     0

```

N27	2	120	5	4	US089003	2	157	6	0
N27	3	143	6	2	US089003	3	143	7	2
N27	4	130	9	0	US089003	4	144	1	0
N27	5	101	11	0	US089003	5	131	1	0
HoCP85845	1	117	0	0	N24	1	91	6	0
HoCP85845	2	123	1	0	N24	2	88	0	0
HoCP85845	3	131	0	0	N24	3	87	3	0
HoCP85845	4	117	3	0	N24	4	83	3	0
HoCP85845	5	114	3	0	N24	5	71	0	0
Ho06563	1	153	74	0	Ho06537	1	130	9	0
Ho06563	2	139	48	10	Ho06537	2	113	8	5
Ho06563	3	131	17	2	Ho06537	3	131	13	3
Ho06563	4	146	0	0	Ho06537	4	132	9	1
Ho06563	5	146	17	7	Ho06537	5	120	2	0
HOCP05961	1	143	14	1	US0140	1	127	12	2
HOCP05961	2	135	9	0	US0140	2	134	11	1
HOCP05961	3	121	5	1	US0140	3	128	14	0
HOCP05961	4	120	3	1	US0140	4	117	2	0
HOCP05961	5	118	8	3	US0140	5	120	3	0
US089003	1	110	5	1					

```

;
ODS HTML FILE='C:\Documents and Settings\treagan\Desktop\Blake Wilson\Variety
Test by plot.html' style = minimal
;
proc glimmix data=data1 ;
class Var rep;
model Bored/Tot = Var / htype=3 ddfm=kr dist=binomial ;
random Rep ;
lsmeans var / ilink diff cl adjust=tukey;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.05,sort=yes);
run;

```

The GLIMMIX Procedure
Probability of a bored internode
Model Information

Data Set	WORK.DATA1
Response Variable (Events)	bored
Response Variable (Trials)	tot
Response Distribution	Binomial
Link Function	Logit
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Residual PL
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
VAR	25	HOCP0596 HO07613 HO07617 Ho06537 Ho06563 Ho069610 Ho07604 Ho07612 HoCP0095 HoCP0483

HoCP0590 HoCP8584 HoCP9654 L01299 L03371
 L0757 L0768 N17 N21 N24 N27 US0140 US089001
 US089003 US9315

rep 5 1 2 3 4 5

Number of Observations Read 123
 Number of Observations Used 123
 Number of Events 1057
 Number of Trials 14898

Dimensions

G-side Cov. Parameters 1
 Columns in X 26
 Columns in Z 5
 Subjects (Blocks in V) 1
 Max Obs per Subject 123

Optimization Information

Optimization Technique Dual Quasi-Newton
 Parameters in Optimization 1
 Lower Boundaries 1
 Upper Boundaries 0
 Fixed Effects Profiled
 Starting From Data

Iteration History

Iteration	Restarts	Subiterations	Objective Function	Change	Max Gradient
0	0	4	410.88103698	2.00000000	0.000017
1	0	3	514.45822984	0.49515945	0.000079
2	0	2	541.27249692	0.09674837	2.933E-7
3	0	1	543.41936418	0.00246621	5.978E-9
4	0	0	543.44404146	0.00000084	1.086E-6
5	0	0	543.44404706	0.00000000	1.086E-6

Convergence criterion (PCONV=1.11022E-8) satisfied.

Fit Statistics

-2 Res Log Pseudo-Likelihood 543.44
 Generalized Chi-Square 467.51
 Gener. Chi-Square / DF 4.77

Covariance Parameter

Estimates

Cov Parm	Estimate	Standard Error
rep	0.3903	0.2804

Type III Tests of Fixed Effects

Effect	Num DF	Den DF	F Value	Pr > F
VAR	24	98	17.68	<.0001

----- Effect=VAR Method=Tukey-Kramer(P<.05) Set=1 -----

Letter	Obs VAR	Standard		Standard Error of		Alpha	Lower	Upper	Lower	Upper
		Estimate	Error	Mean	Mean				Mean	Mean
1	Ho06563	-1.3689	0.2947	0.2028	0.04765	0.05	-2.1361	-0.6016	0.1056	0.3540 A
2	HoCP0590	-1.7838	0.3060	0.1438	0.03768	0.05	-2.5465	-1.0210	0.07266	0.2648 AB
3	HoCP0483	-2.1023	0.3037	0.1089	0.02946	0.05	-2.8655	-1.3391	0.05388	0.2077 BC
4	Ho07612	-2.2005	0.3114	0.09970	0.02795	0.05	-2.9631	-1.4379	0.04912	0.1919 BCD
5	L03371	-2.2599	0.3109	0.09450	0.02660	0.05	-3.0225	-1.4974	0.04642	0.1828 BCD
6	HoCP9654	-2.4665	0.3136	0.07824	0.02262	0.05	-3.2293	-1.7037	0.03808	0.1540 BCDE
7	L0757	-2.5778	0.3203	0.07058	0.02101	0.05	-3.3423	-1.8133	0.03415	0.1402 CDEF
8	Ho07604	-2.7068	0.3225	0.06257	0.01892	0.05	-3.4722	-1.9415	0.03011	0.1255 CDEF
9	US0140	-2.7824	0.3231	0.05828	0.01773	0.05	-3.5480	-2.0169	0.02798	0.1174CDEFG
10	N27	-2.8020	0.3232	0.05722	0.01743	0.05	-3.5676	-2.0364	0.02745	0.1154CDEFG
11	Ho06537	-2.8028	0.3240	0.05717	0.01747	0.05	-3.5688	-2.0369	0.02742	0.1154CDEFG
12	H007613	-2.8587	0.3271	0.05424	0.01678	0.05	-3.6261	-2.0912	0.02593	0.1100CDEFG
13	N17	-2.8708	0.3249	0.05361	0.01648	0.05	-3.6372	-2.1045	0.02565	0.1087 DEFG
14	US089001	-2.9075	0.3293	0.05179	0.01617	0.05	-3.6761	-2.1388	0.02470	0.1054 DEFG
15	H0CP0596	-2.9077	0.3260	0.05177	0.01600	0.05	-3.6746	-2.1408	0.02473	0.1052 DEFG
16	Ho069610	-2.9648	0.3348	0.04904	0.01561	0.05	-3.7370	-2.1926	0.02327	0.1004 DEFG
17	HoCP0095	-3.0543	0.3395	0.04503	0.01460	0.05	-3.8300	-2.2786	0.02125	0.09291DEFGH
18	L0768	-3.1744	0.3313	0.04014	0.01276	0.05	-3.9443	-2.4046	0.01900	0.08282 EFGH
19	H007617	-3.2171	0.3432	0.03853	0.01271	0.05	-3.9958	-2.4384	0.01806	0.08029 EFGH
20	US089003	-3.6077	0.3609	0.02640	0.009277	0.05	-4.4040	-2.8114	0.01208	0.05671 FGH
21	N24	-3.7114	0.4062	0.02386	0.009461	0.05	-4.5673	-2.8556	0.01028	0.05440 FGH
22	L01299	-3.7891	0.3978	0.02212	0.008603	0.05	-4.6327	-2.9455	0.009635	0.04995 FGH
23	US9315	-4.3926	0.4988	0.01222	0.006020	0.05	-5.4013	-3.3839	0.004491	0.03280 GH
24	HoCP8584	-4.6005	0.4728	0.009947	0.004656	0.05	-5.5637	-3.6372	0.003820	0.02565 H
25	N21	-4.6073	0.4727	0.009881	0.004625	0.05	-5.5705	-3.6441	0.003794	0.02548 H

EMERGENCE PER STALK

```
dm'output;clear;log;clear';
Title1'Variety Test by Plot';
data data1;
input VAR$ rep$ emer Emerg;
cards;
```

US9315	1	0	0.0001	Ho07617	4	0	0.0001
US9315	5	0	0.0001	Ho07617	5	0	0.0001
US9315	3	0	0.0001	US089001	1	0	0.0001
US9315	4	0	0.0001	US089001	2	0	0.0001
L0768 1	0.4	0.4001		US089001	3	0.2	0.2001
L0768 2	0	0.0001		US089001	4	0	0.0001
L0768 3	0	0.0001		US089001	5	0	0.0001
L0768 4	0	0.0001		HO07613	1	0.1	0.1001
L0768 5	0.2	0.2001		HO07613	2	0	0.0001
N21 1	0	0.0001		HO07613	3	0	0.0001
N21 2	0	0.0001		HO07613	4	0	0.0001
N21 3	0	0.0001		HO07613	5	0	0.0001
N21 4	0	0.0001		HoCP05902	1	0	0.0001
N21 5	0	0.0001		HoCP05902	2	0.6	0.6001
Ho07617	1	0.1	0.1001	HoCP05902	3	1	1.0001
Ho07617	2	0.1	0.1001	HoCP05902	4	0	0.0001
Ho07617	3	0.1	0.1001	HoCP05902	5	0	0.0001

L03371	1	0	0.0001	HoCP85845	1	0	0.0001
L03371	2	0.111111		HoCP85845	2	0	0.0001
	0.111211			HoCP85845	3	0	0.0001
L03371	3	0.1	0.1001	HoCP85845	4	0	0.0001
L03371	4	0.1	0.1001	HoCP85845	5	0	0.0001
L03371	5	0.4	0.4001	Ho06563	1	0	0.0001
N17	1	0.22222222	0.22232222	Ho06563	2	1	1.0001
N17	2	0	0.0001	Ho06563	3	0.2	0.2001
N17	3	0.2	0.2001	Ho06563	4	0	0.0001
N17	4	0	0.0001	Ho06563	5	0.7	0.7001
N17	5	0	0.0001	H0CP05961	1	0.090909091	
HoCP96540	1	0	0.0001		0.091009091		
HoCP96540	2	0.2	0.2001	H0CP05961	2	0	0.0001
HoCP96540	3	0.1	0.1001	H0CP05961	3	0.1	0.1001
HoCP96540	4	0.1	0.1001	H0CP05961	4	0.1	0.1001
HoCP96540	5	0	0.0001	H0CP05961	5	0.3	0.3001
L01299	1	0	0.0001	US089003	1	0.1	0.1001
L01299	2	0	0.0001	US089003	2	0	0.0001
L01299	3	0.2	0.2001	US089003	3	0.2	0.2001
L01299	4	0	0.0001	US089003	4	0	0.0001
L01299	5	0	0.0001	US089003	5	0	0.0001
Ho07612	1	0	0.0001	N24	1	0	0.0001
Ho07612	2	0.5	0.5001	N24	2	0	0.0001
Ho07612	3	0.1	0.1001	N24	3	0	0.0001
Ho07612	4	0.3	0.3001	N24	4	0	0.0001
Ho07612	5	0	0.0001	N24	5	0	0.0001
HoCP00950	1	0	0.0001	Ho06537	1	0	0.0001
HoCP00950	2	0	0.0001	Ho06537	2	0.555555556	
HoCP00950	3	0	0.0001		0.555655556		
HoCP00950	4	0.1	0.1001	Ho06537	3	0.3	0.3001
HoCP00950	5	0.1	0.1001	Ho06537	4	0.1	0.1001
L0757	1	0	0.0001	Ho06537	5	0	0.0001
L0757	2	0.8	0.8001	US0140	1	0.2	0.2001
L0757	3	0	0.0001	US0140	2	0.090909091	
L0757	5	0.5	0.5001		0.091009091		
Ho069610	1	0.1	0.1001	US0140	3	0	0.0001
Ho069610	2	0	0.0001	US0140	4	0	0.0001
Ho069610	3	0	0.0001	US0140	5	0	0.000
Ho069610	4	0.1	0.1001				
Ho069610	5	0	0.0001				
HoCP04838	1	0	0.0001				
HoCP04838	2	0	0.0001				
HoCP04838	3	0.7	0.7001				
HoCP04838	4	0.2	0.2001				
HoCP04838	5	0.1	0.1001				
Ho07604	1	0	0.0001				
Ho07604	2	0.2	0.2001				
Ho07604	3	0	0.0001				
Ho07604	4	0	0.0001				
Ho07604	5	0	0.0001				
N27	1	0	0.0001				
N27	2	0.4	0.4001				
N27	3	0.2	0.2001				
N27	4	0	0.0001				
N27	5	0	0.0001				

```

proc glimmix data=data1 ;
class Var rep;
model Emerg = Var / htype=3 ddfm=kr ;
random Rep ;
lsmeans var / ilink diff cl ;
ods output diffs=ppp lsmeans=mmm;
ods listing exclude diffs lsmeans;
run;
%include 'E:\Stats\pdmix800.sas';
%pdmix800(ppp,mmm,alpha=.1,sort=yes);
run;

```

The GLIMMIX Procedure

Emergence

Model Information

Data Set	WORK.DATA1
Response Variable	Emerg
Response Distribution	Gaussian
Link Function	Identity
Variance Function	Default
Variance Matrix	Not blocked
Estimation Technique	Restricted Maximum Likelihood
Degrees of Freedom Method	Kenward-Roger
Fixed Effects SE Adjustment	Kenward-Roger

Class Level Information

Class	Levels	Values
VAR	25	HOCP0596 HO07613 Ho06537 Ho06563 Ho069610 Ho07604 Ho07612 Ho07617 HoCP0095 HoCP0483 HoCP0590 HoCP8584 HoCP9654 L01299 L03371 L0757 L0768 N17 N21 N24 N27 US0140 US089001 US089003 US9315
rep	5	1 2 3 4 5
		Number of Observations Read 123
		Number of Observations Used 123

Dimensions

G-side Cov. Parameters	1
R-side Cov. Parameters	1
Columns in X	26
Columns in Z	5
Subjects (Blocks in V)	1
Max Obs per Subject	123

Optimization Information

Optimization Technique	Dual Quasi-Newton
Parameters in Optimization	1
Lower Boundaries	1
Upper Boundaries	0
Fixed Effects	Profiled
Residual Variance	Profiled
Starting From	Data

Iteration History

Iteration	Restarts	Evaluations	Objective Function	Change	Max Gradient
0	0	4	-9.073056736	.	0.619005
1	0	5	-9.073545369	0.00048863	0.019953

2	0	2	-9.073545899	0.00000053	0.000614
3	0	2	-9.0735459	0.00000000	6.33E-7

Convergence criterion (GCONV=1E-8) satisfied.

Fit Statistics

-2 Res Log Likelihood	-9.07
AIC (smaller is better)	-5.07
AICC (smaller is better)	-4.95
BIC (smaller is better)	-5.85
CAIC (smaller is better)	-3.85
HQIC (smaller is better)	-7.17
Generalized Chi-Square	3.36
Gener. Chi-Square / DF	0.03

Covariance Parameter Estimates

		Standard
Cov Parm	Estimate	Error
rep	0.002046	0.002458
Residual	0.03428	0.005001

Type III Tests of Fixed Effects

	Num	Den		
Effect	DF	DF	F Value	Pr > F
VAR	24	94.01	1.56	0.0671

----- Effect=VAR Method=LSD(P<.1) Set=1 -----

Obs	VAR	Estimate	Standard Err	Mean	Stand Err of Mean	Alpha	Lower	Upper	Lower Mean	Upper Mean	Letter Group
1	Ho06563	0.3801	0.08523	0.3801	0.08523	0.05	0.2108	0.5494	0.2108	0.5494	A
2	HoCP0590	0.3201	0.08523	0.3201	0.08523	0.05	0.1508	0.4894	0.1508	0.4894	AB
3	L0757	0.3175	0.09514	0.3175	0.09514	0.05	0.1286	0.5063	0.1286	0.5063	ABC
4	HoCP0483	0.2001	0.08523	0.2001	0.08523	0.05	0.03080	0.3694	0.03080	0.3694	ABCD
5	Ho06537	0.1912	0.08523	0.1912	0.08523	0.05	0.02191	0.3605	0.02191	0.3605	ABCDE
6	Ho07612	0.1801	0.08523	0.1801	0.08523	0.05	0.01080	0.3494	0.01080	0.3494	BCDE
7	L03371	0.1423	0.08523	0.1423	0.08523	0.05	-0.02698	0.3116	-0.02698	0.3116	BCDE
8	N27	0.1201	0.08523	0.1201	0.08523	0.05	-0.04920	0.2894	-0.04920	0.2894	CDE
9	L0768	0.1201	0.08523	0.1201	0.08523	0.05	-0.04920	0.2894	-0.04920	0.2894	CDE
10	HoCP0596	0.1183	0.08523	0.1183	0.08523	0.05	-0.05102	0.2876	-0.05102	0.2876	CDE
11	N17	0.08454	0.08523	0.08454	0.08523	0.05	-0.08476	0.2538	-0.08476	0.2538	DE
12	HoCP9654	0.08010	0.08523	0.08010	0.08523	0.05	-0.08920	0.2494	-0.08920	0.2494	DE
13	US089003	0.06010	0.08523	0.06010	0.08523	0.05	-0.1092	0.2294	-0.1092	0.2294	DE
14	Ho07617	0.06010	0.08523	0.06010	0.08523	0.05	-0.1092	0.2294	-0.1092	0.2294	DE
15	US0140	0.05828	0.08523	0.05828	0.08523	0.05	-0.1110	0.2276	-0.1110	0.2276	DE
16	Ho07604	0.04010	0.08523	0.04010	0.08523	0.05	-0.1292	0.2094	-0.1292	0.2094	DE
17	Ho069610	0.04010	0.08523	0.04010	0.08523	0.05	-0.1292	0.2094	-0.1292	0.2094	DE
18	HoCP0095	0.04010	0.08523	0.04010	0.08523	0.05	-0.1292	0.2094	-0.1292	0.2094	DE
19	L01299	0.04010	0.08523	0.04010	0.08523	0.05	-0.1292	0.2094	-0.1292	0.2094	DE
20	US089001	0.04010	0.08523	0.04010	0.08523	0.05	-0.1292	0.2094	-0.1292	0.2094	DE
21	H007613	0.02010	0.08523	0.02010	0.08523	0.05	-0.1492	0.1894	-0.1492	0.1894	DE
22	US9315	0.01179	0.09514	0.01179	0.09514	0.05	-0.1771	0.2007	-0.1771	0.2007	DE
23	N24	0.000100	0.08523	0.000100	0.08523	0.05	-0.1692	0.1694	-0.1692	0.1694	E
24	HoCP8584	0.000100	0.08523	0.000100	0.08523	0.05	-0.1692	0.1694	-0.1692	0.1694	E
25	N21	0.000100	0.08523	0.000100	0.08523	0.05	-0.1692	0.1694	-0.1692	0.1694	E

VITA

Blake Emerson Wilson was born into a loving family in 1987 in Tulsa, Oklahoma. He spent the early years of his life in Austin, Texas, and Shreveport, Louisiana, before landing in Mandeville, Louisiana, at the age of 9. He graduated from Mandeville High School in 2005. He received his Bachelor of Science degree in biology from LSU in 2009. While an undergraduate, Blake held student worker positions in various fields of biology before becoming an assistant in Dr. Reagan's sugarcane entomology lab. It was during his final semester as an undergraduate that he developed an interest in entomology. With help and encouragement from Dr. Reagan, Blake enrolled in a master's program in entomology at Louisiana State University.

Under the guidance of Dr. Reagan and Dr. Showler, Blake conducted his thesis research in the Lower Rio Grande Valley during the summer of 2010. Blake plans to continue to reside in Baton Rouge, Louisiana, and continue to be active in the scientific community. Blake hopes to continue to conduct entomological research and aspires to make meaningful contributions to agriculture in Louisiana.